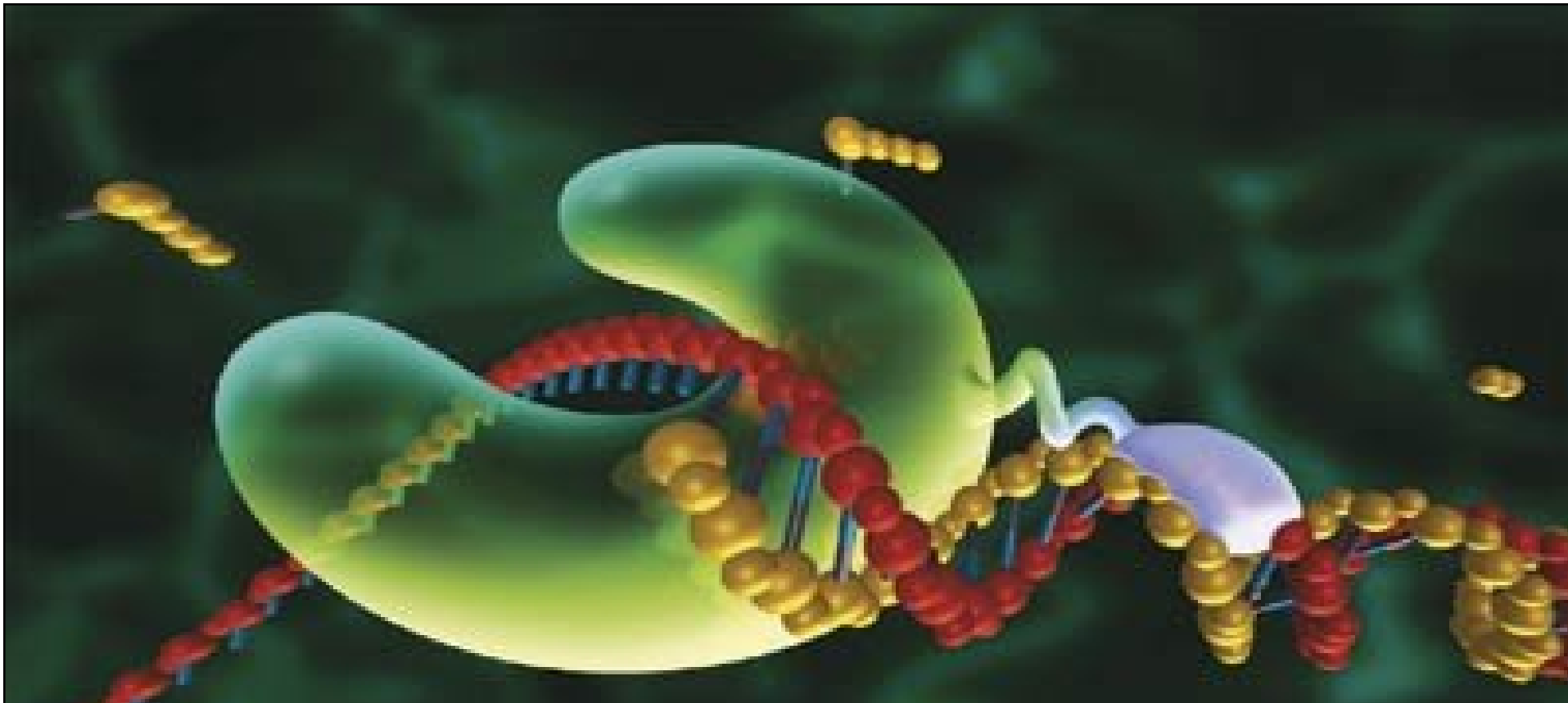




Fundamentals of Real Time PCR

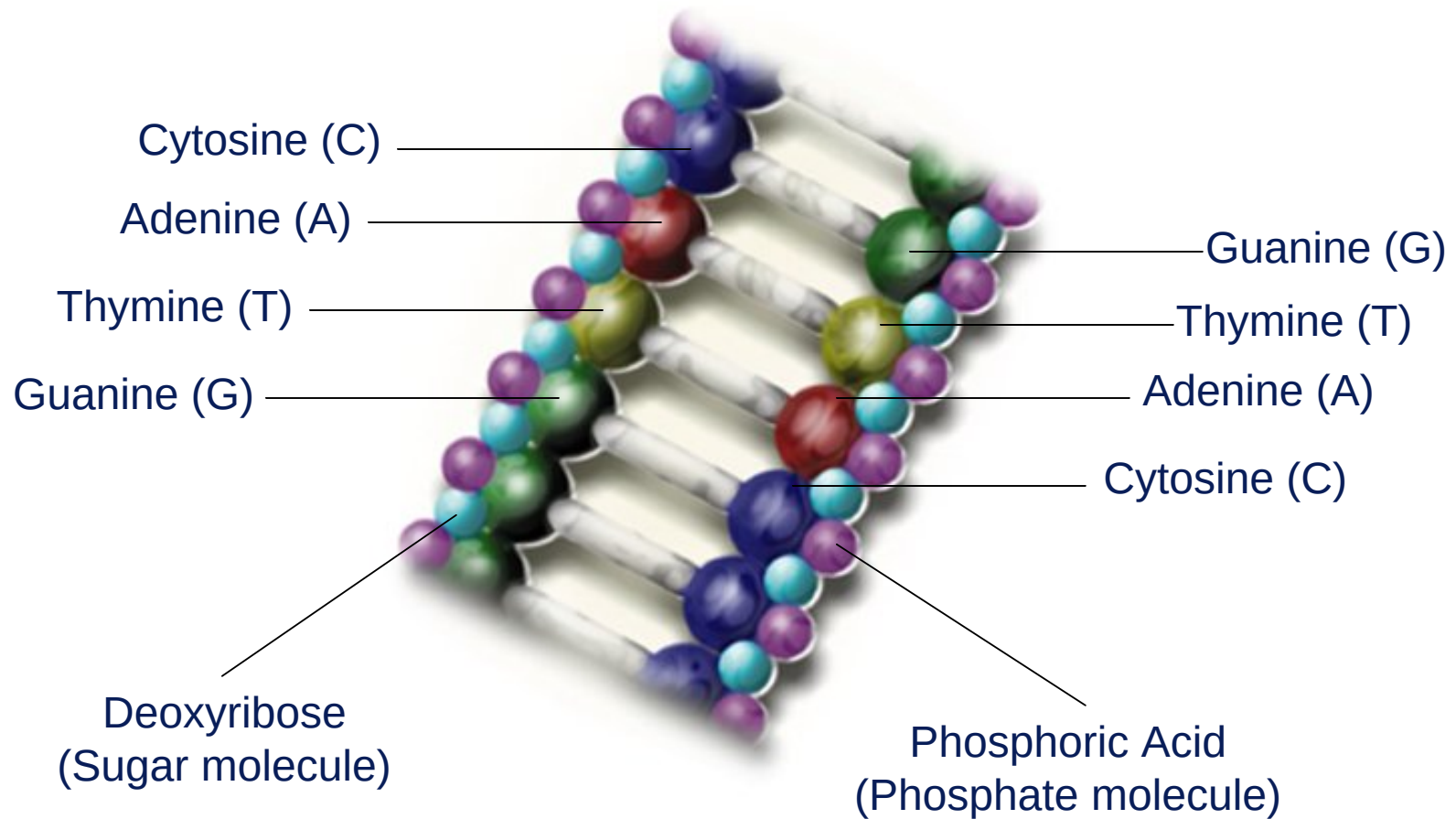
Mohamed Abdel Fattah

*Senior Technical Specialist
Scientific Support - Molecular Biology Dept.
AnalysisAB Co.*

Mobile: 012 27 906 74

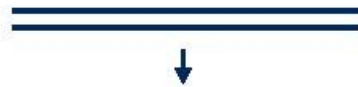
E-mail: analysis@analysis-ab.com

What is that DNA ?

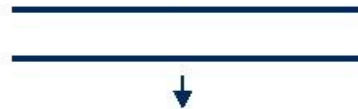


Polymerase Chain Reaction (PCR)

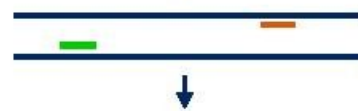
Unamplified DNA



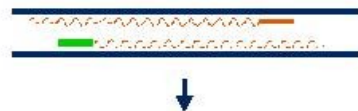
Strand Denaturation



Primer annealing



Primer extension



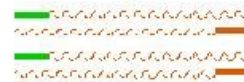
Cycling:
Exponential amplification
of PCR products

Denaturation

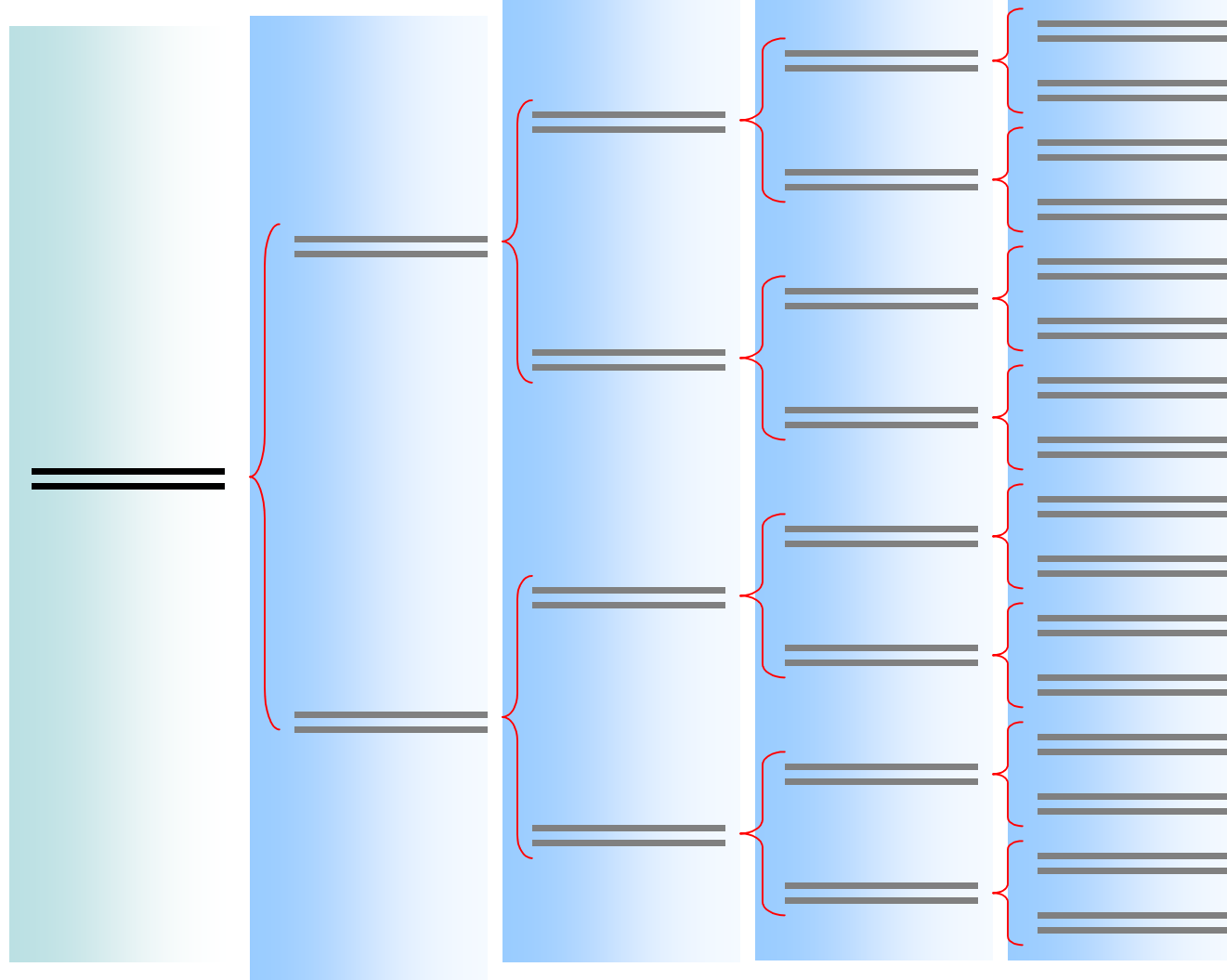


Annealing

Extension



PCR As Exponential Amplification



25 cycles

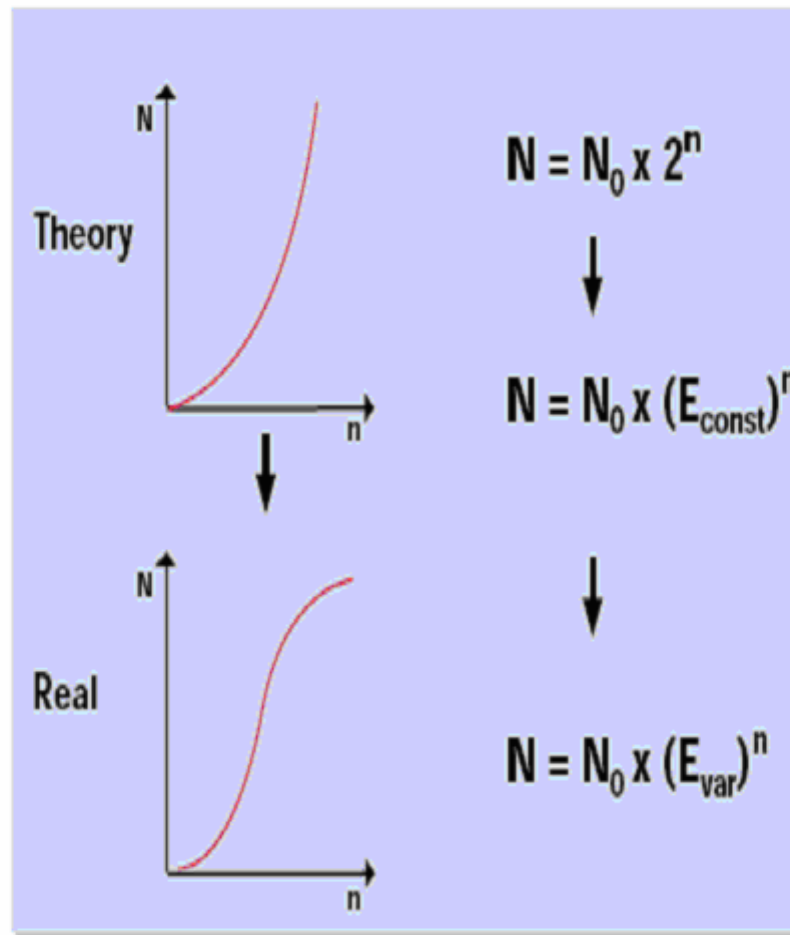
2^{25} copies
(33,554,432)

PCR In Theoretical Aspect

$$N = N_0 \times 2^n$$

N: number of amplified molecules
N₀: initial number of molecules
n: number of amplification cycles

PCR In Graphical Aspect



N : number of amplified molecules; N_0 : initial number of molecules; n : number of amplification cycles; E : amplification efficiency

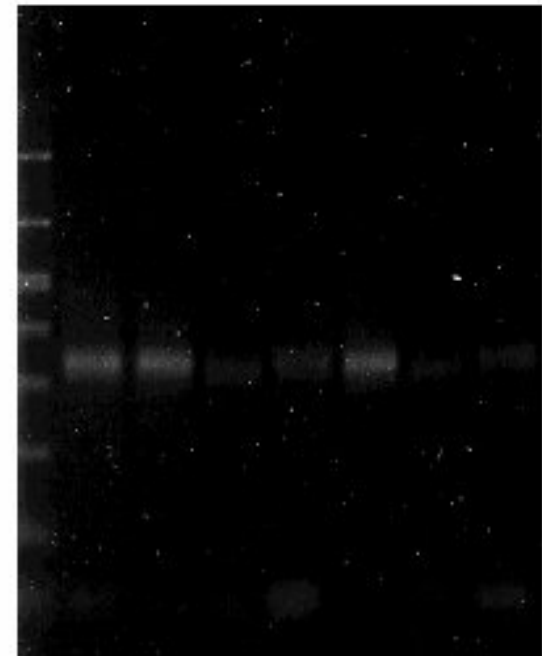
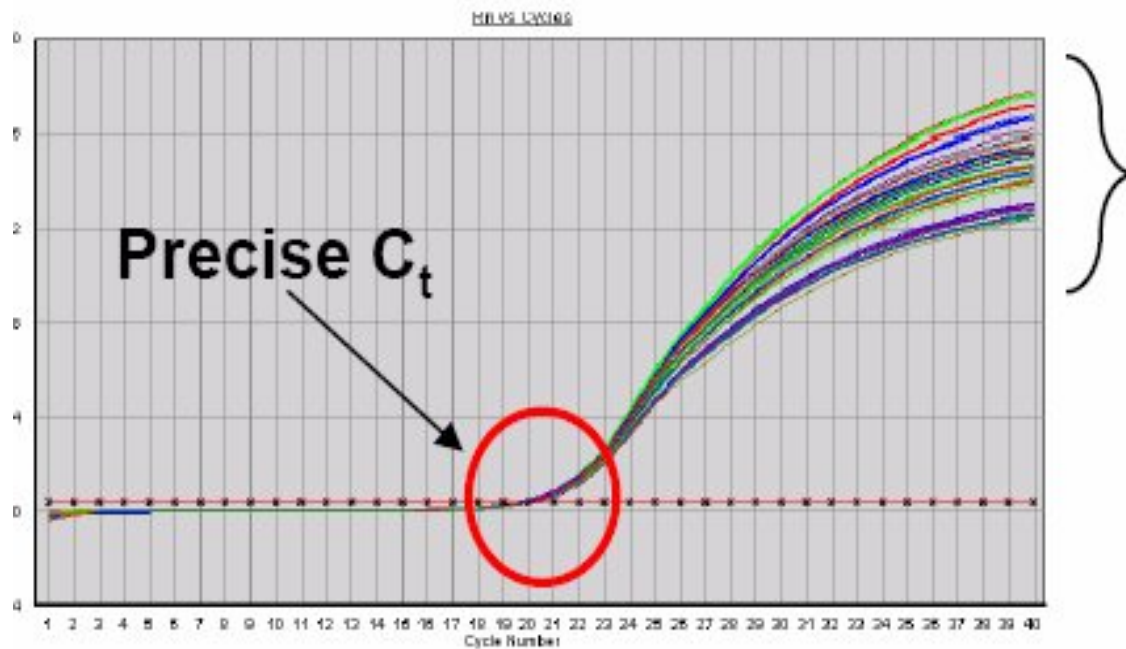
Limitations of PCR Efficiency, Plateau Effect

The Plateau Effect is caused by:

- Enzyme instability.
- Reagents consumption.
- Product inhibition. e.g. pyrophosphate accumulation.
- Competition of re-annealing amplification product with primers.

