

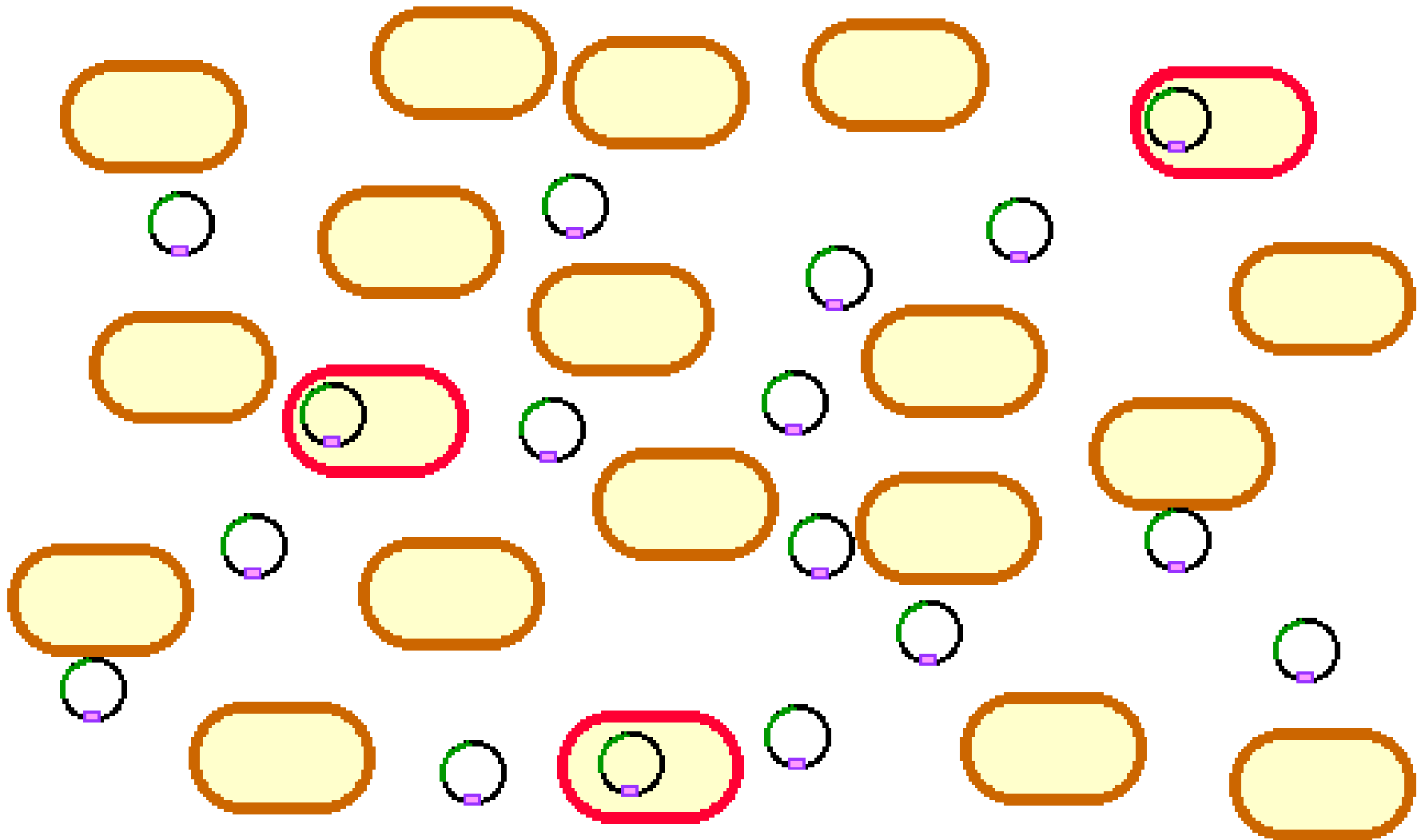
DETECTION OF YOUR CLONING AND PICK UP YOUR RIGHT CLONE

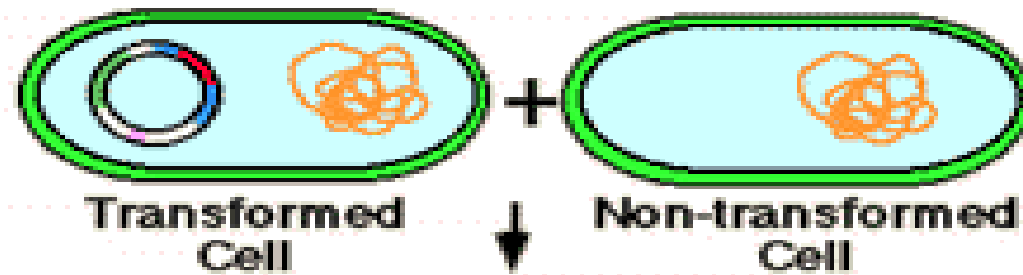
SELECTION OF THE
CELLS WITH GENE OF
INTEREST

Introduction

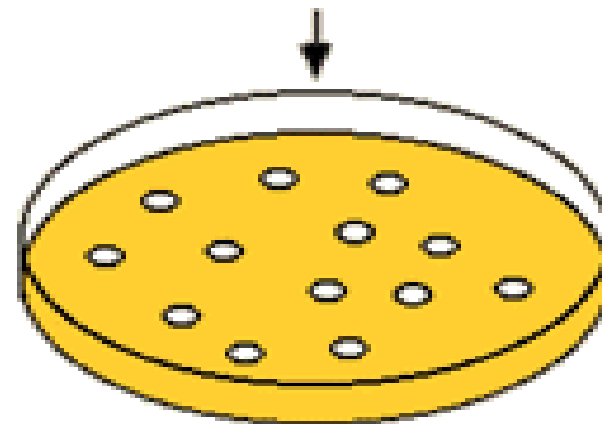
- Once recombinant plasmid is constructed → Introduced into recipient cells.
- Introduction of recombinant DNA into recipient cells is called transformation: *introduction of foreign DNA changes (transforms) properties of the organism.*
- Special treatment makes *cells competent* - capable of accepting foreign DNA.
- Usually, these treatments make cell membrane more permeable for a DNA molecule.
- When competent cells are mixed with DNA some cells (actually, very few) become transformed.