Limitations of PCR Efficiency

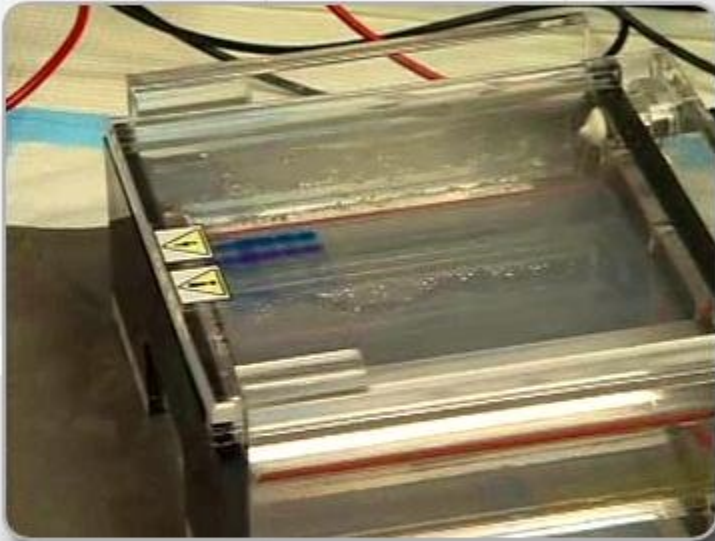
Variable PCR Plateau



Amplification Plot of 96 Sample Replicates

Real-Time PCR System

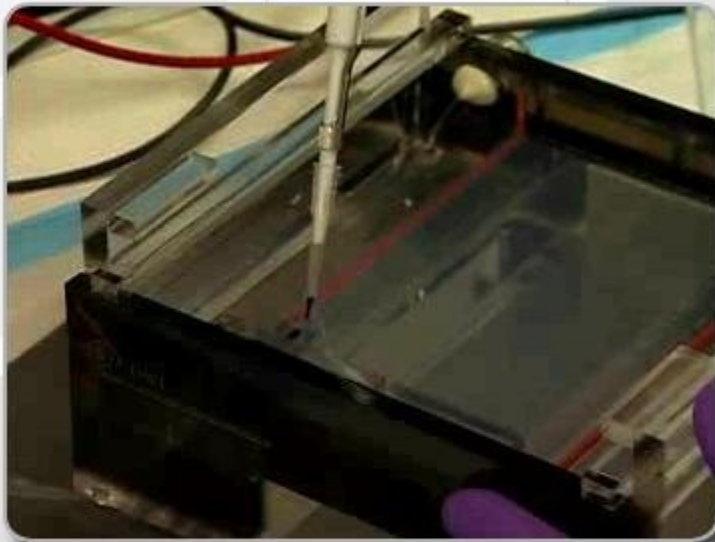
**time
consuming**



messy



Real-Time PCR System



waiting

Real-Time PCR System



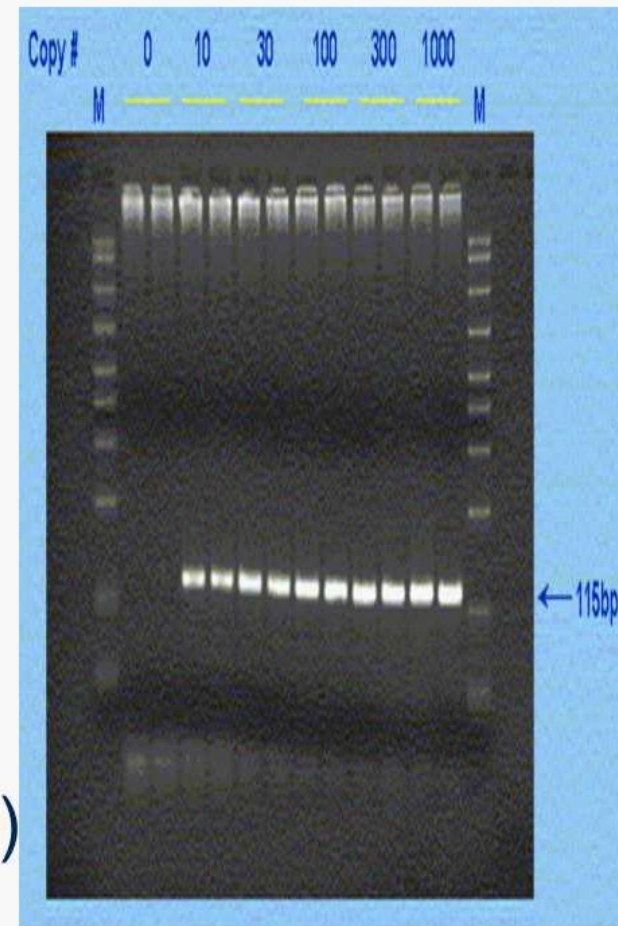
result A

result B

result C

Limitations of Traditional End-Point PCR

- Low sensitivity
- Poor precision
- Results are not expressed as numbers
- Ethidium bromide staining is not quantitative
- Post-PCR processing required
- Narrow dynamic range (<2 logs)



From *PCR* to *Real-Time PCR*

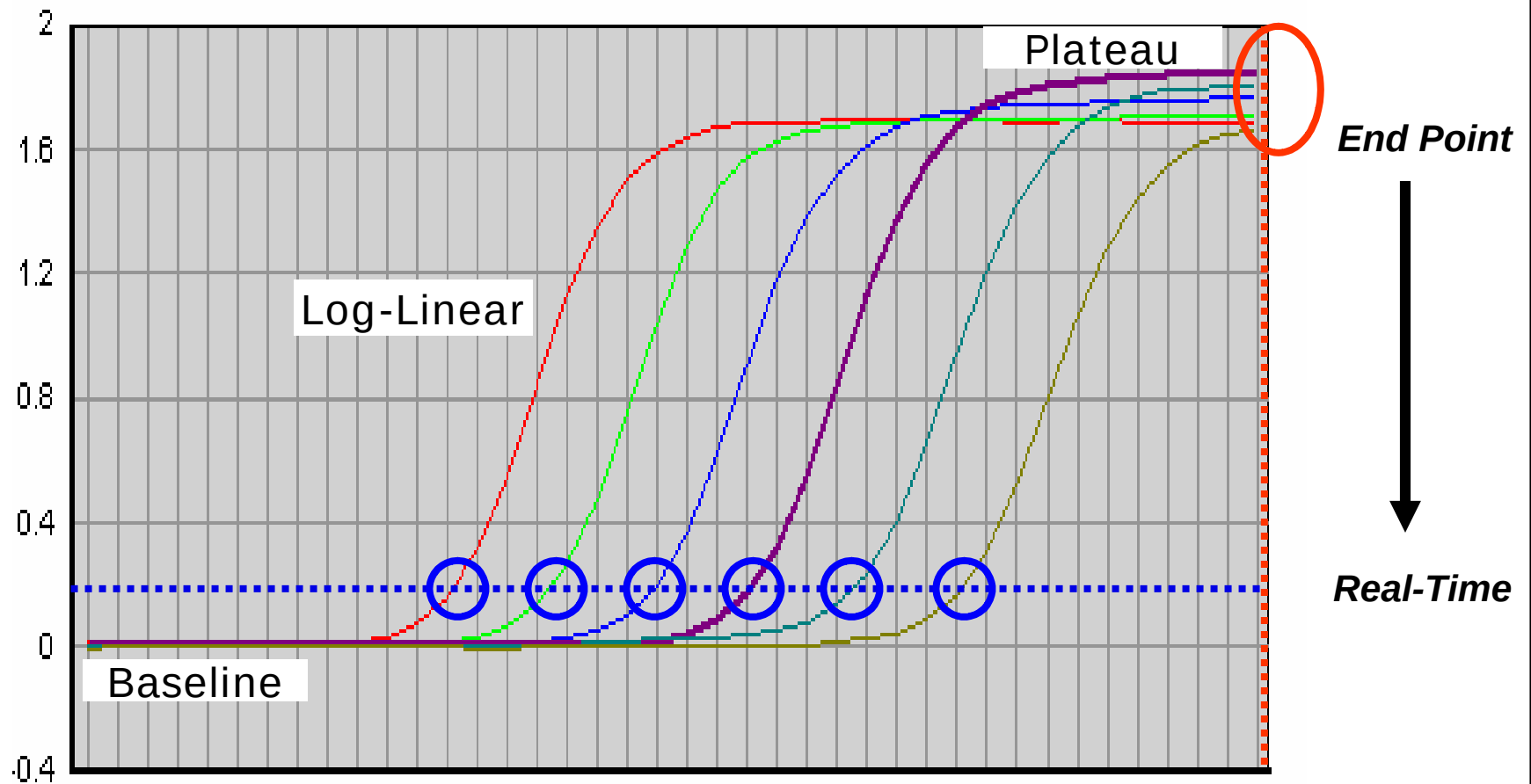
Real-Time PCR turns the question

HOW MUCH?

Into the question

WHEN?

From *PCR* to *Real-Time PCR*



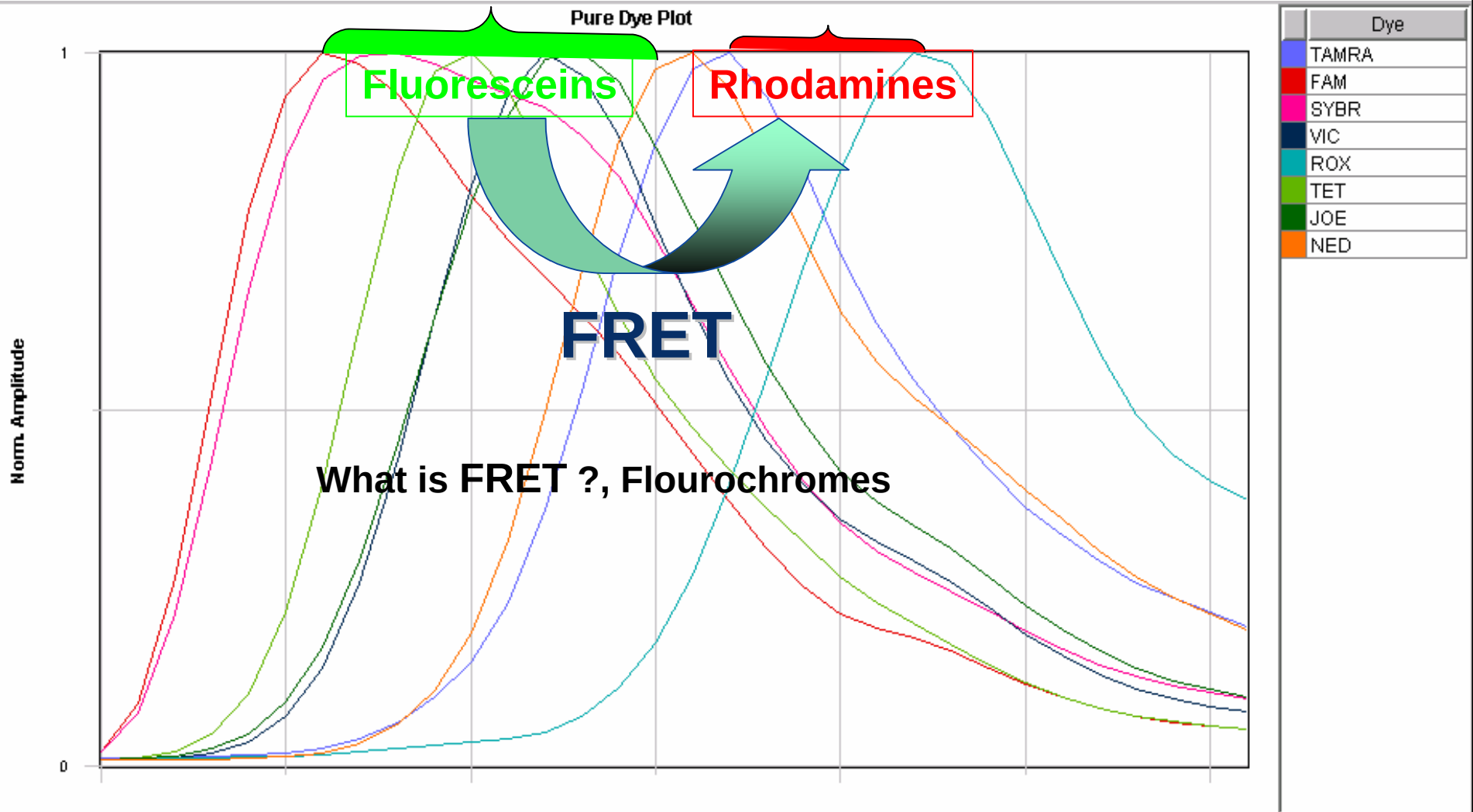
Towards *Real-Time PCR*

1st Point

Detection of Specific PCR Product by
utilizing the 5' to 3' Exonuclease
Activity of *Thermus Aquaticus* DNA
Polymerase