Competent cells transformation





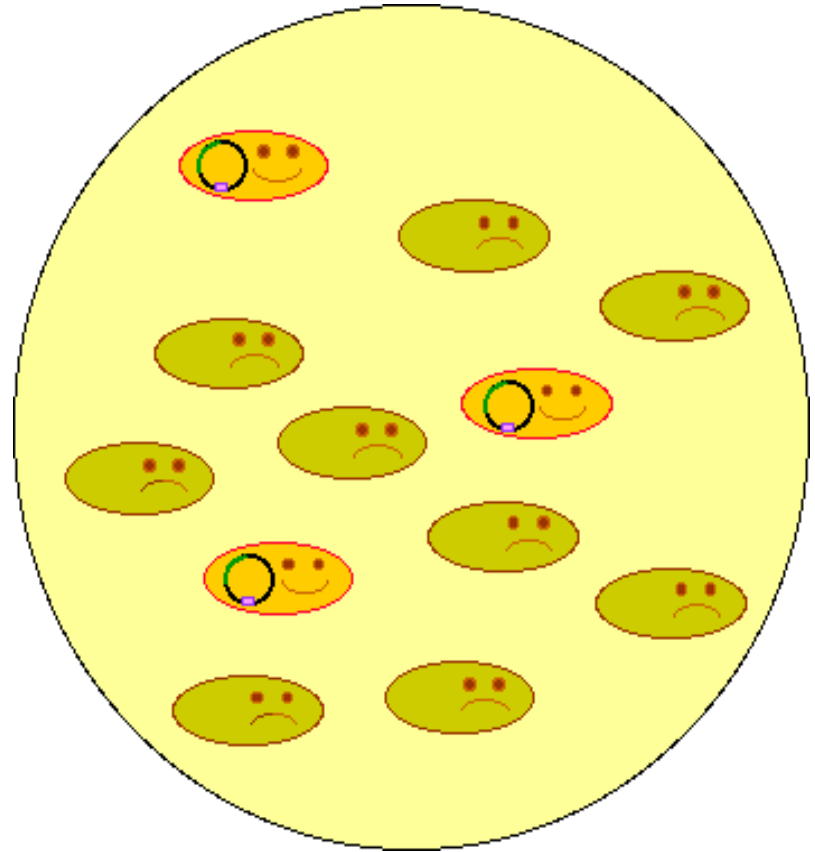
Overnight Growth



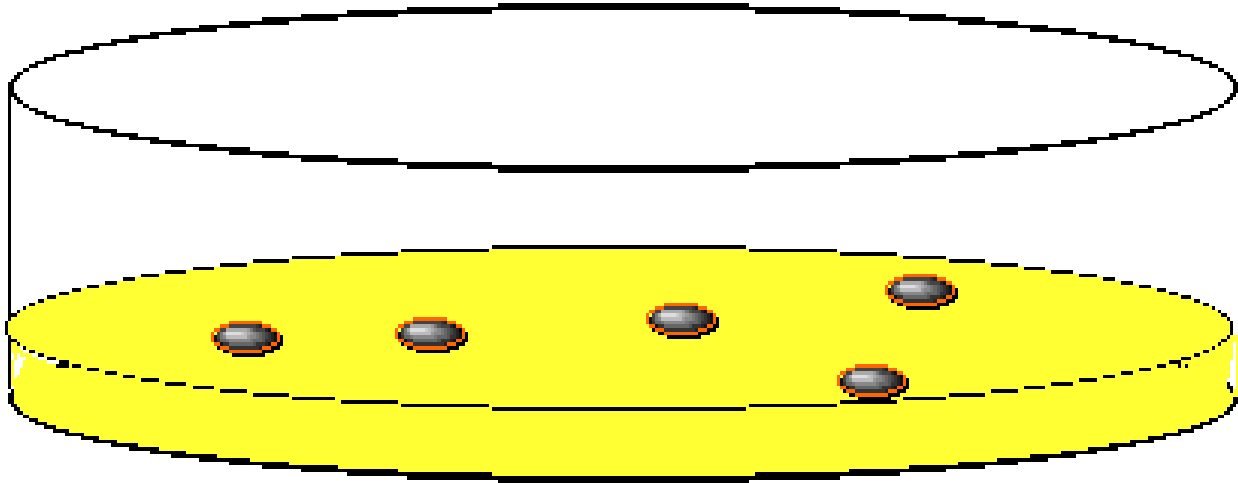
Competent cells culture:

After transformation, cells are plated onto agar medium that contains selective antibiotic:

Only transformed cells, will survive and form colonies. All the untransformed cells will die.

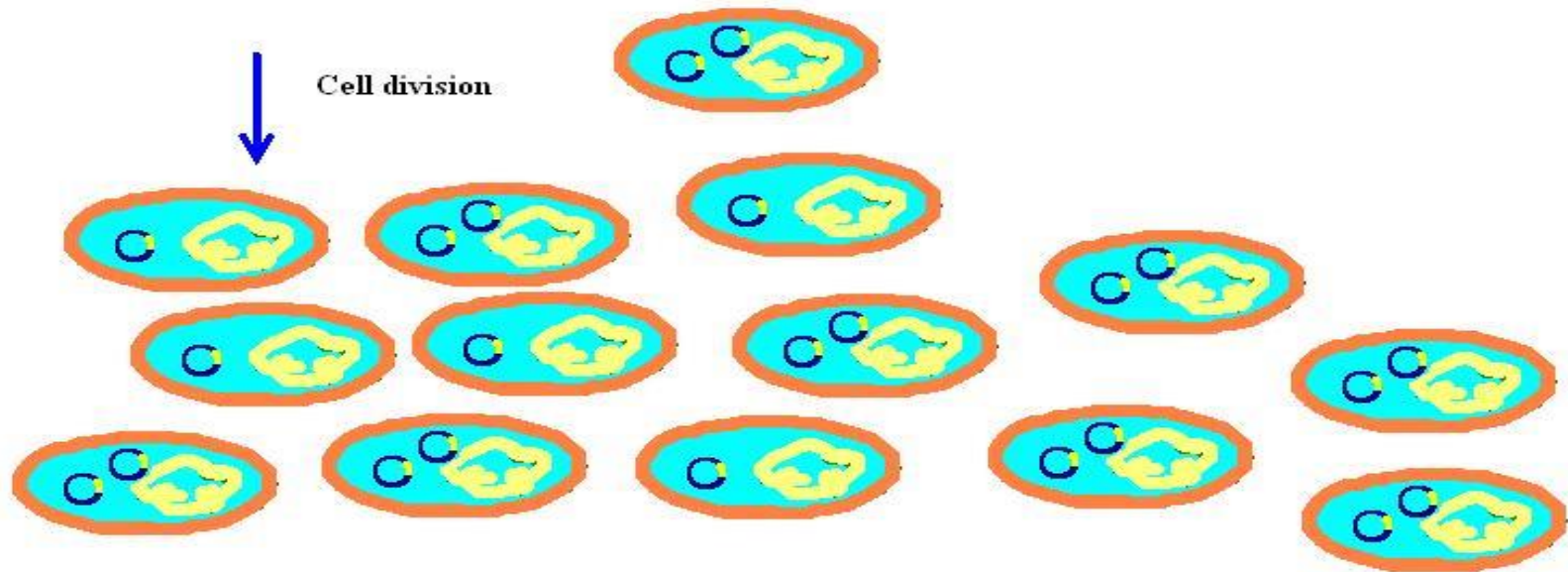
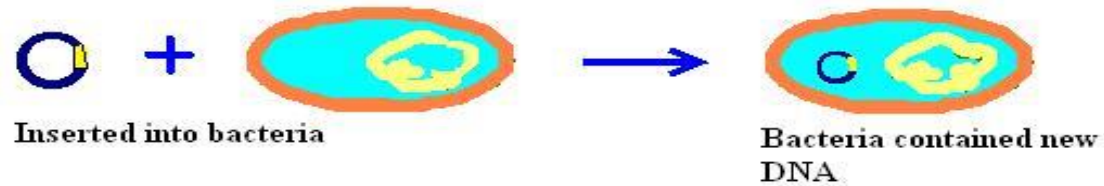