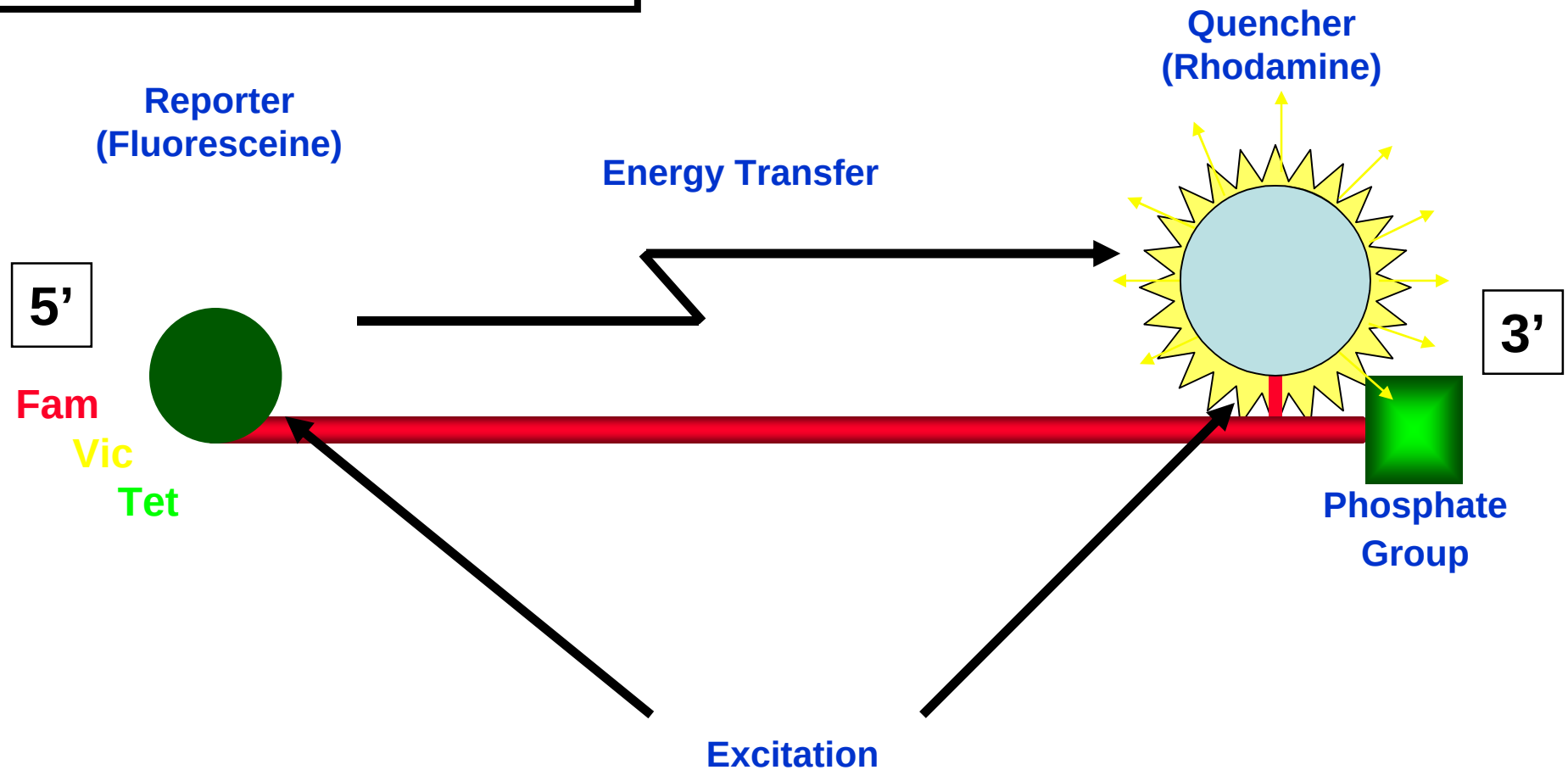
Towards *Real-Time PCR*

2nd Point



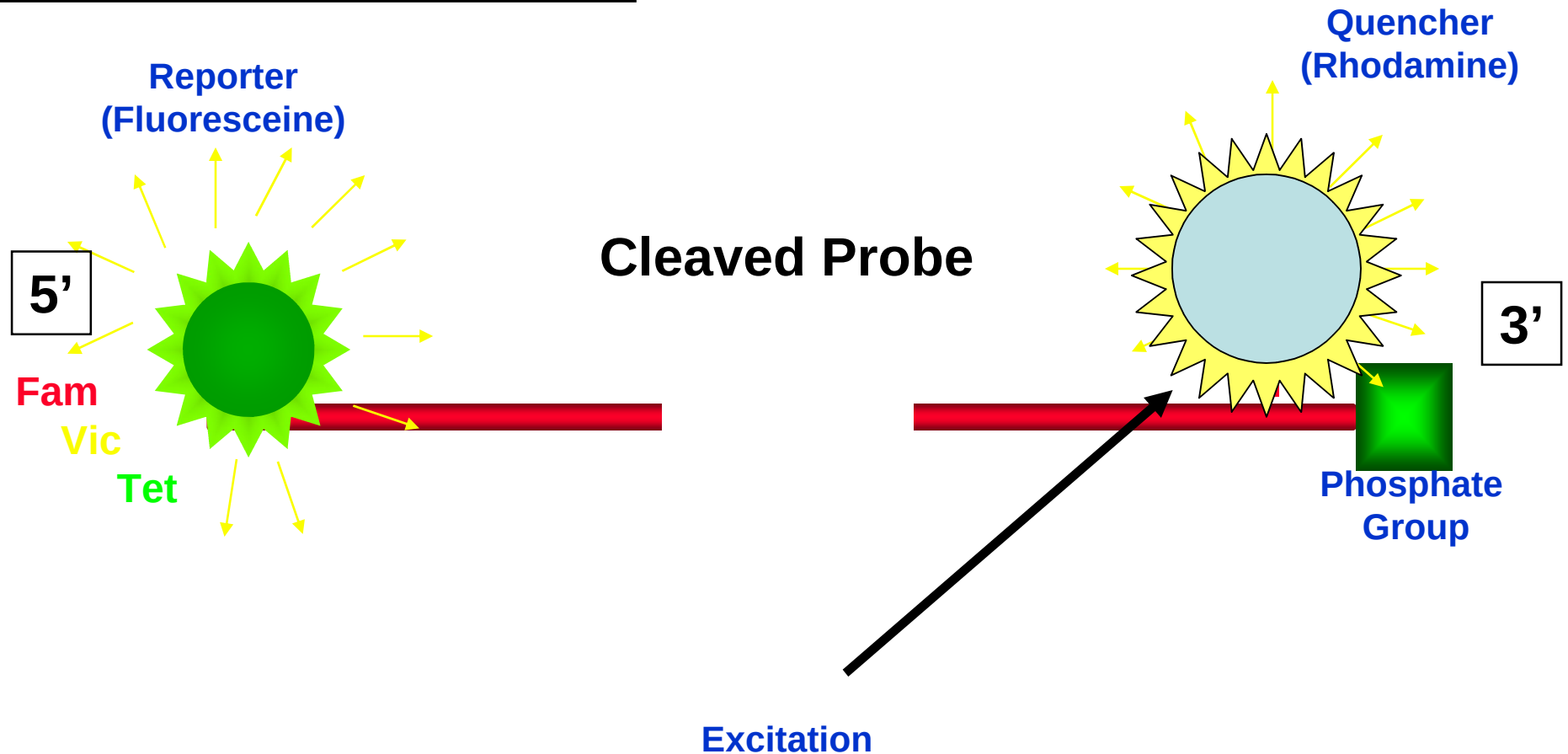
Towards *Real-Time PCR*

What is FRET ?, Flourochromes



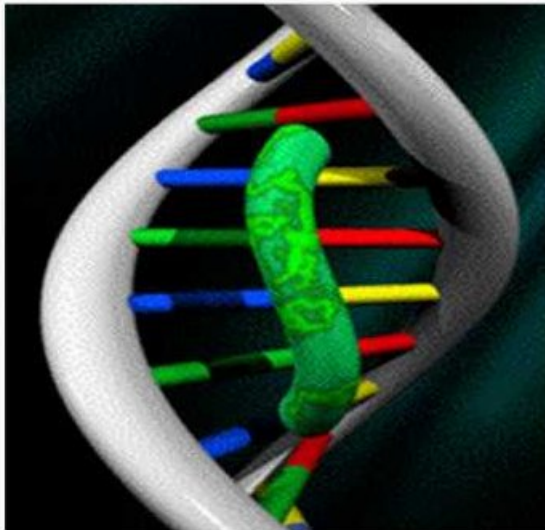
Towards *Real-Time PCR*

What is FRET ?, Flourochromes



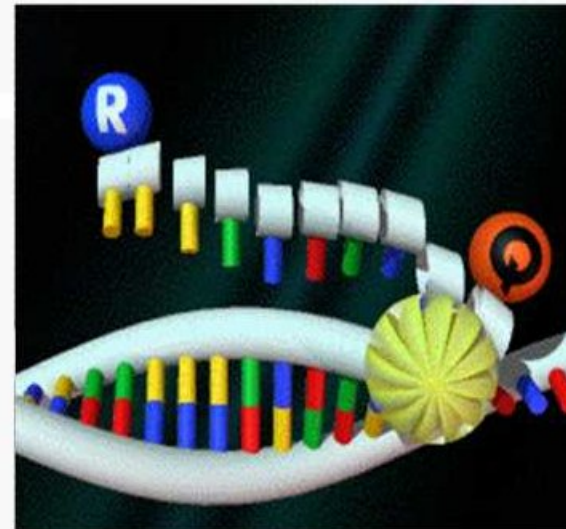
Real-Time PCR Chemistries

SYBR[®] Green I dye



Binds double
stranded DNA

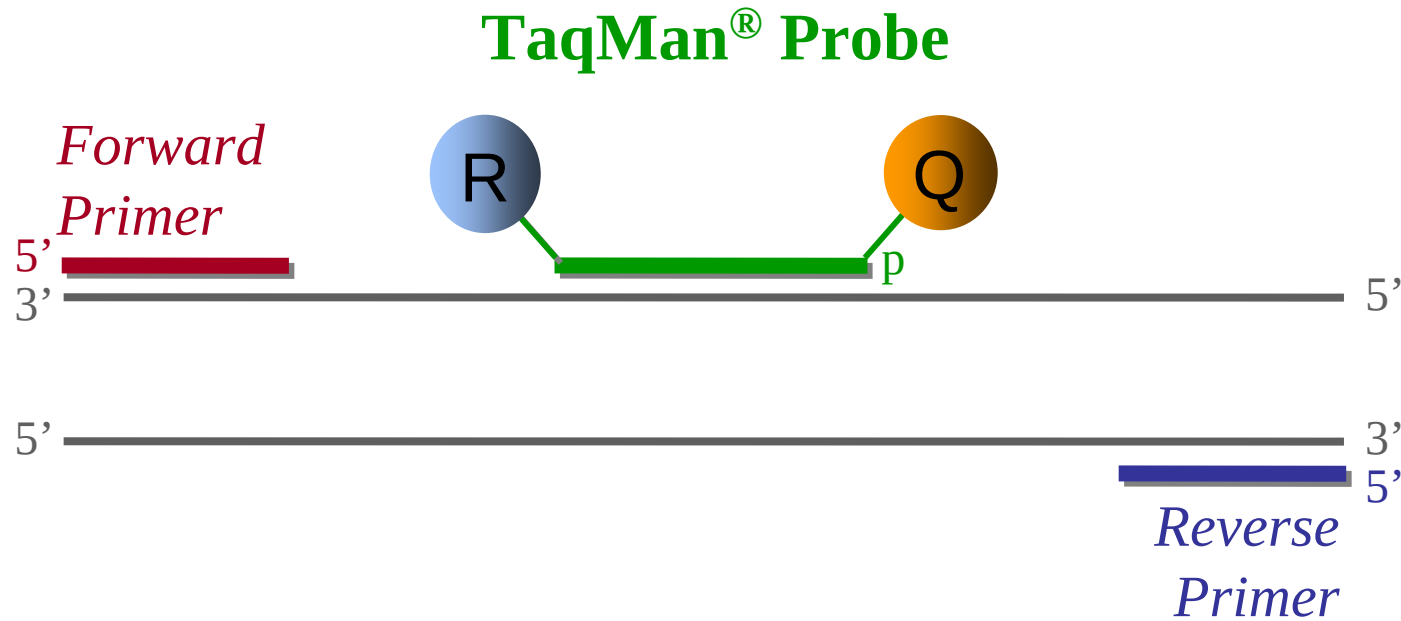
Fluorogenic 5' Nuclease Assay



Uses a TaqMan[®] probe

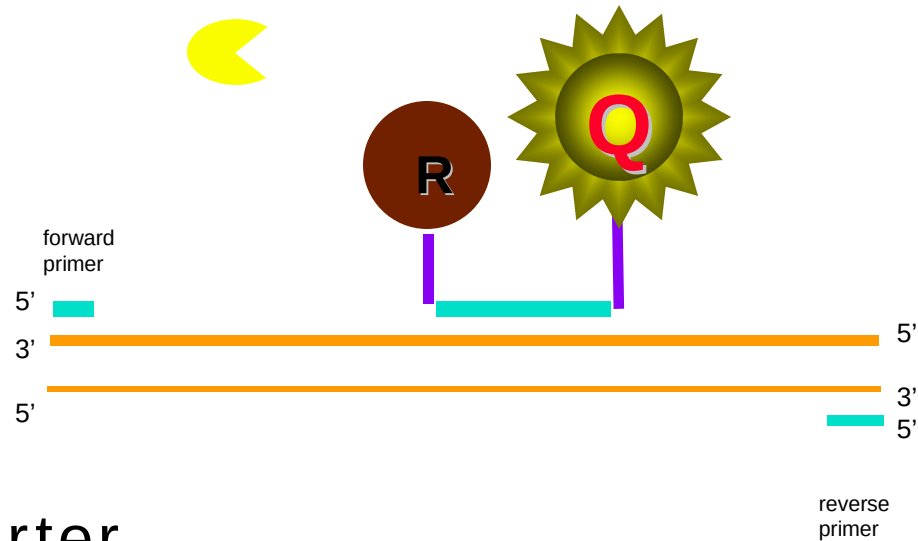
Real-Time PCR Chemistries

TaqMan Probe-Based Chemistry



TaqMan Probe-Based Chemistry

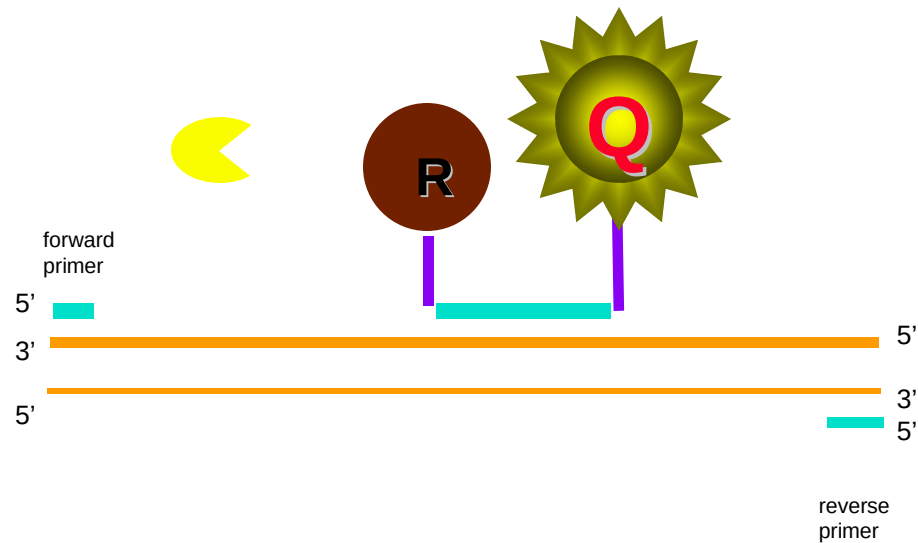
Denaturation → Annealing → Polymerization



R----- Reporter
Q----- Quencher

TaqMan Probe-Based Chemistry

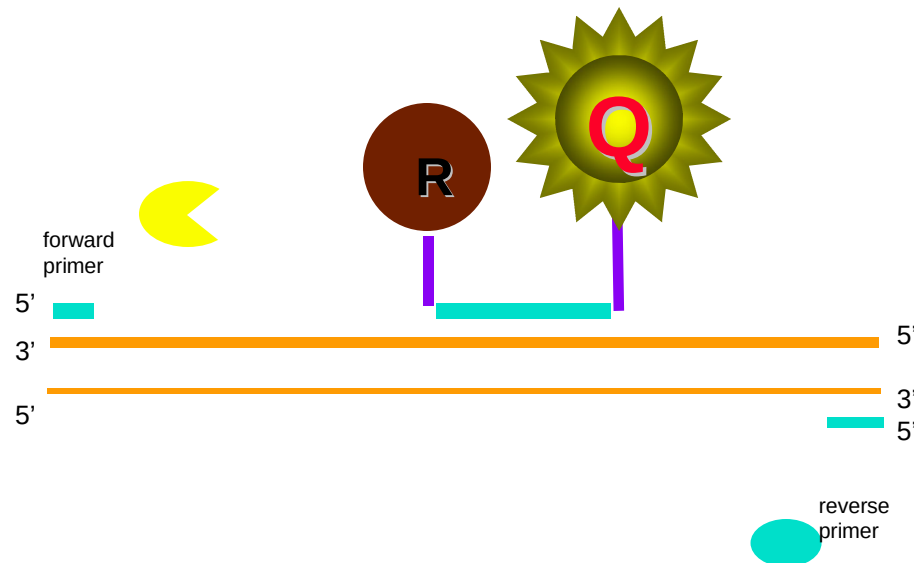
Denaturation → Annealing → Polymerization



R----- Reporter
Q----- Quencher

TaqMan Probe-Based Chemistry

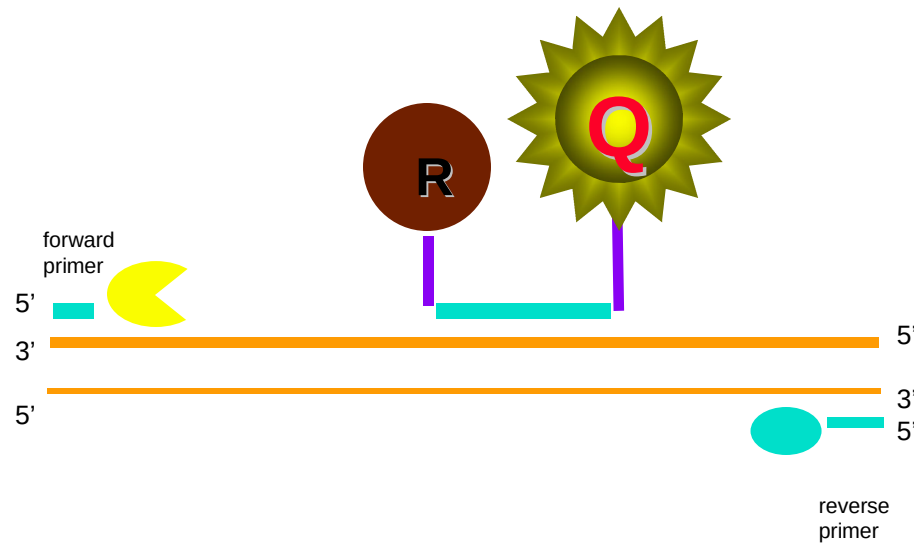
Denaturation → Annealing → Polymerization



R----- Reporter
Q----- Quencher

TaqMan Probe-Based Chemistry

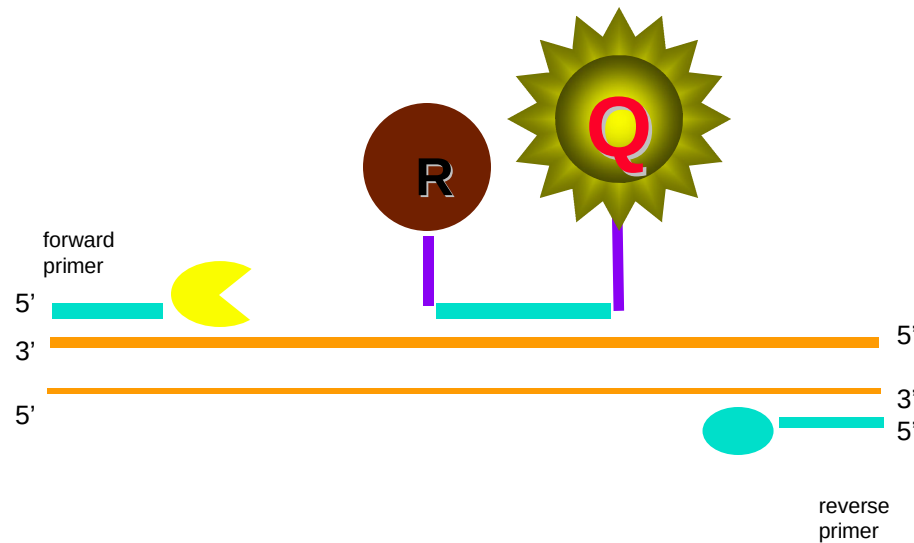
Denaturation → Annealing → Polymerization



R----- Reporter
Q----- Quencher

TaqMan Probe-Based Chemistry

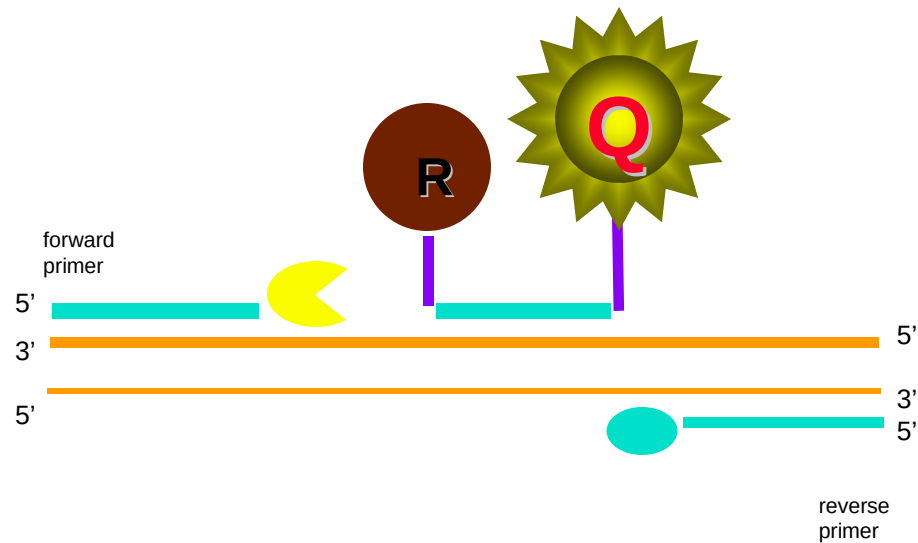
Denaturation → Annealing → Polymerization



R----- Reporter
Q----- Quencher

TaqMan Probe-Based Chemistry

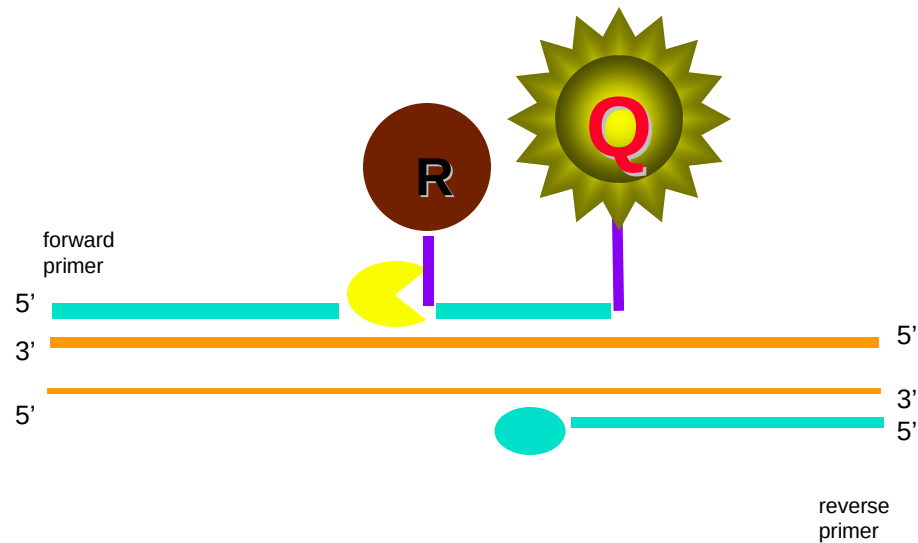
Denaturation → Annealing → Polymerization



R----- Reporter
Q----- Quencher

TaqMan Probe-Based Chemistry

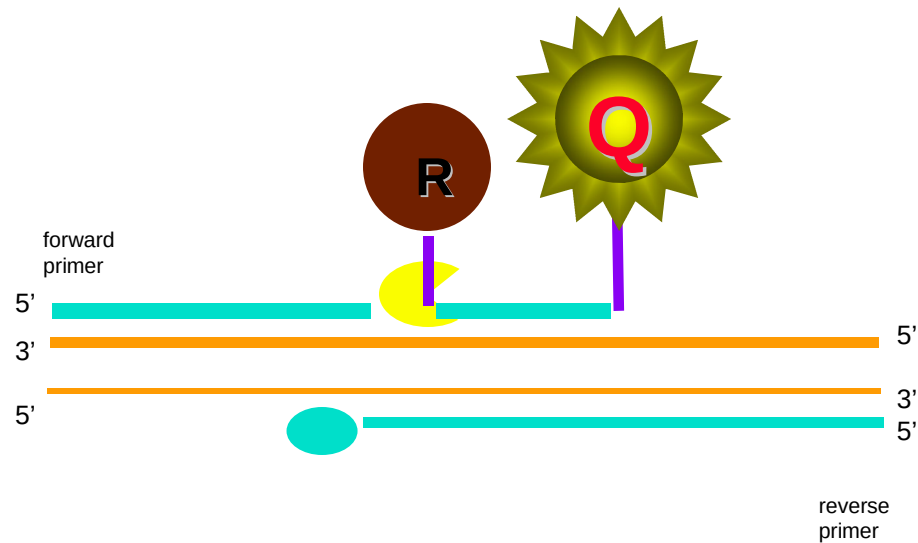
Denaturation → Annealing → Polymerization



R----- Reporter
Q----- Quencher

TaqMan Probe-Based Chemistry

Denaturation → Annealing → Polymerization

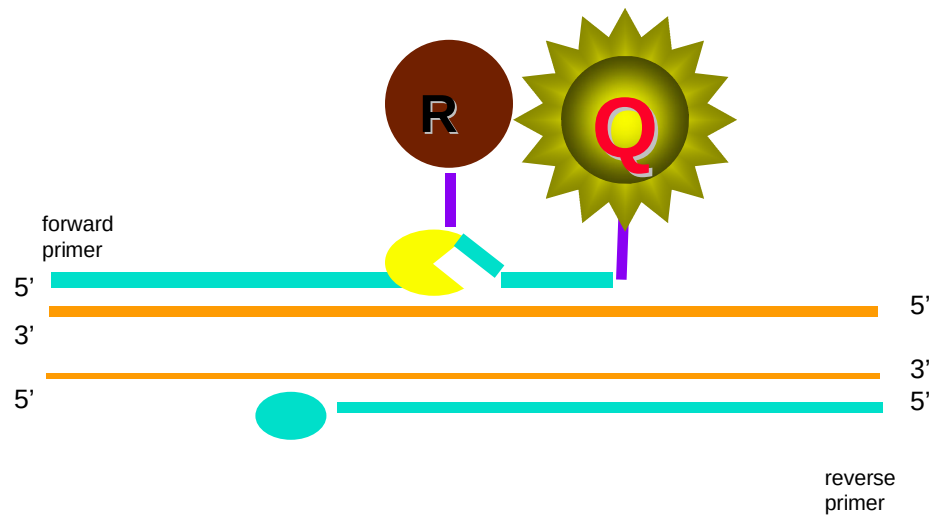


R----- Reporter
Q----- Quencher

TaqMan Probe-Based Chemistry

Denaturation → Annealing → Polymerization

. Strand Displacement

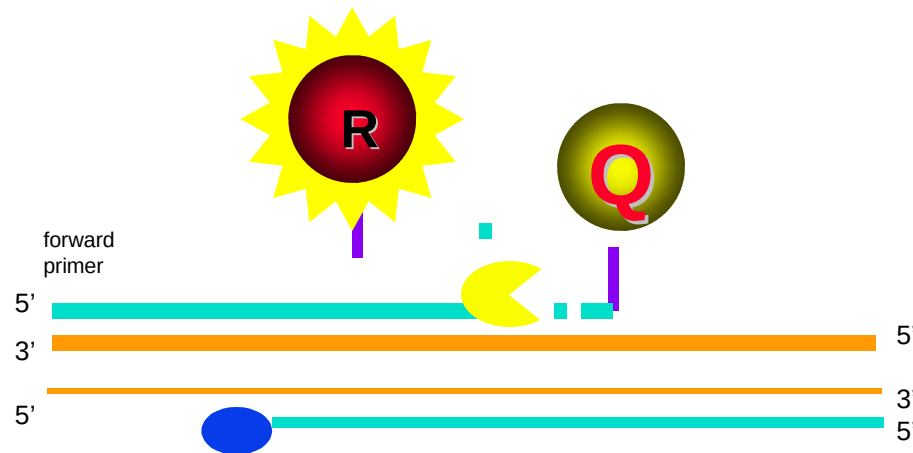


R----- Reporter
Q----- Quencher

TaqMan Probe-Based Chemistry

Denaturation → Annealing → Polymerization

. Cleavage



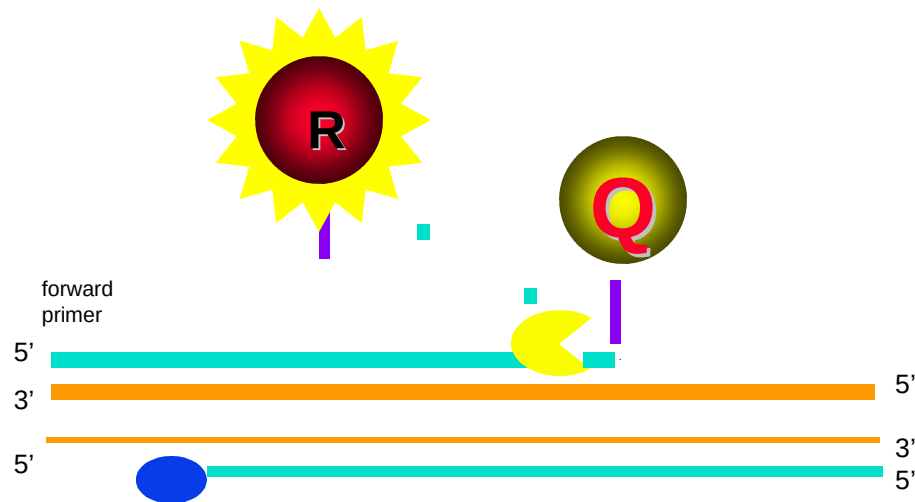
R----- Reporter

Q----- Quencher

TaqMan Probe-Based Chemistry

Denaturation → Annealing → Polymerization

. Cleavage



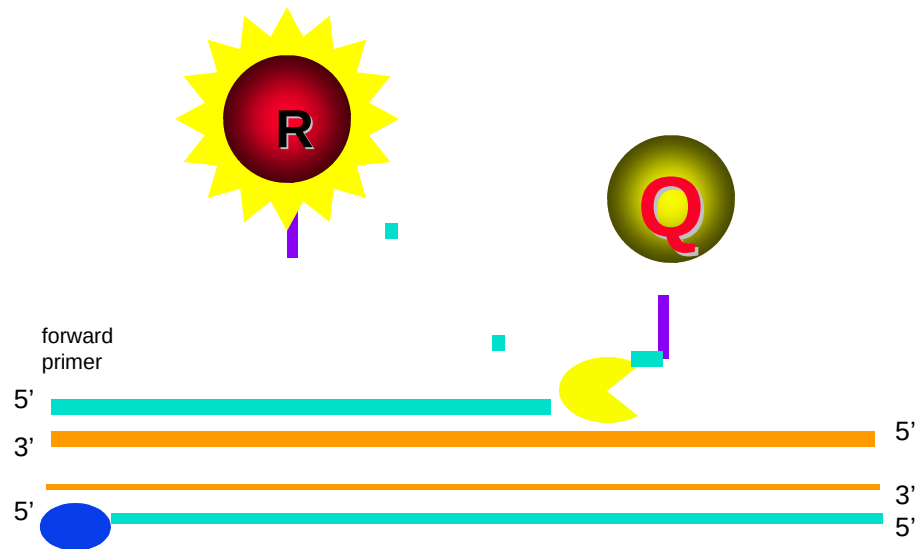
R----- Reporter

Q----- Quencher

TaqMan Probe-Based Chemistry

Denaturation → Annealing → Polymerization

. Cleavage

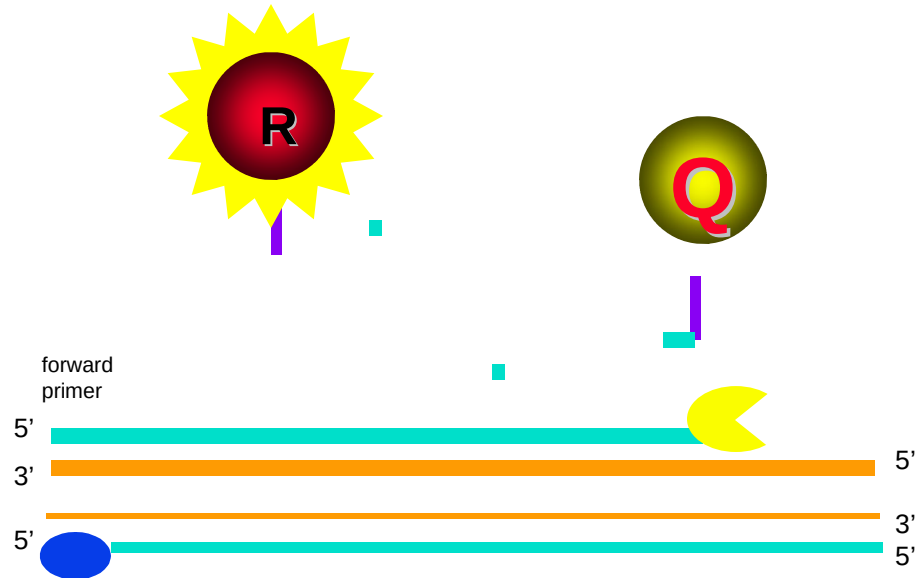


R----- Reporter
Q----- Quencher

TaqMan Probe-Based Chemistry

Denaturation → Annealing → Polymerization

. Cleavage

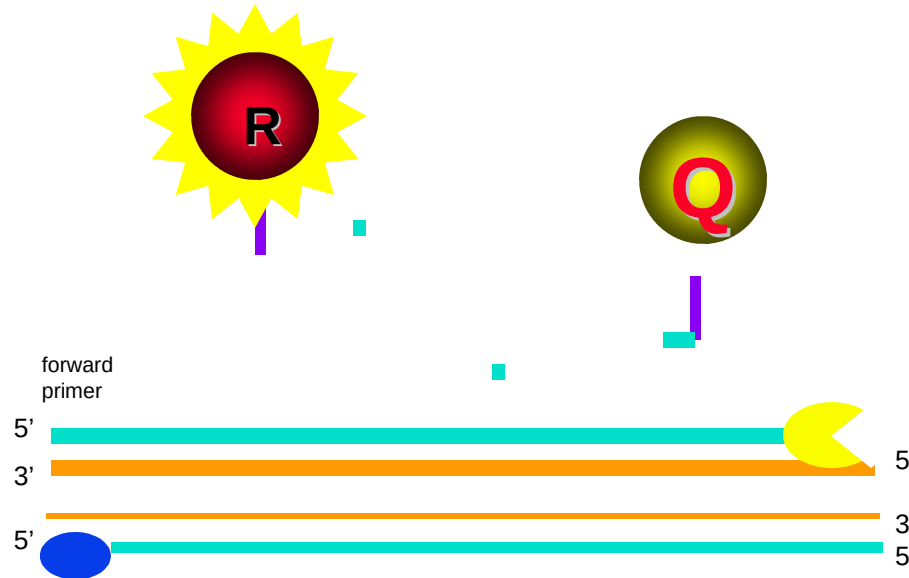


R----- Reporter
Q----- Quencher

TaqMan Probe-Based Chemistry

Denaturation → Annealing → Polymerization

Polymerization Completed



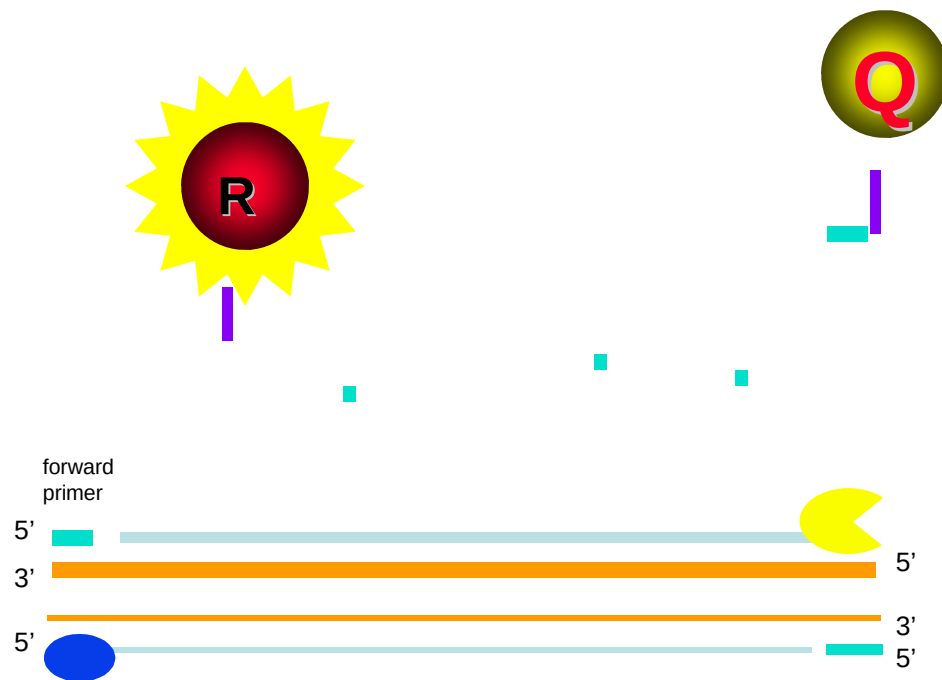
R----- Reporter