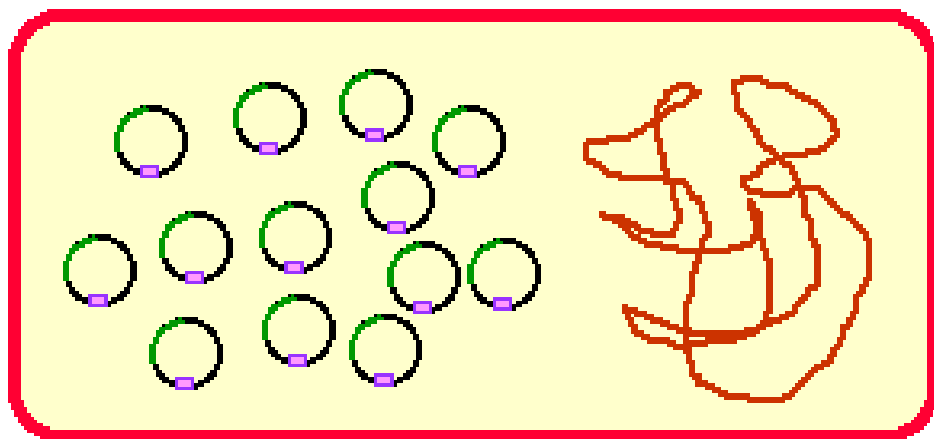
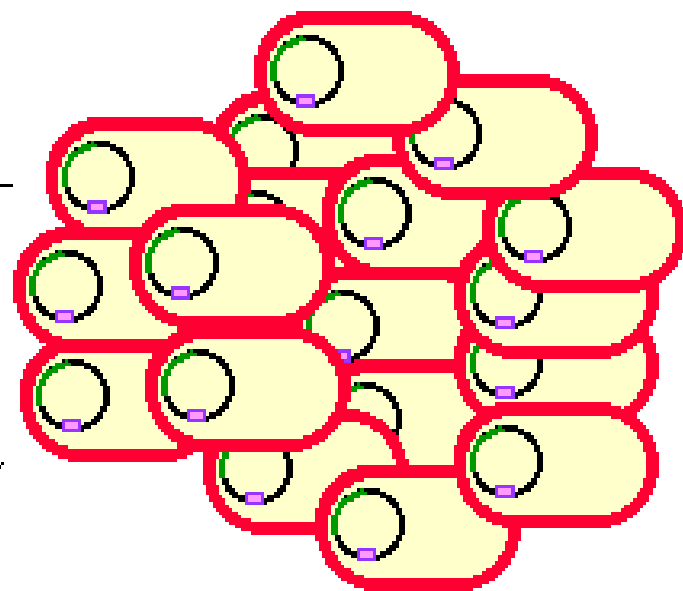
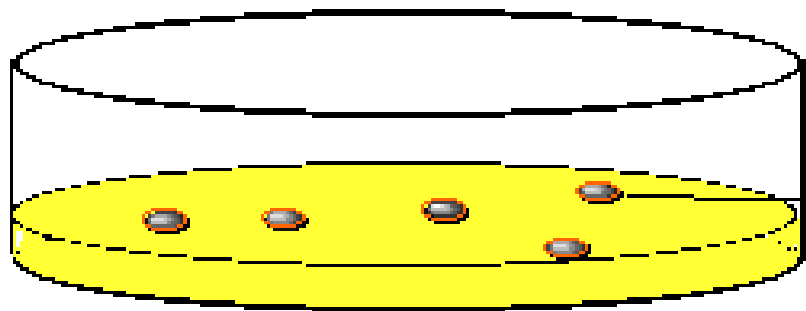
In each colony formed on the agar plate, all cells are descendants of one transformed cell.



agar medium with antibiotic

All cells in the clone are genetically identical and contain the same recombinant vector





How to Find and Pick Up the Right Clone?

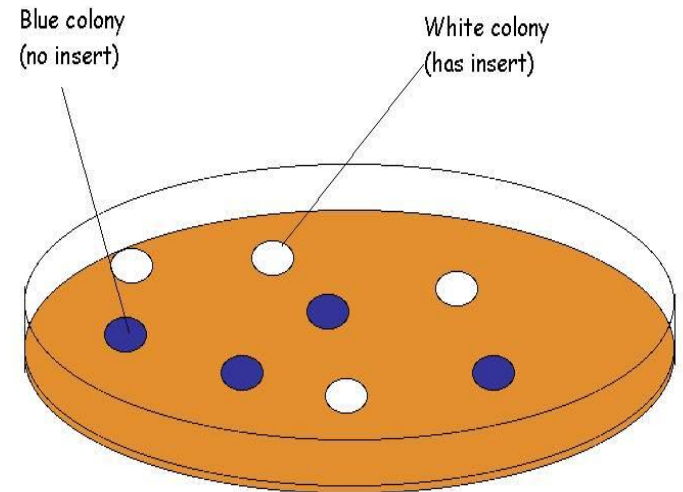
The most common methods include:

- 1. Phenotypic screening.**
- 2. Screening with antibodies.**
- 3. DNA hybridization.**

Phenotypic screening

Phenotypic screening is used when cloned gene is expressed and changes properties of the cell in an "obvious way".