Q----- Quencher

TaqMan Probe-Based Chemistry

Denaturation → Annealing → Polymerization

Polymerization Completed

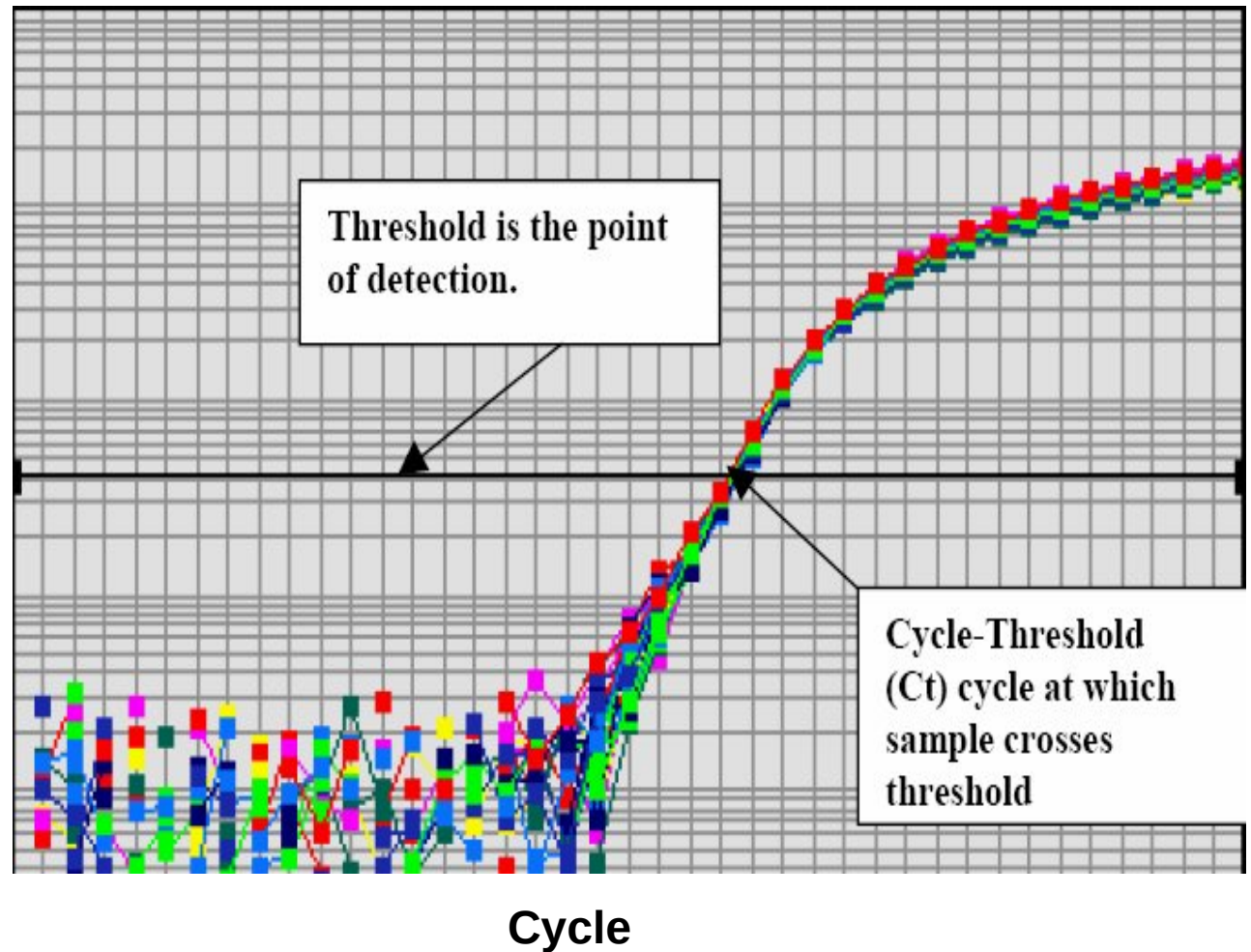


R----- Reporter
Q----- Quencher

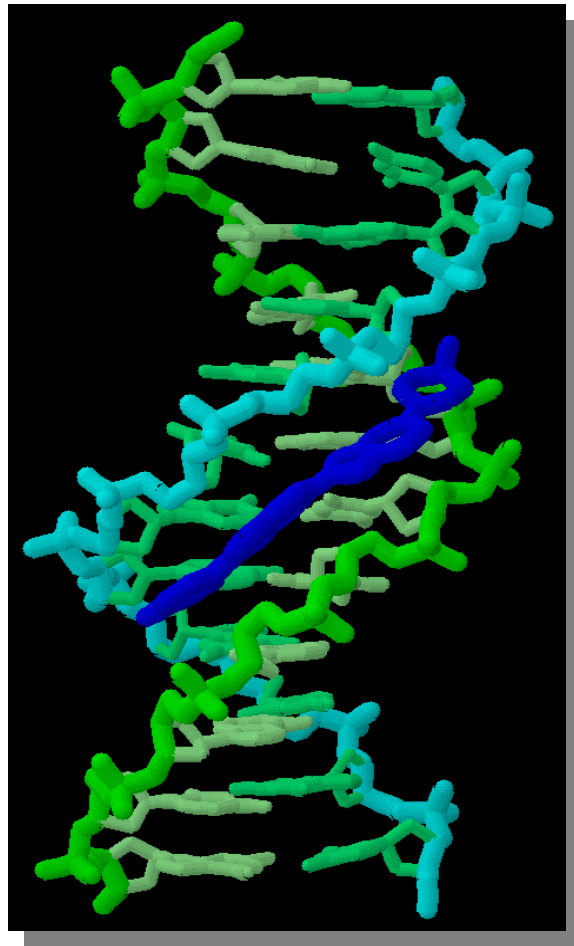
Real-Time PCR Principles

Real-Time Amplification Plots

Rn: Reporter Signal

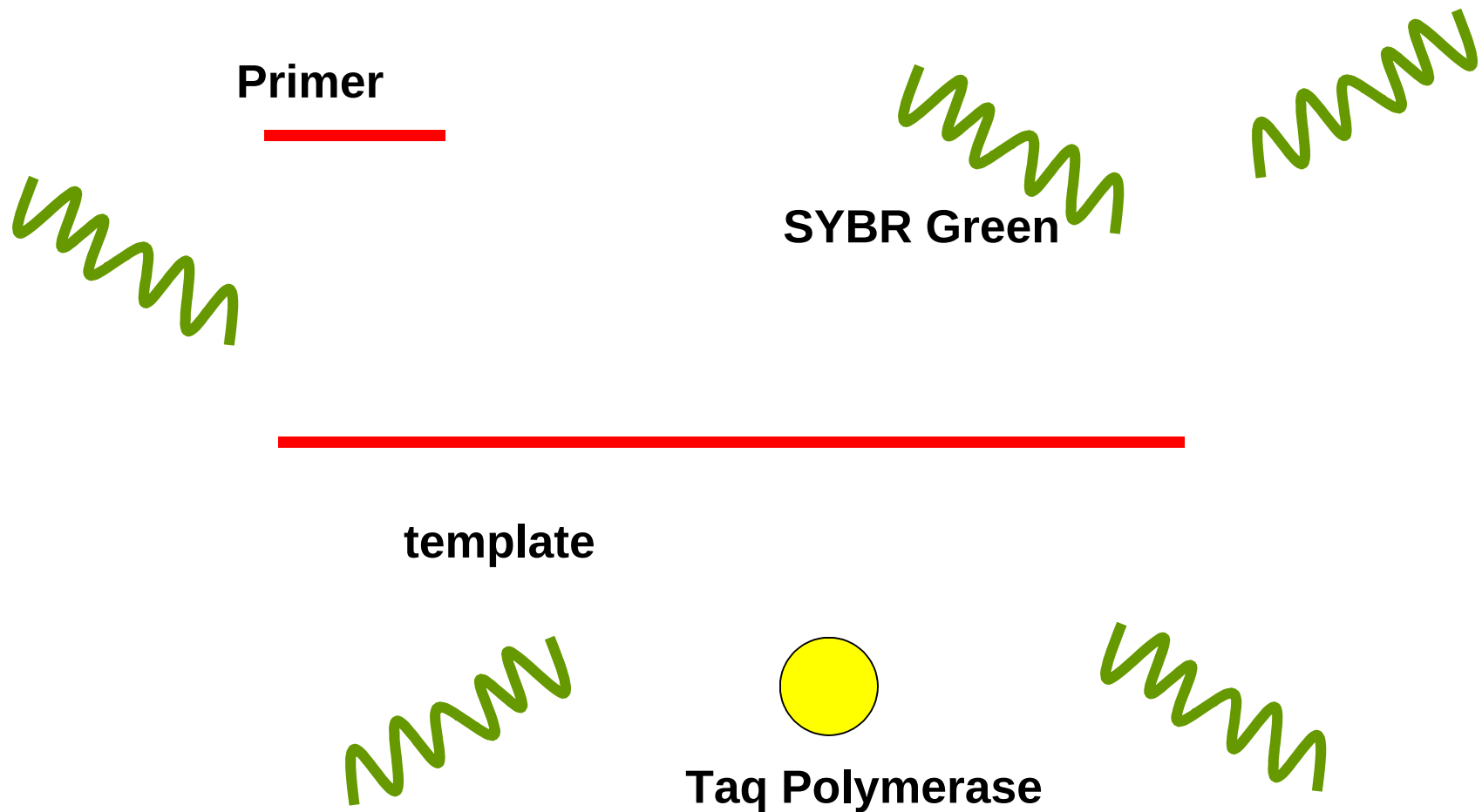


SYBR[®] green chemistry

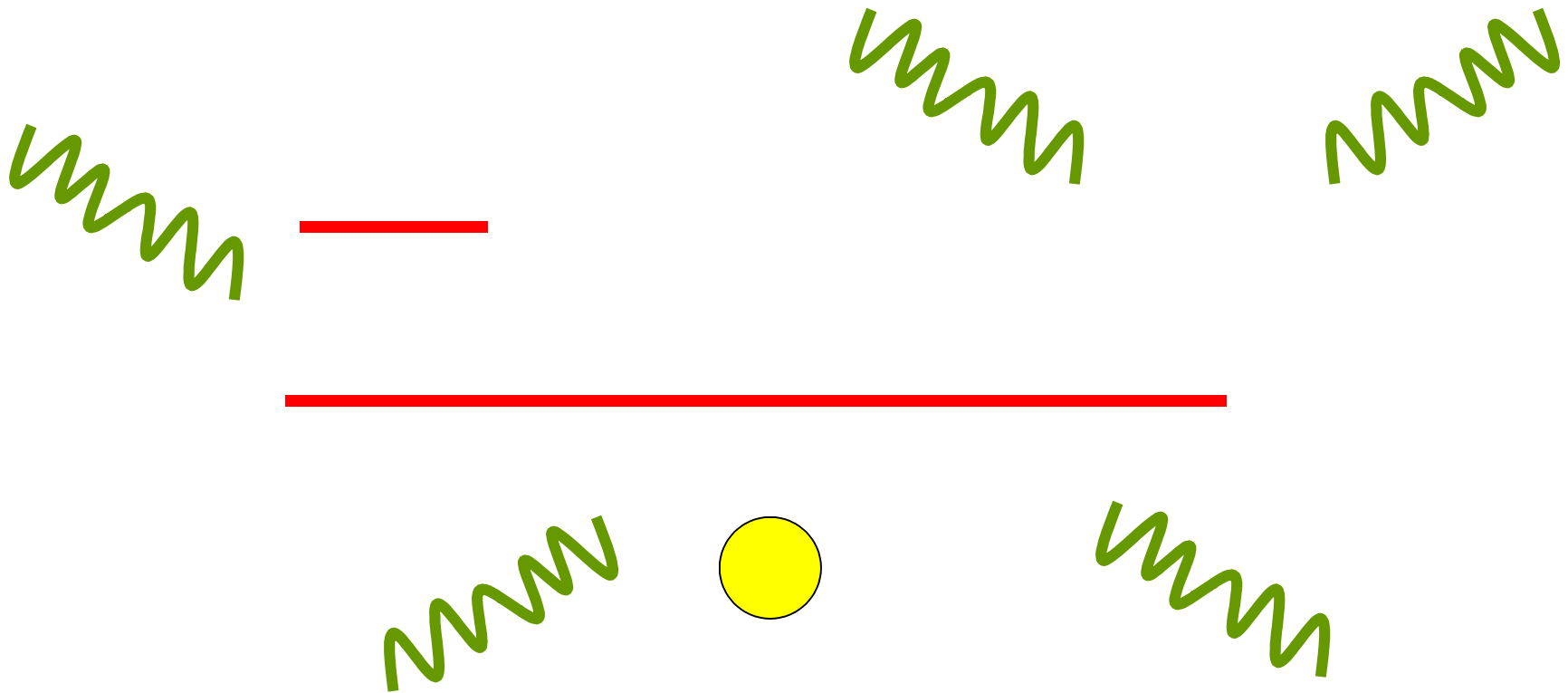


**Double stranded DNA
Binding Dye**

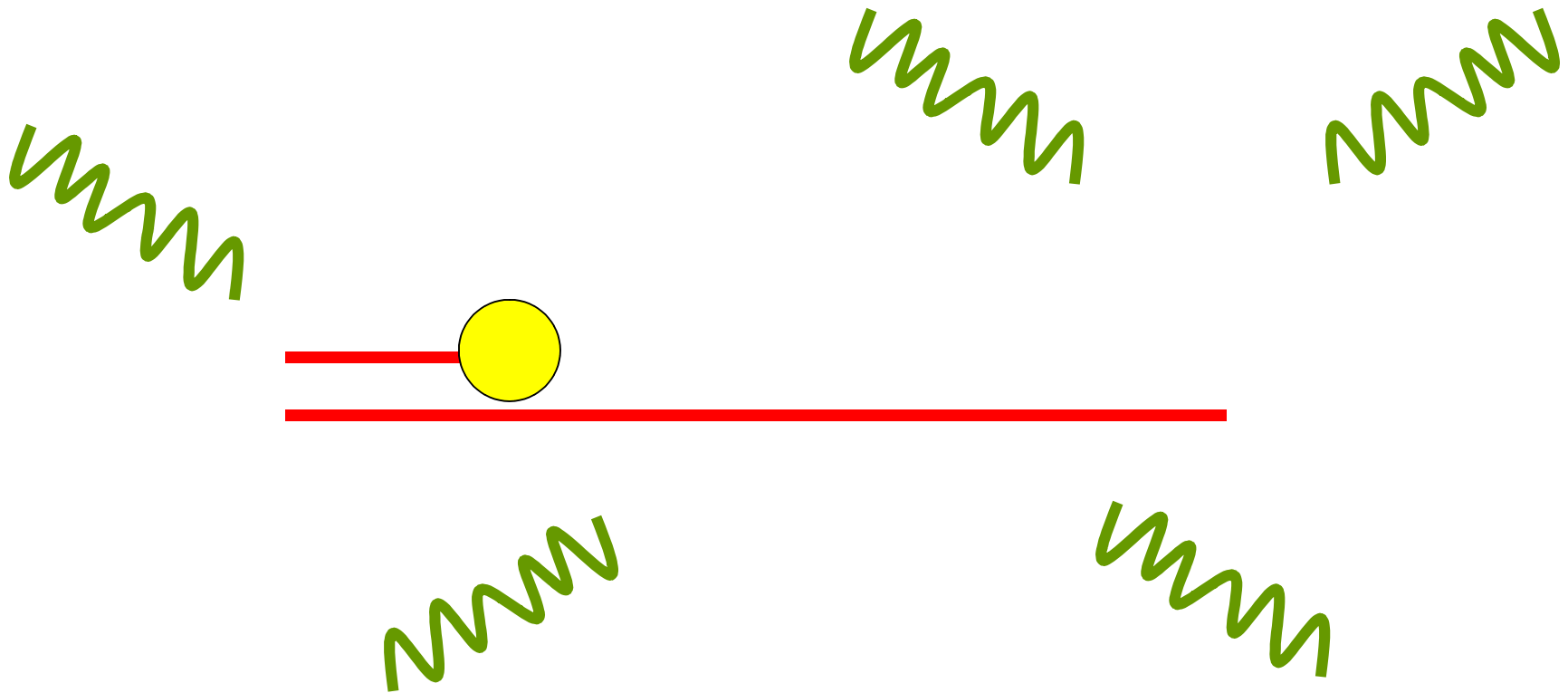
SYBR Green detection



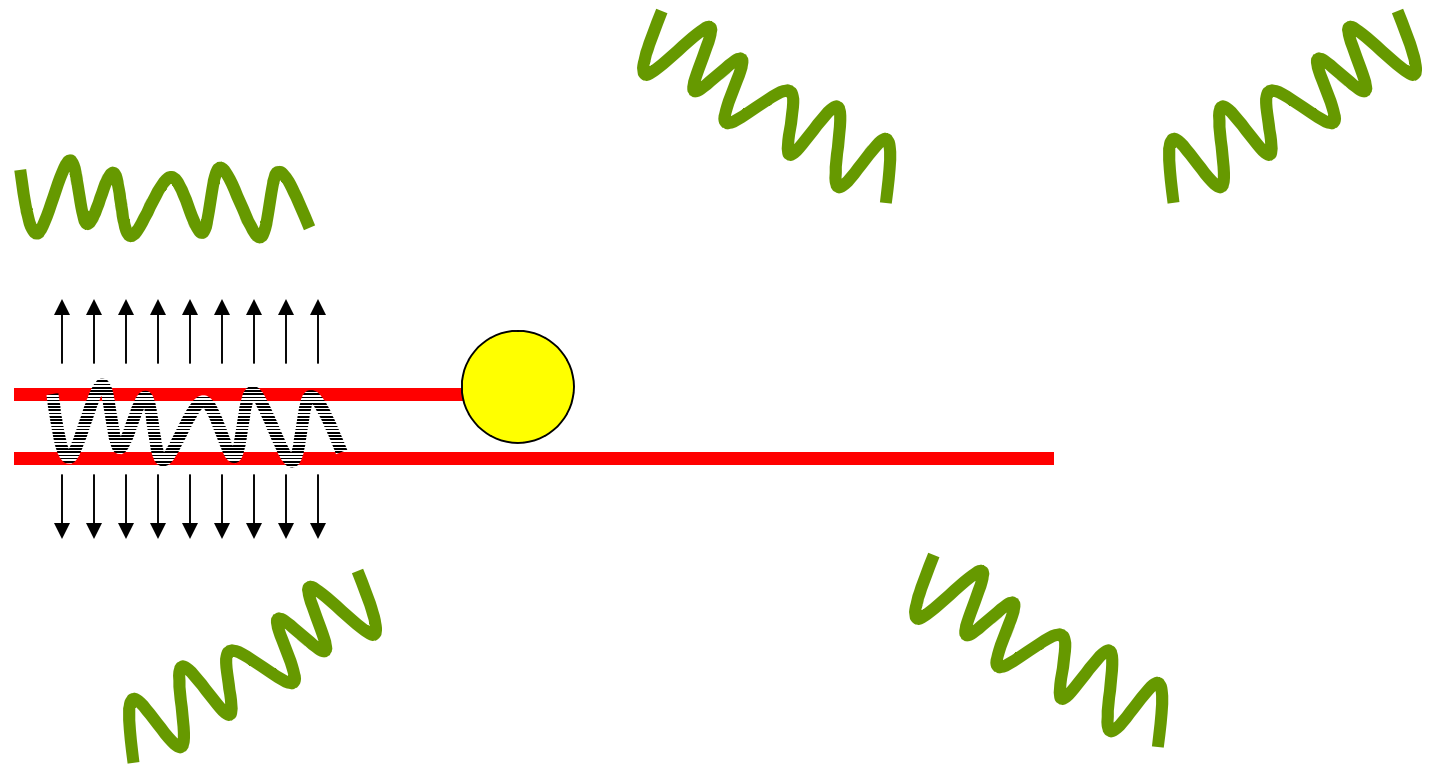
SYBR Green detection



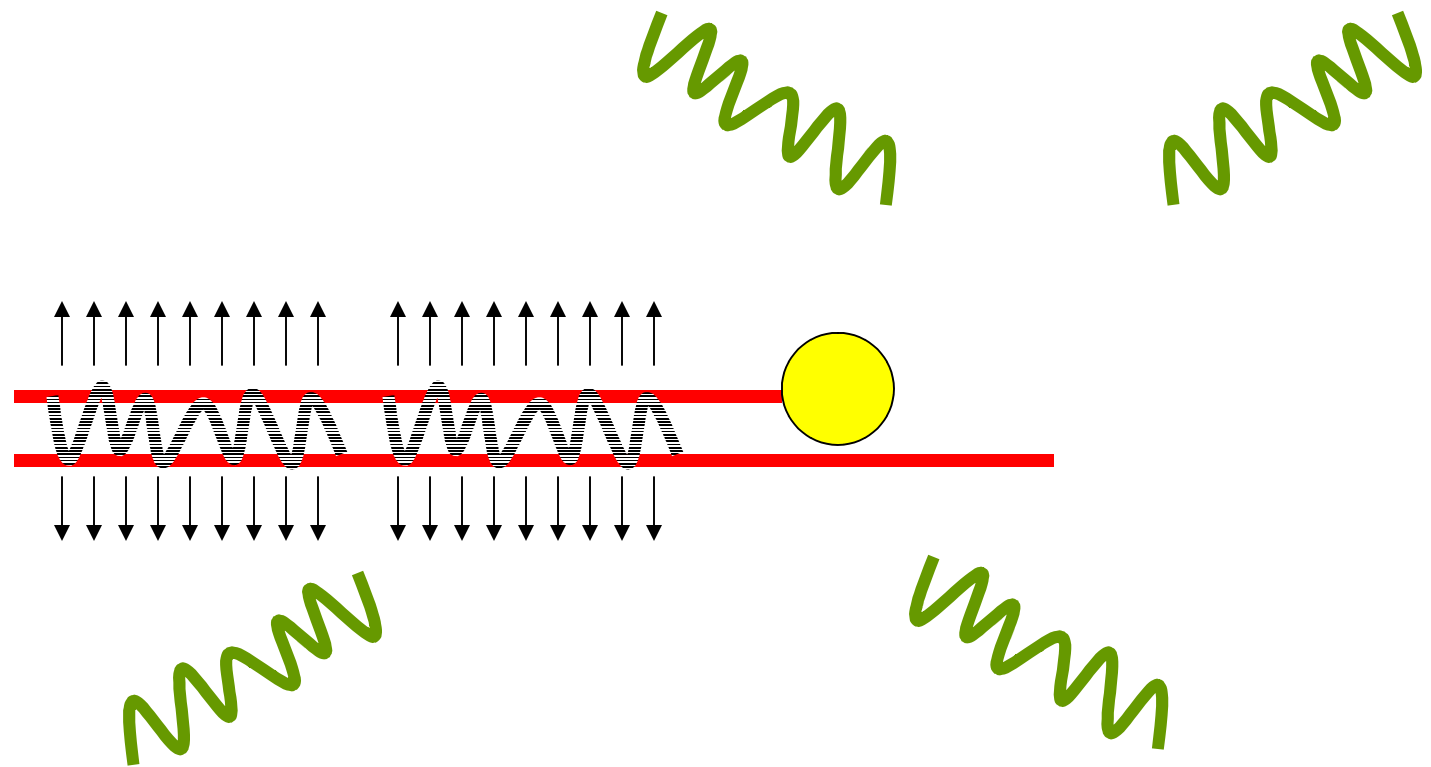
SYBR Green detection



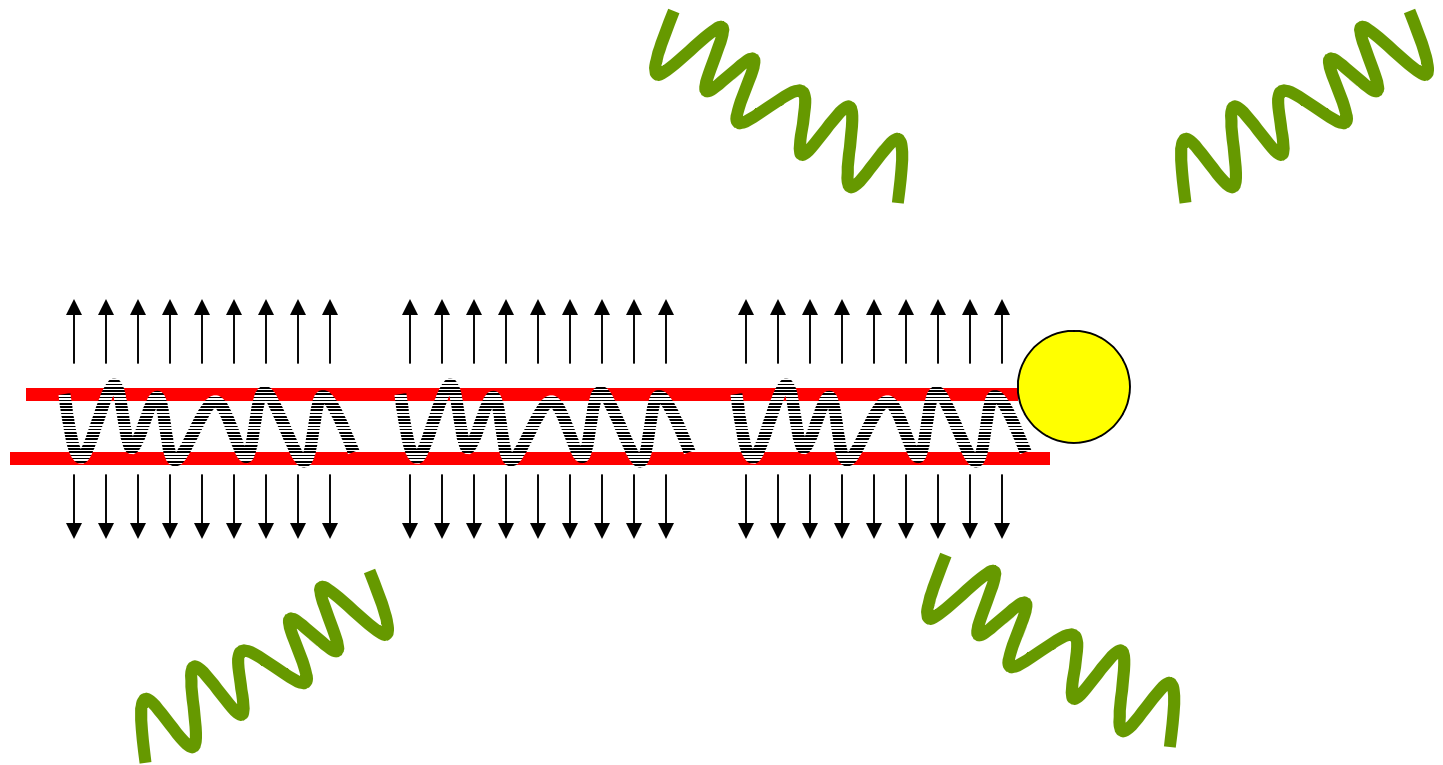
SYBR Green detection



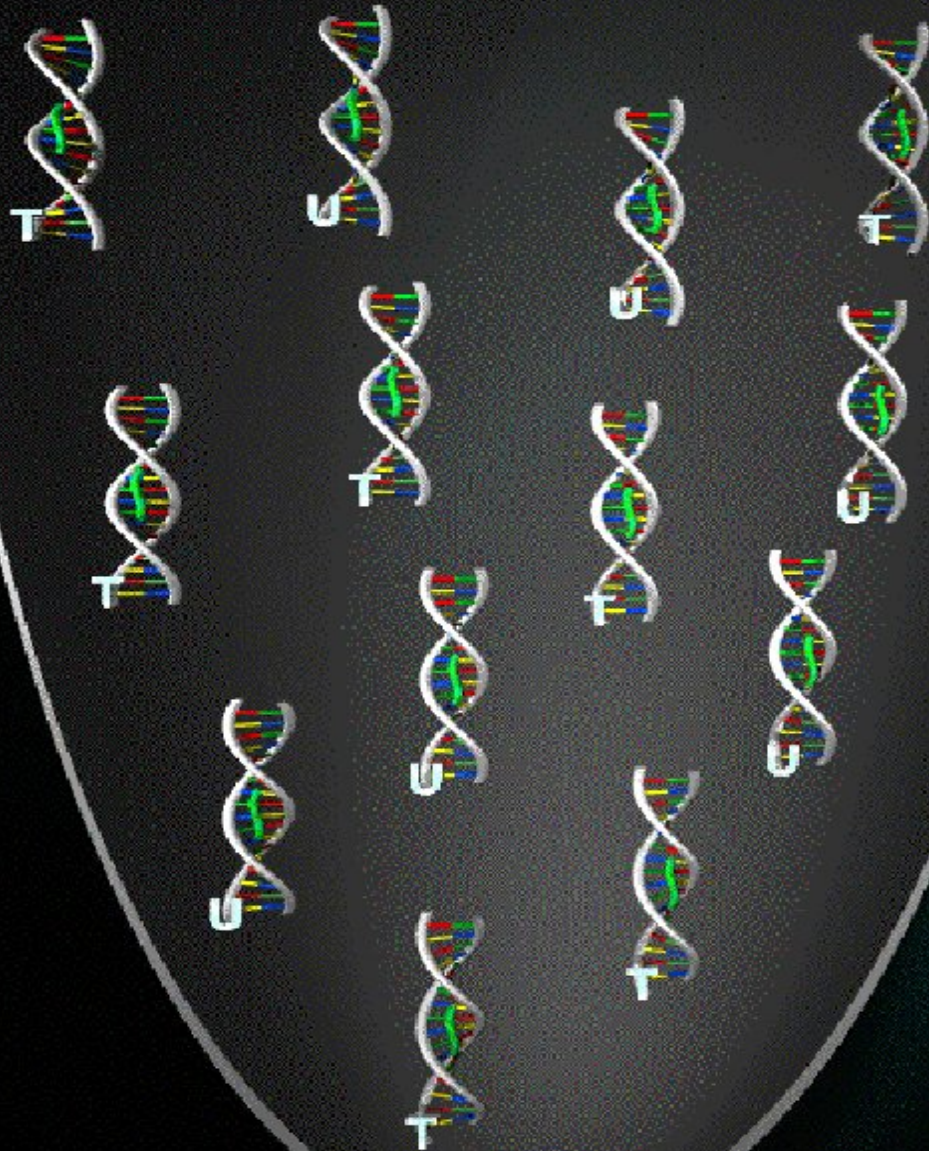
SYBR Green detection



SYBR Green detection



CARRY-OVER PREVENTION WITH UNG

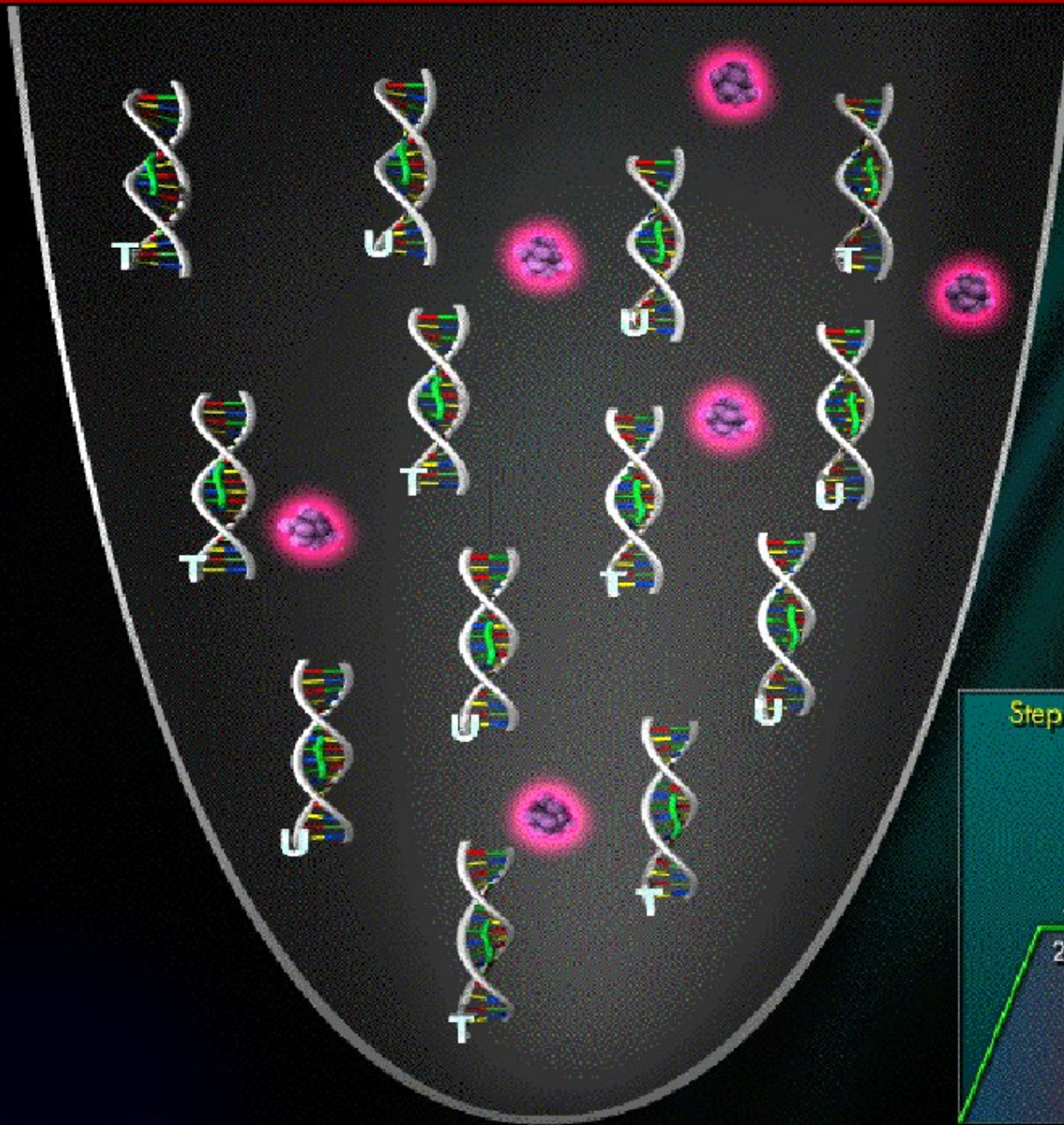


First, the system incubates the reaction for two minutes at 50 °C, activating the UNG.

Step 1

50
2:00

CARRY-OVER PREVENTION WITH UNG



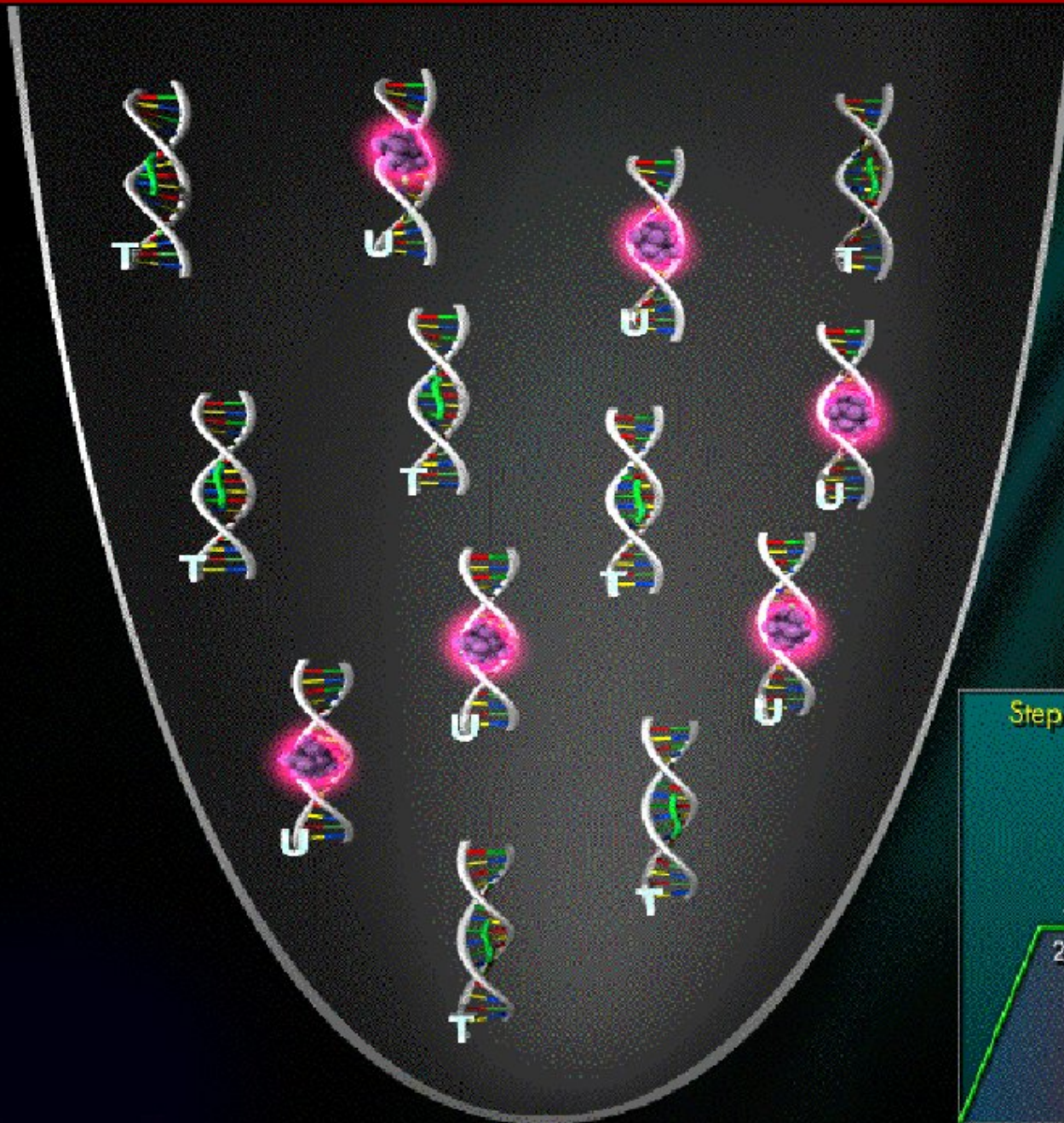
First, the system incubates the reaction for two minutes at 50 °C, activating the UNG.

Step 1

50

2:00

CARRY-OVER PREVENTION WITH UNG



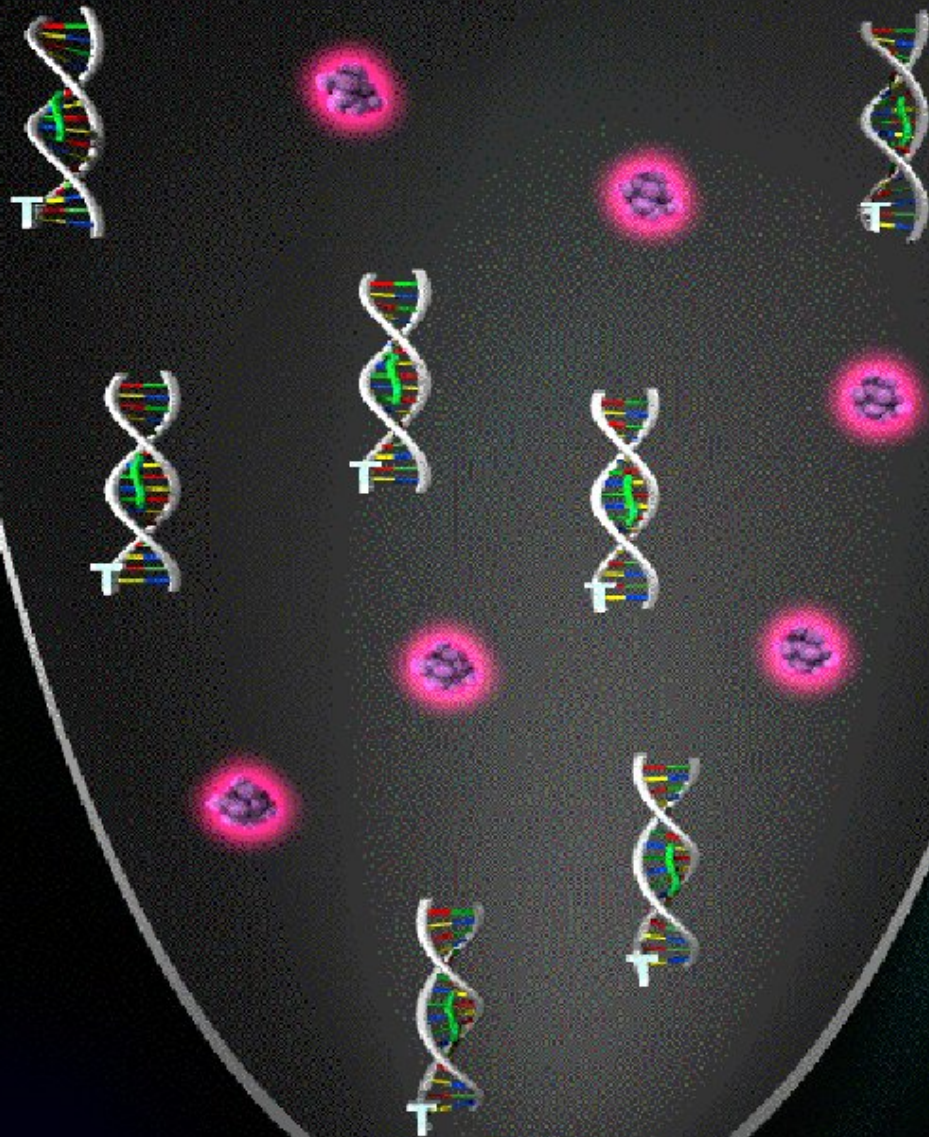
Since amplicons contain U bases instead of T, UNG digests all the amplicons during this incubation, leaving the genomic DNA intact.

Step 1

50

2:00

CARRY-OVER PREVENTION WITH UNG



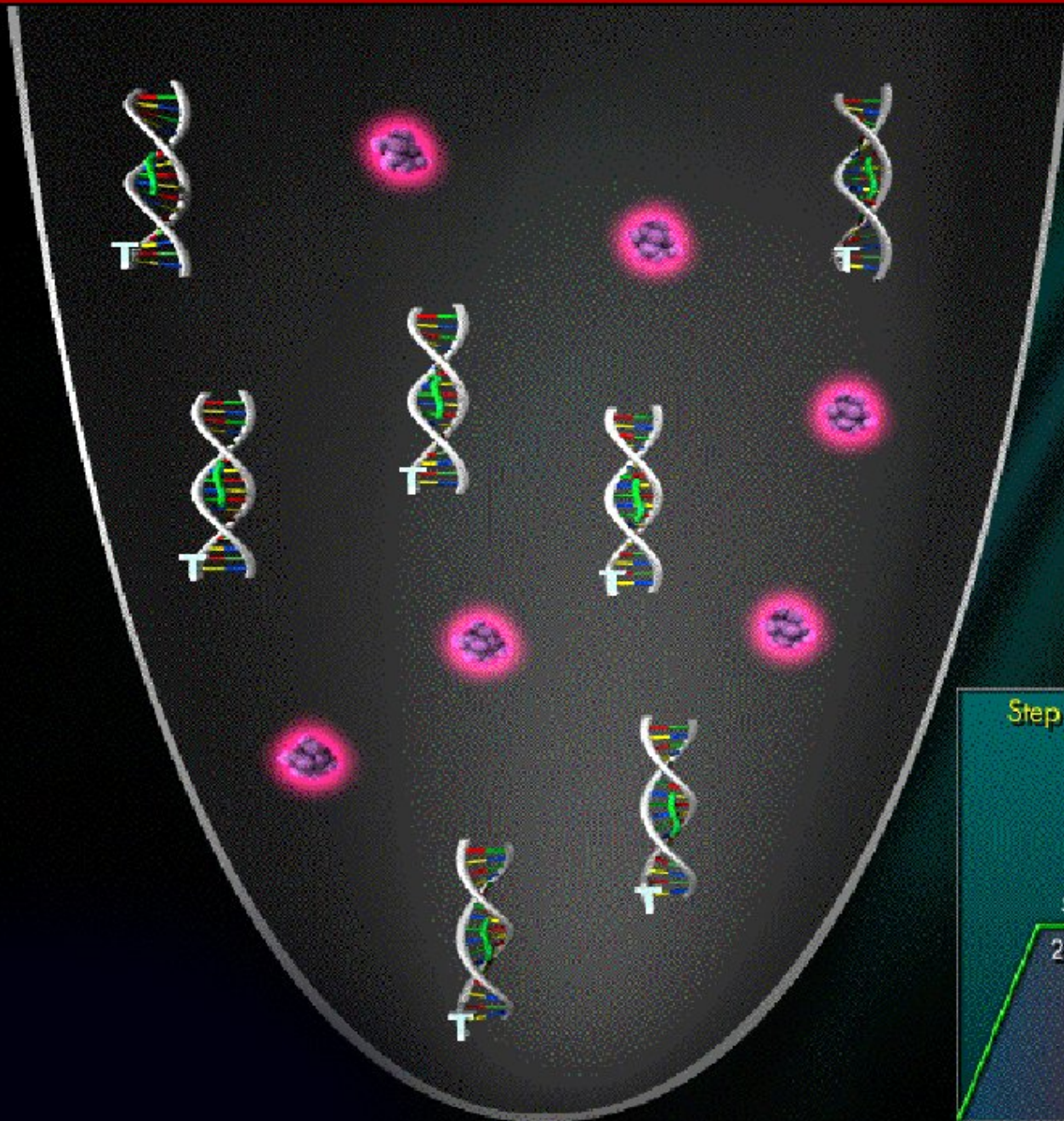
Since amplicons contain U bases instead of T, UNG digests all the amplicons during this incubation, leaving the genomic DNA intact.

Step 1

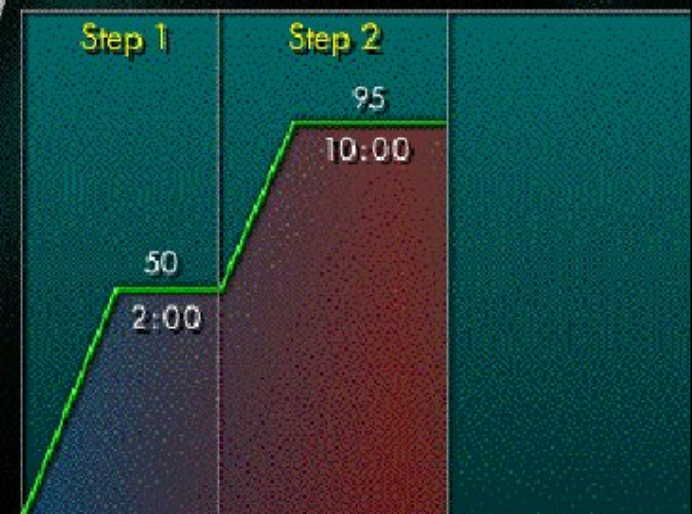
50

2:00

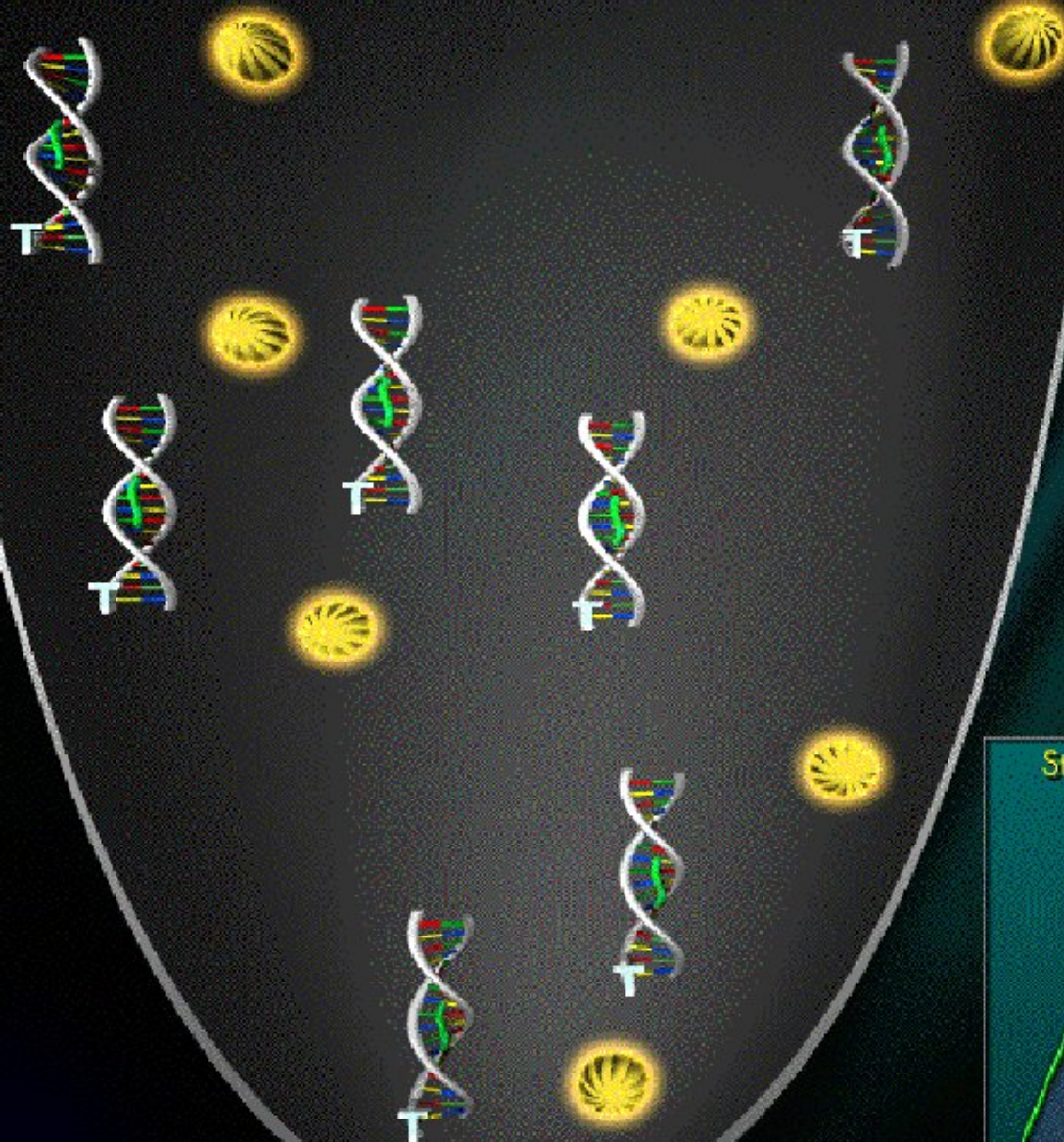
CARRY-OVER PREVENTION WITH UNG



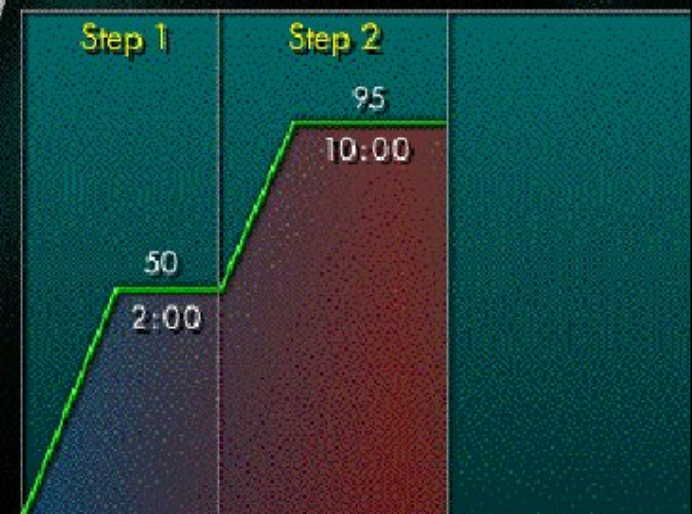
Next, the system raises the temperature to 95 °C for ten minutes, deactivating the UNG and activating the AmpliTaq Gold® enzyme.



CARRY-OVER PREVENTION WITH UNG / AmpliTaq Gold™ ACTIVATION

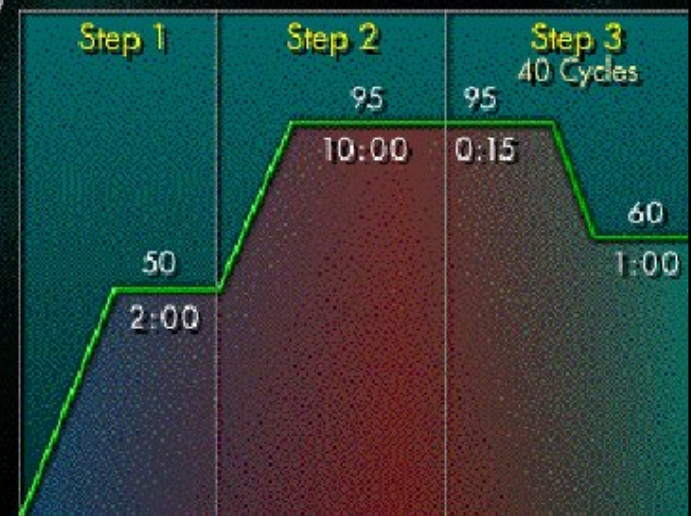


Next, the system raises the temperature to 95 °C for ten minutes, deactivating the UNG and activating the AmpliTaq Gold® enzyme.



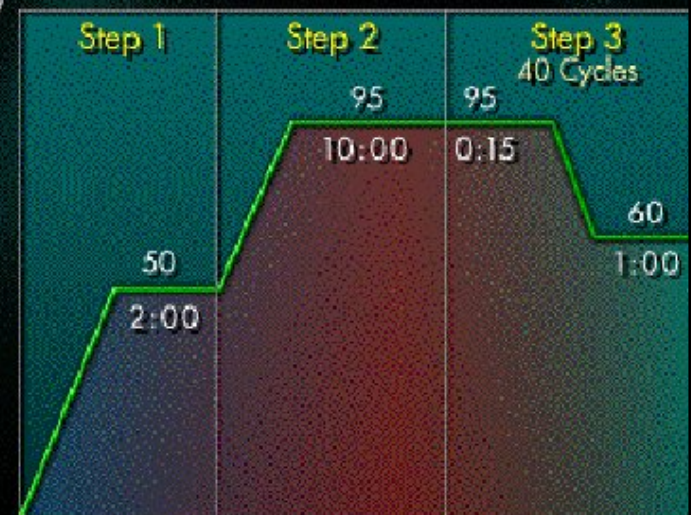
CARRY-OVER PREVENTION WITH UNG / AmpliTaq Gold™ CYCLING

During cycling, the reaction amplifies the target...

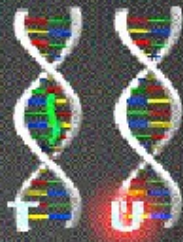
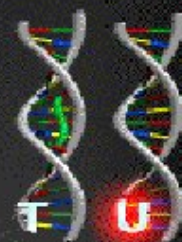
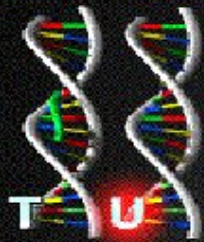


CARRY-OVER PREVENTION WITH UNG / AmpliTaq Gold™ CYCLING

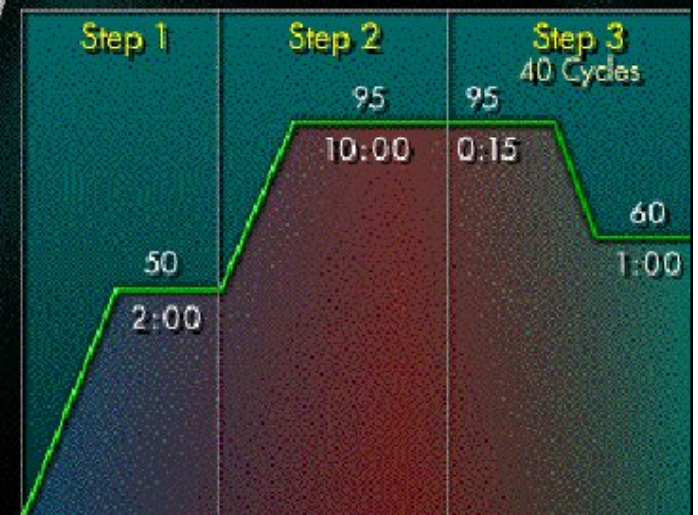
During cycling, the reaction amplifies the target...



CARRY-OVER PREVENTION WITH UNG / AmpliTaq Gold™ CYCLING



Creating U-containing amplicons.



Types of Quantitation Assays

```
graph TD; A[Types of Quantitation Assays] --> B[Absolute quantitation]; A --> C[Relative quantitation]; B --> D[Provides absolute measurement of starting copy number]; D --> E[–Requires standards of known quantity]; D --> F[–e.g. Detecting DNA copy number for forensics purposes];
```

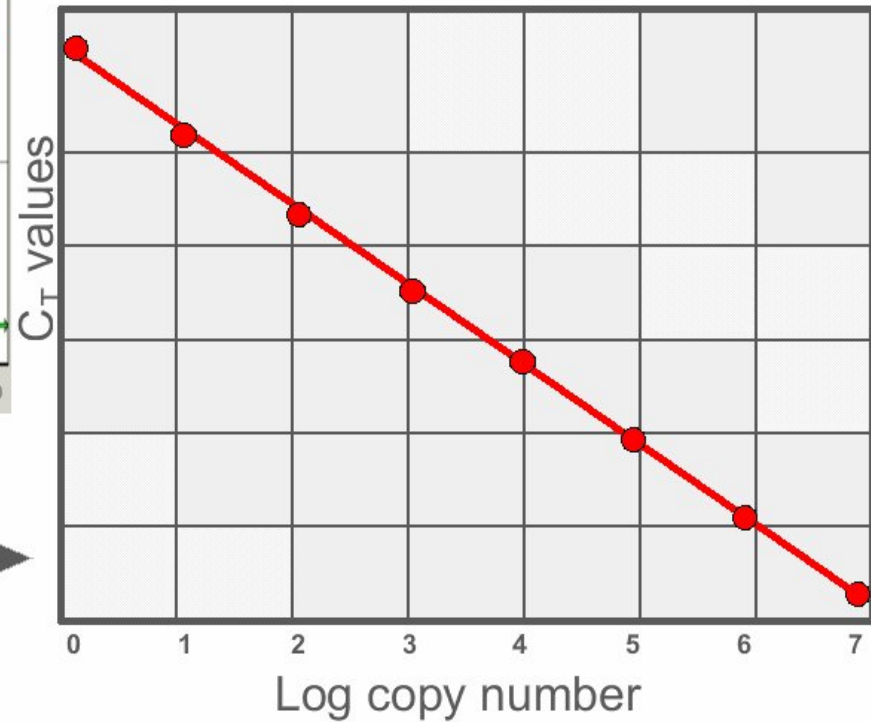
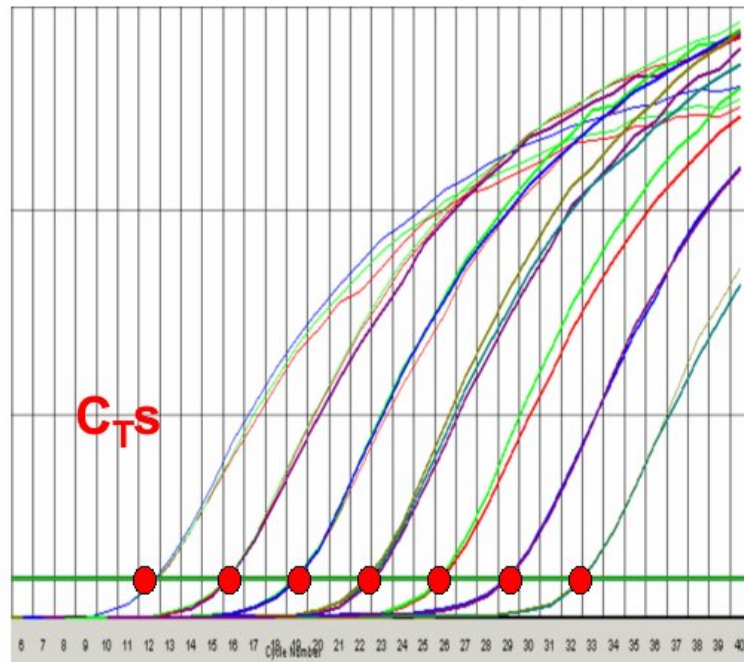
Absolute quantitation

Relative quantitation

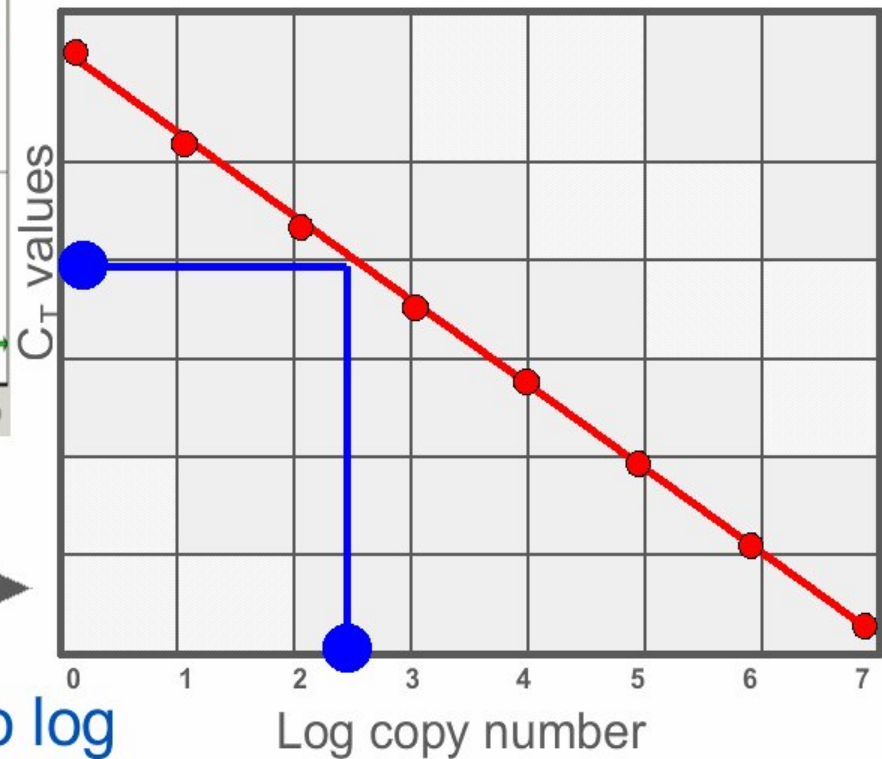
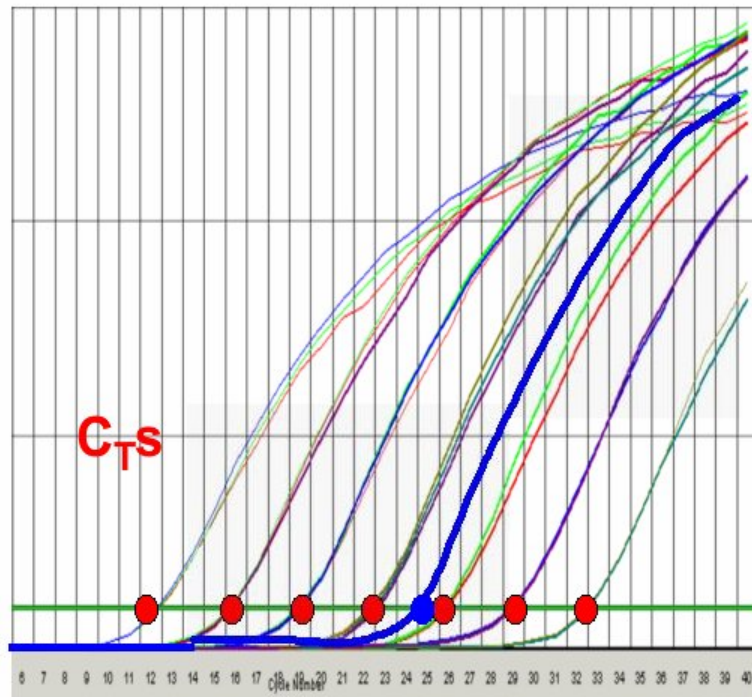
Provides absolute measurement of starting copy number

- Requires standards of known quantity
- e.g. Detecting DNA copy number for forensics purposes

From Fluorescence to Copy Number

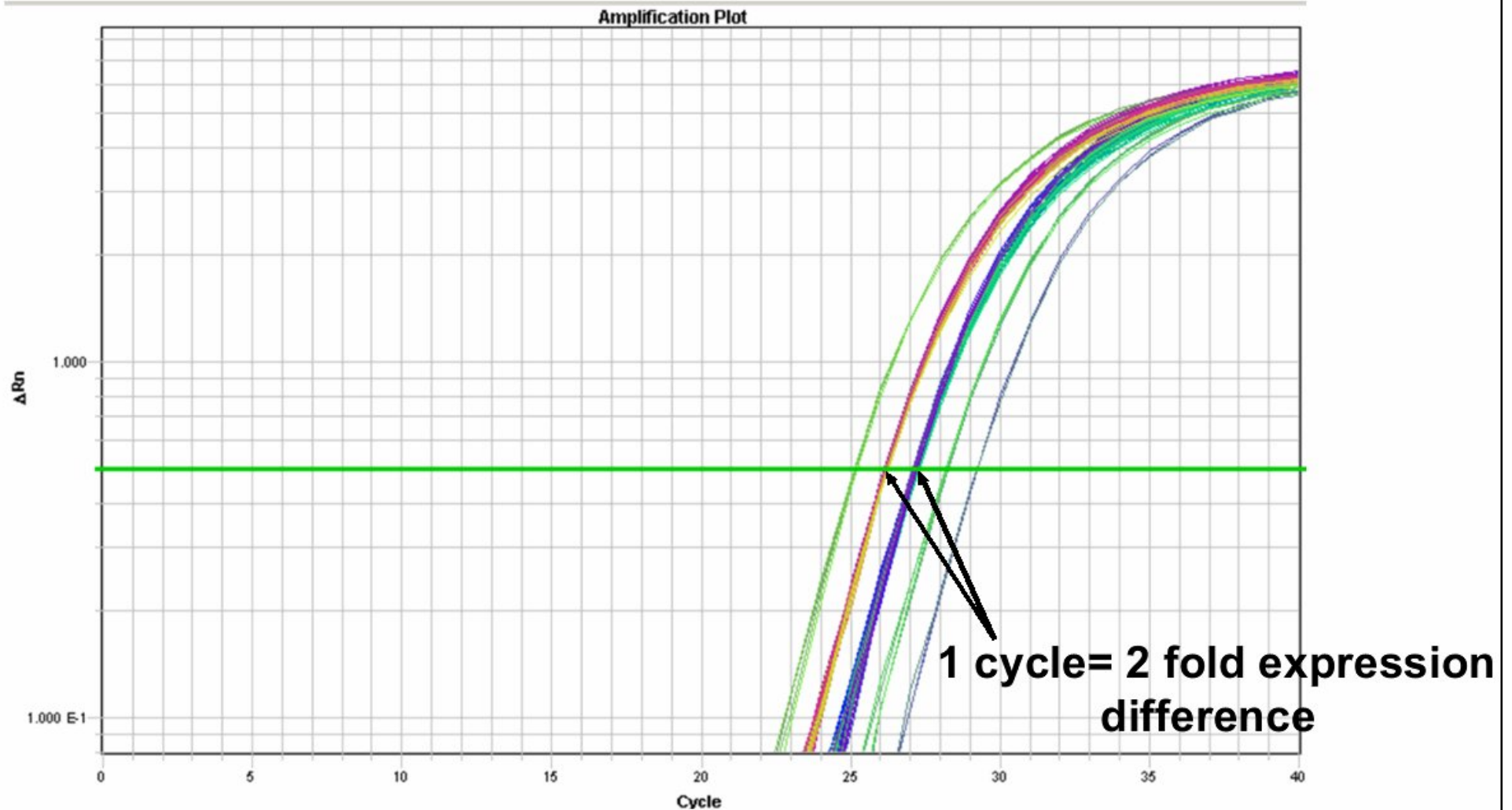


From Fluorescence to Copy Number



C_T is directly proportional to log of amount of input template

High (100%) Amplification Efficiency



Types of Quantitation Assays

```
graph TD; A[Types of Quantitation Assays] --> B[Absolute quantitation]; A --> C[Relative quantitation]; C --> D[Provides accurate discrimination between relative amounts of starting material]; D --> E["-e.g. Comparing expression levels of wildtype vs. mutated alleles"]; D --> F["-e.g. Comparing expression levels of a gene across different tissues or between different biological conditions"]; D --> G["-e.g. Validating array results"];
```

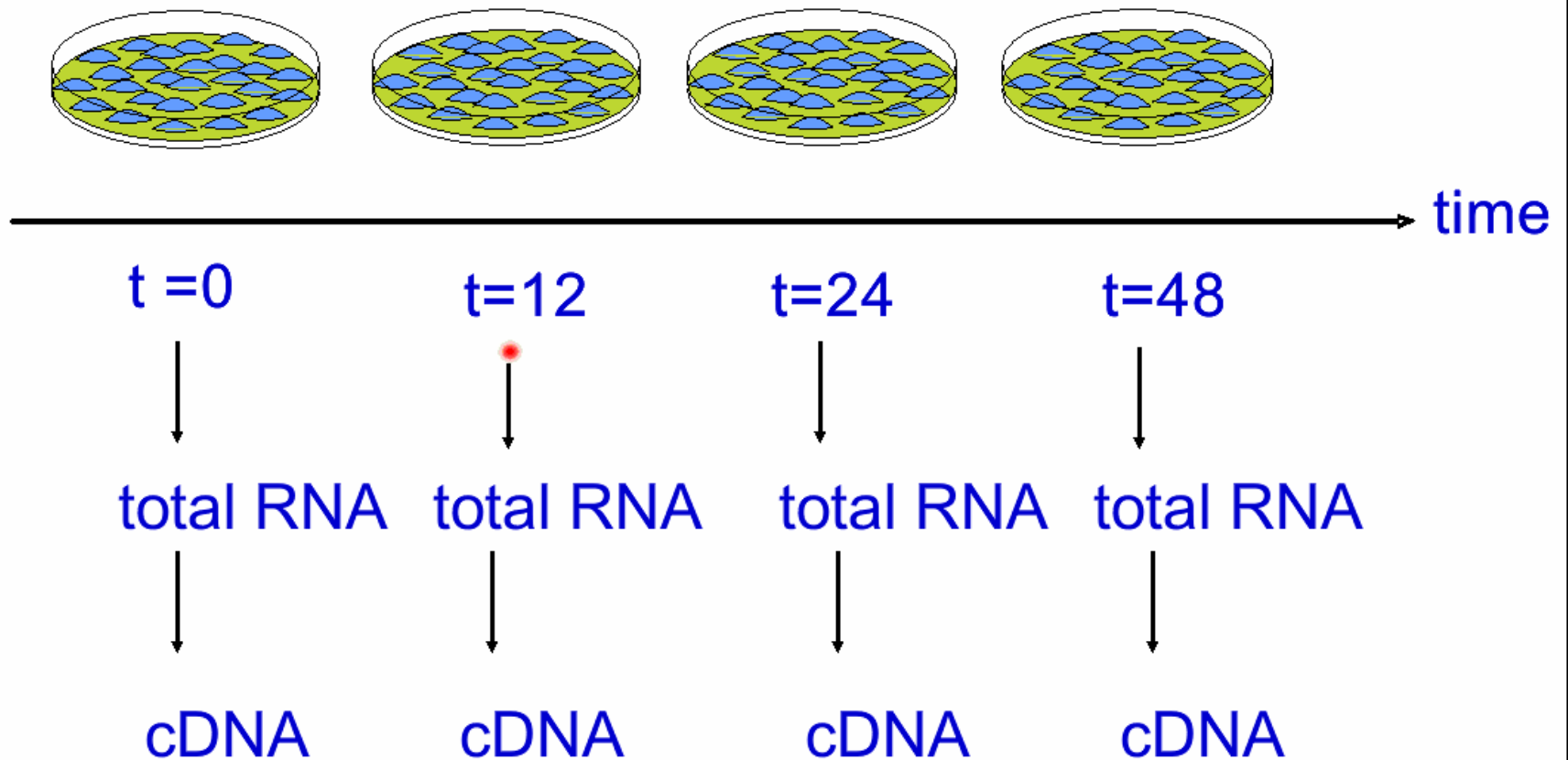
Absolute quantitation

Relative quantitation

Provides accurate discrimination between relative amounts of starting material

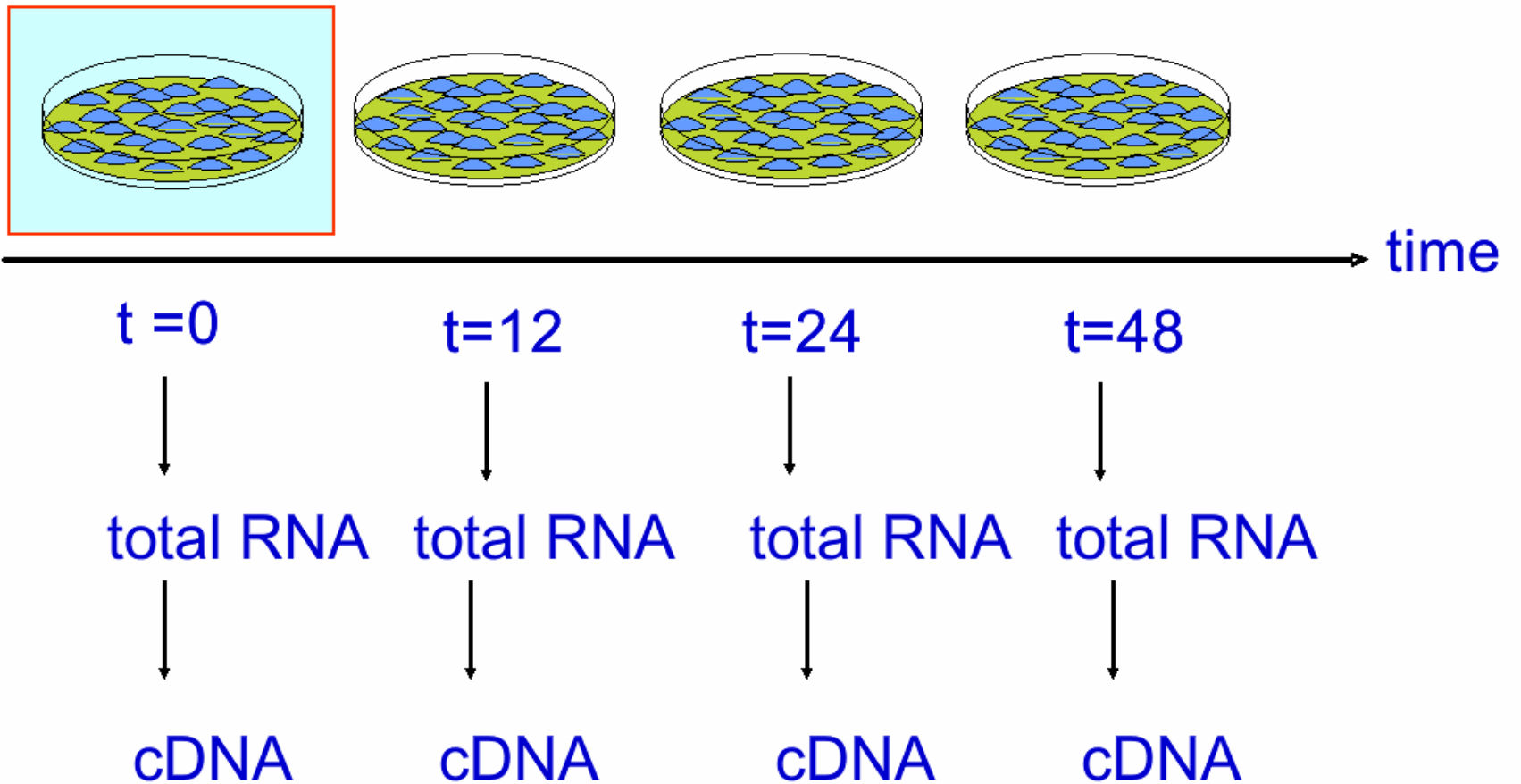
- e.g. Comparing expression levels of wildtype vs. mutated alleles
- e.g. Comparing expression levels of a gene across different tissues or between different biological conditions
- e.g. Validating array results

Relative Quantitation



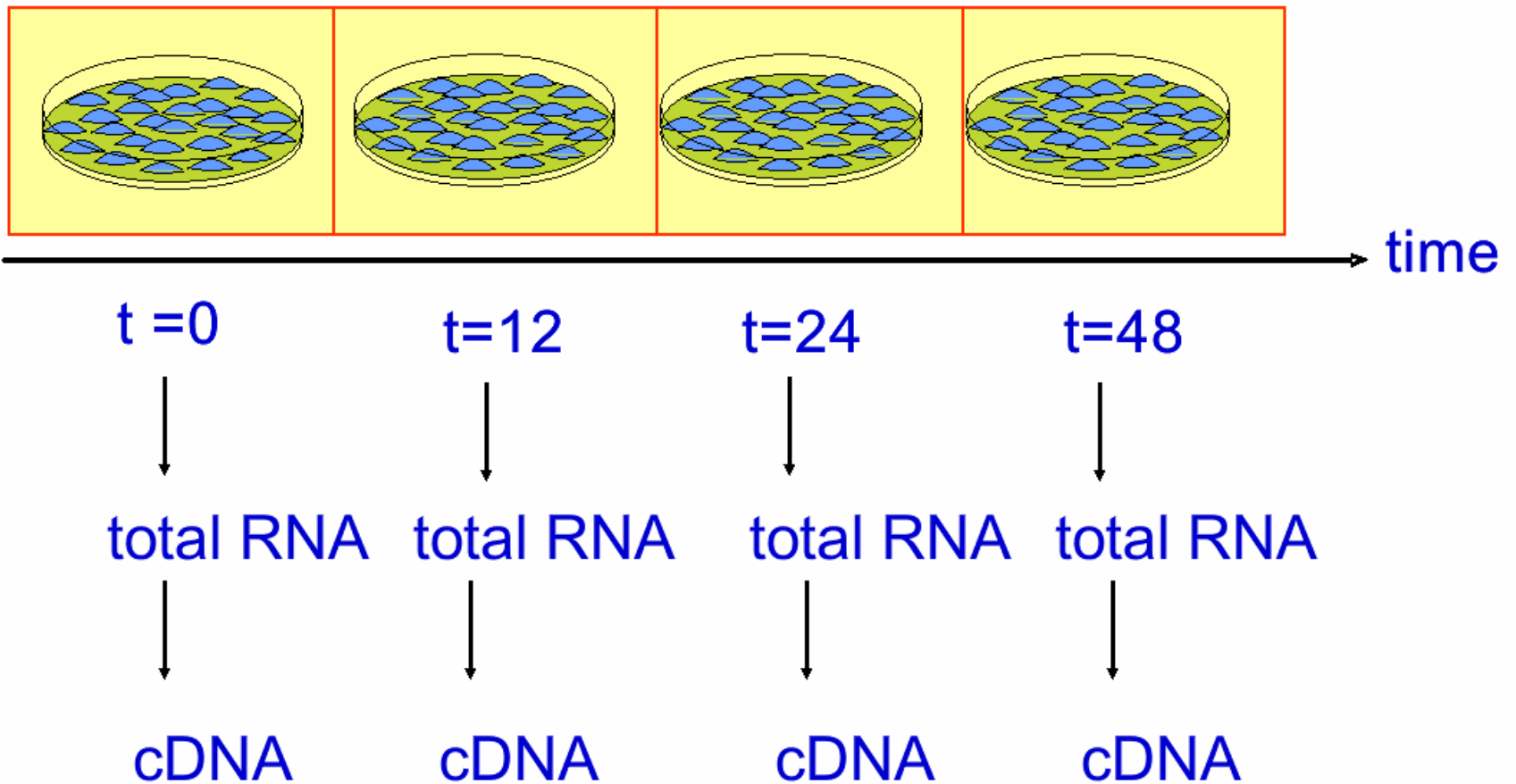
Relative Quantitation

Calibrator = The sample used as the basis for comparative results

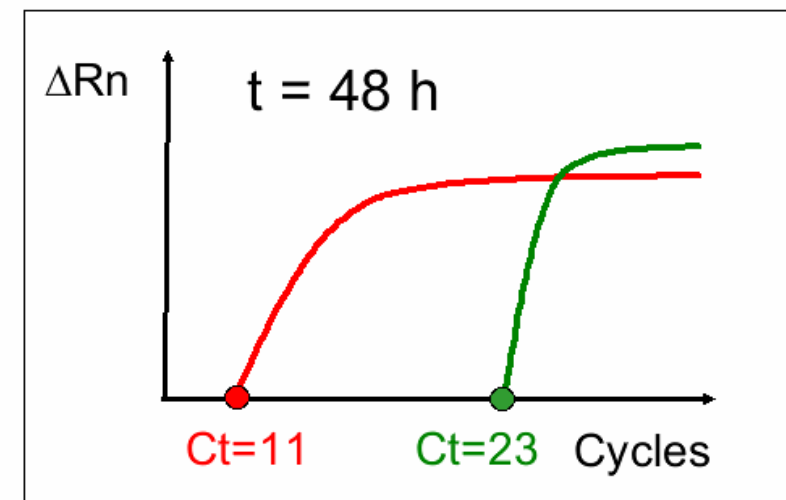
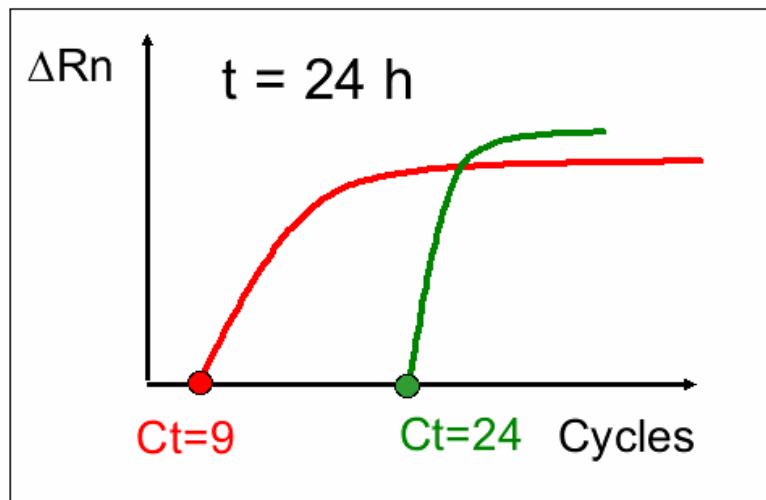
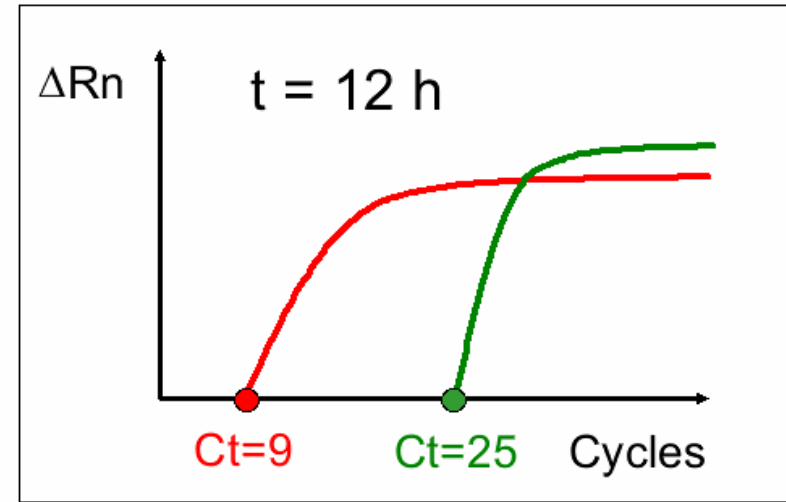
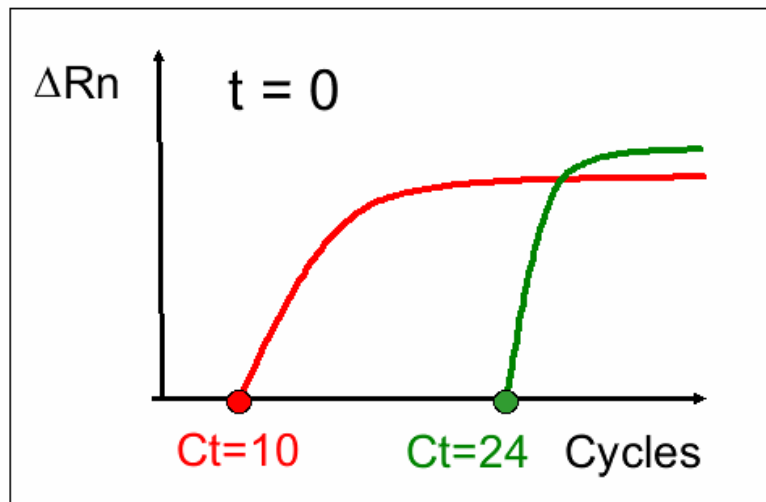


Relative Quantitation

Endogenous Control = Target used to normalize for sample handling
(e.g. 18S rRNA, GAPDH, β -actin)



Comparative C_T Method



■ Endogenous control ■ Target gene

Comparative C_T Method Calculation:

Normalized to endogenous control:

$$C_{T\ 48hrs} \text{ (23)} - C_{T\ Endo.\ Control} \text{ (11)} = \Delta C_{T\ 48hrs} \text{ (12)}$$

$$C_{T\ 0hrs, Calibrator} \text{ (24)} - C_{T\ Endo.\ Control} \text{ (10)} = \Delta C_{T\ 0hrs} \text{ (14)}$$

Normalized to calibrator sample:

$$\Delta C_{T\ 48hrs} \text{ (12)} - \Delta C_{T\ 0hrs, Calibrator} \text{ (14)} = \Delta\Delta C_{T} \text{ (-2)}$$

Relative fold change:

$$2^{(-\Delta\Delta C_T)} = 2^{\text{(2)}} = \text{(4)}$$

There is a 4-fold overexpression of my gene at T=48h compared T=0h...48hrs after drug treatment!

Applications

◆ Real-Time Detection

- Absolute Quantitation
- Relative Quantitation

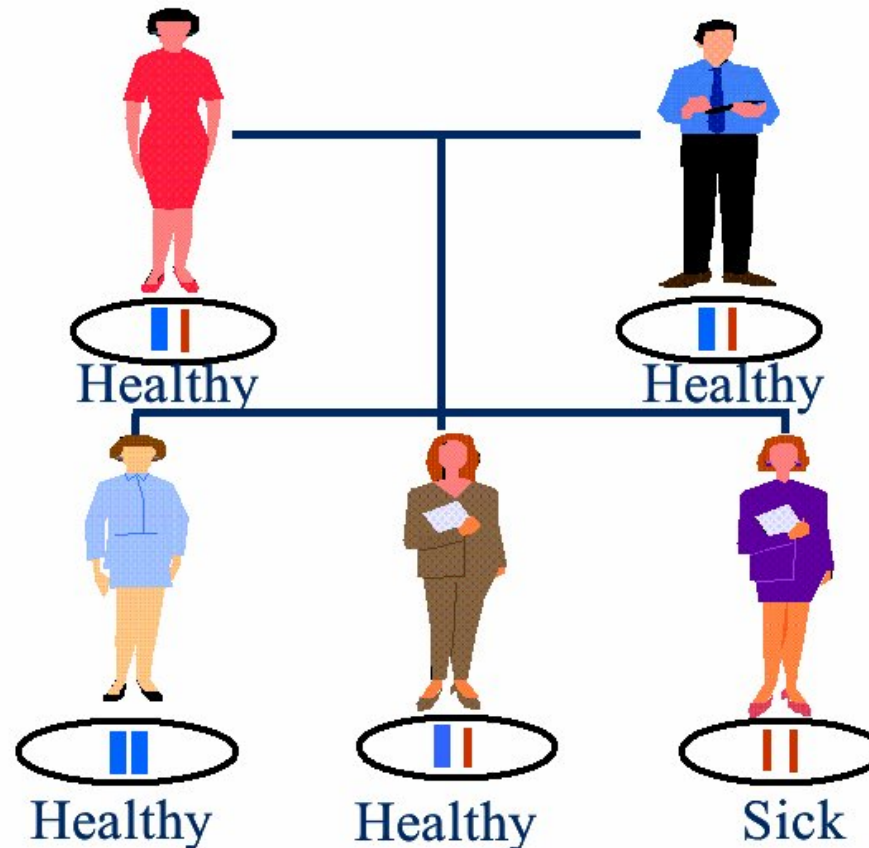
◆ End Point Detection

- Allelic Discrimination (SNP)
- +/- Assay (IPC)
 - Pathogen Detection

Allelic Discrimination (SNP)

◆ Determines the genotype of samples

- Possible to differentiate a single nucleotide polymorphism (SNP)

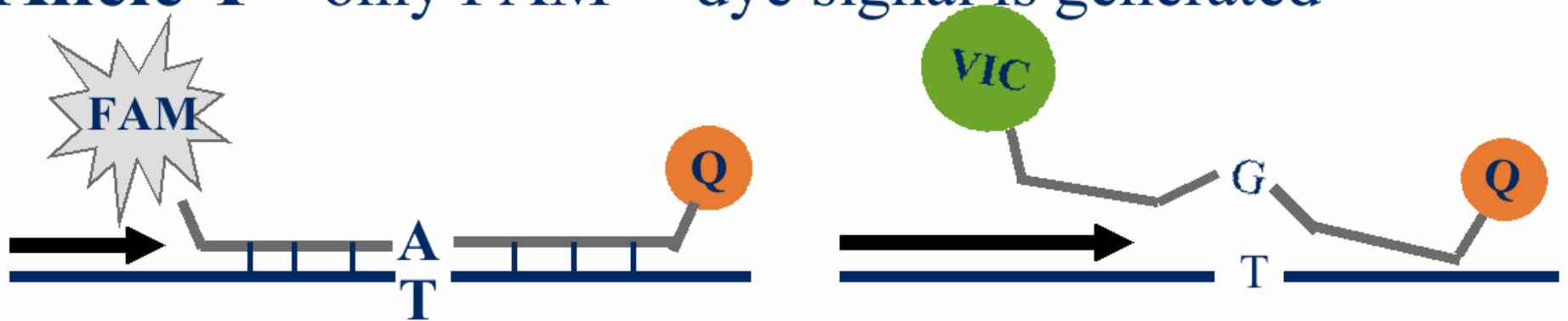


Allelic Discrimination Assay

Allele C - only VIC^R dye signal is generated



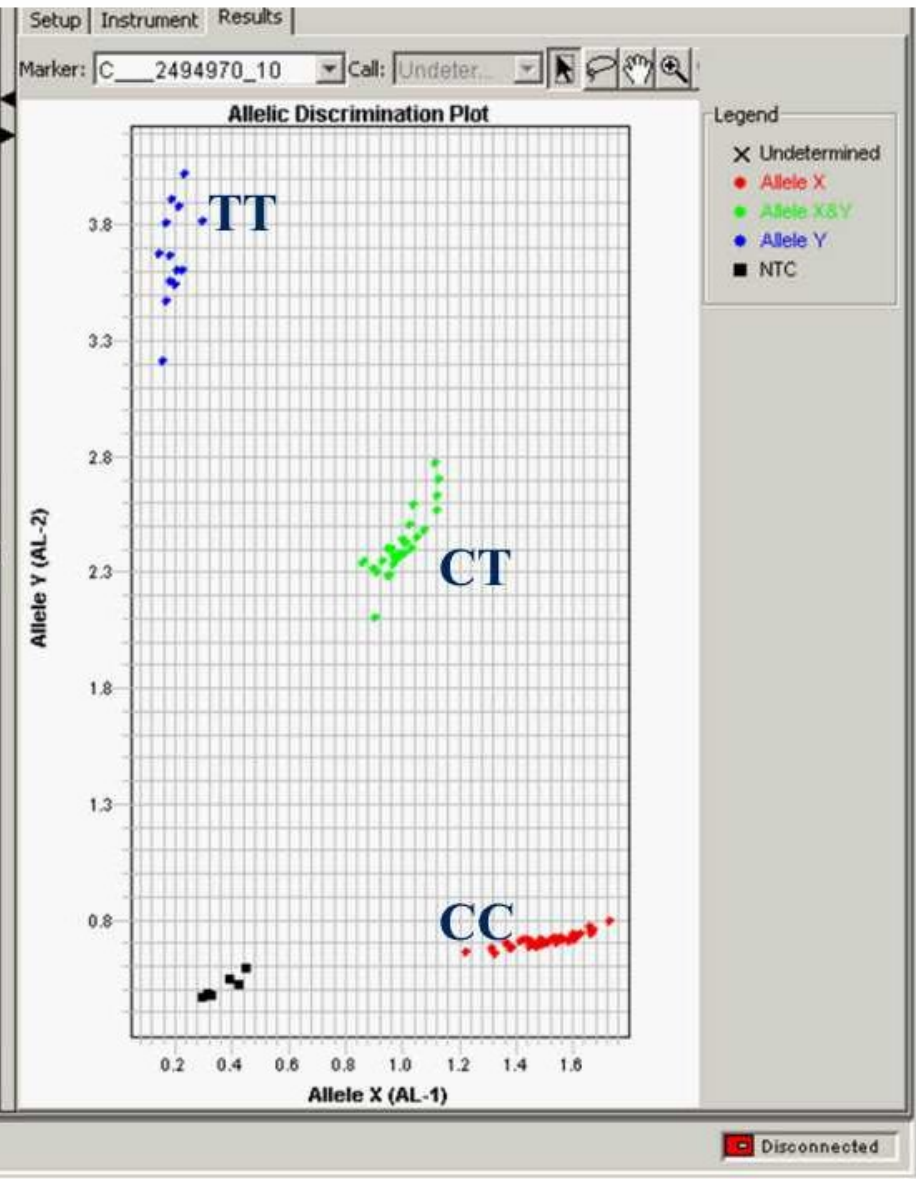
Allele T - only FAMTM dye signal is generated



Allelic Discrimination (SNP)

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23
A	NT	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
B	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	PO	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
C	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	PO	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
D	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
E	NT	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
F	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	PO	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
G	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	PO	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
H	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
I	NT	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
J	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	PO	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
K	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	PO	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
L	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA

	Allele X Rn	Allele Y Rn	Passive Ref R	Task	Call	Quality Value	Call Type	?
0	1.4346231222	7.2357E-1	4.07411743E3	Unknown	AL-1	99.83	Automatic	
0	1.436563015	7.1887E-1	4.07931543E3	Unknown	AL-1	99.91	Automatic	
0	1.4762319326	7.1261E-1	4.14455469E3	Unknown	AL-1	99.8	Automatic	
0	9.3739861E-1	2.3027508	3.80921924E3	Unknown	Both	99.85	Automatic	
0	1.4520388842	7.1169E-1	4.23892529E3	Unknown	AL-1	99.87	Automatic	
0	1.5078164339	7.354E-1	3.97991406E3	Unknown	AL-1	99.96	Automatic	
0	1.5383040905	7.4058E-1	4.03561401E3	Unknown	AL-1	99.94	Automatic	
0	9.6396738E-1	2.3712568	3.74589673E3	Unknown	Both	99.95	Automatic	
0	1.4599691629	6.9937E-1	4.06696484E3	Unknown	AL-1	99.32	Automatic	
0	1.4714711905	7.1601E-1	3.99868652E3	Unknown	AL-1	99.91	Automatic	
0	9.3969476E-1	2.3025954	3.78599658E3	Unknown	Both	99.85	Automatic	
0	9.7082245E-1	2.4008629	3.80715747E3	Unknown	Both	99.98	Automatic	
0	9.9780667E-1	2.4470868	3.74816064E3	Unknown	Both	99.96	Automatic	
0	1.0166915655	2.4187698	3.74147754E3	Unknown	Both	99.78	Automatic	
0	1.0388317108	2.4720457	3.79221973E3	Unknown	Both	99.71	Automatic	
0	1.9021322E-1	3.5632982	3.58489087E3	Unknown	AL-2	99.9	Automatic	
0	5.448506E-1	2.3890722	3.78411694E3	Unknown	Both	99.96	Automatic	



Applications

◆ Real Time Detection

- Absolute Quantitation
- Relative Quantitation

◆ End Point Detection

- Allele Detection (SNP)
- +/- Assay (IPC)
 - Pathogen Detection

Internal Positive Control (IPC)

- ◆ Distinguish true target negative from PCR inhibition
- ◆ Co-amplified with target DNA without compromising amplification of the target sequence

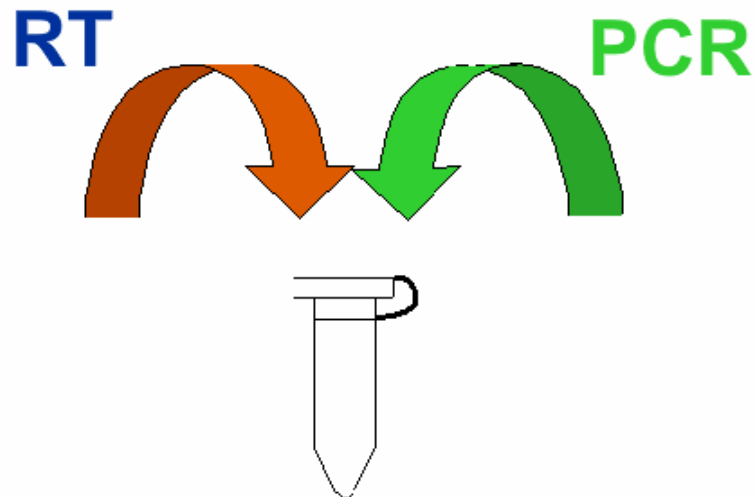
Plus/Minus assay with IPC

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

Disconnected NUM

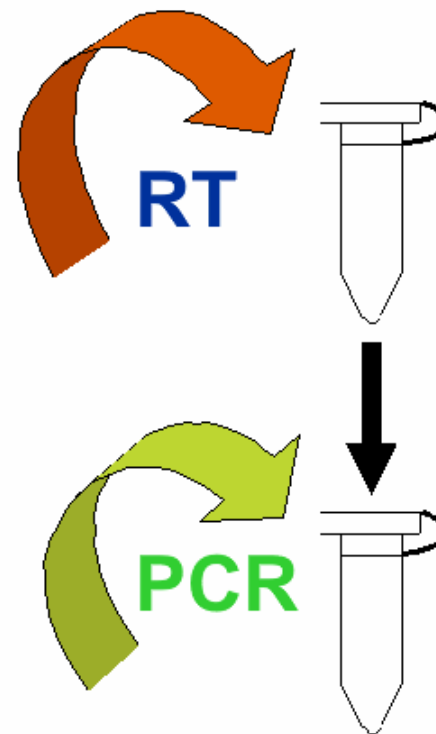
RNA to Amplified cDNA: 1-Step vs. 2-Step

1-STEP



- Closed tube
(no contamination)
- Easy-to-use

2-STEP

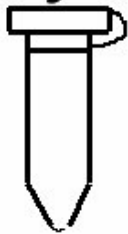


- Archive-ready
sample

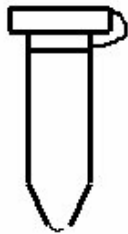
Formats: Master Mix vs. Core Reagent

Core reagents allow flexibility and optimization

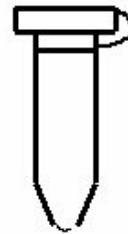
AmpliTaq Gold[®]
DNA Polymerase



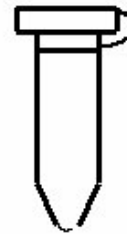
AmpErase[®]
UNG



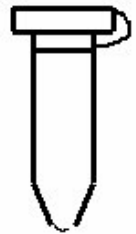
10X Buffer



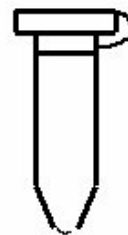
dNTPs



MgCl₂



Master mixes are easy-to-use and convenient



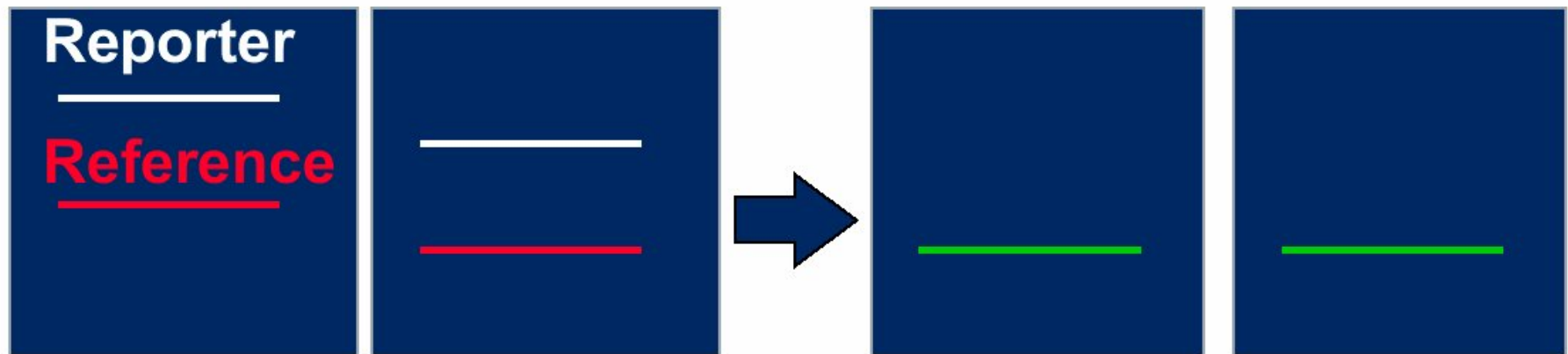
All components in one tube!

Advantage of Using a ROX™ Dye Normalizer

Improves precision

Compensates for small fluorescent fluctuations that can occur from well-to-well

Reporter / Reference



please wait for Q&A



Thank you

Mohamed Abdel Fattah

Senior Technical Specialist

Scientific Support - Molecular Biology Dept.

AnalysisAB Co.

Mobile: 012 27 906 74

E-mail: analysis@analysis-ab.com