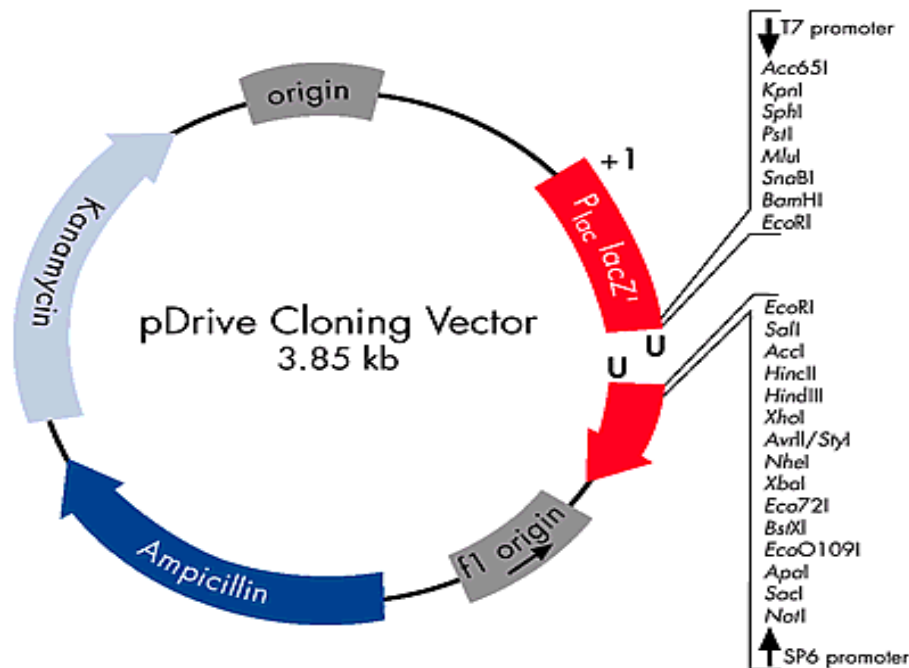
E.g. **Blue-White Screening**
Insertional mutagenesis



Blue-White Screening

LacZ α \rightarrow Provide β -galactosidase activity.

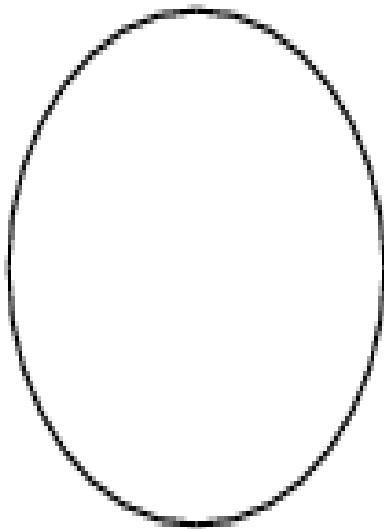
X-gal is a colorless analog of lactose cleaved by β -galactosidase to form a bright blue insoluble pigment.



- No insert \rightarrow intact lacZ \rightarrow Blue
- Insert \rightarrow disrupted lacZ \rightarrow White

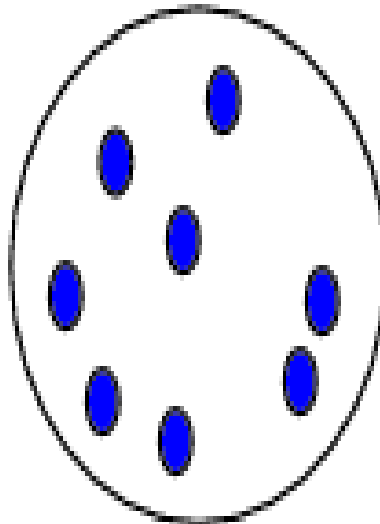
BLUE – WHITE SCREENING

A)



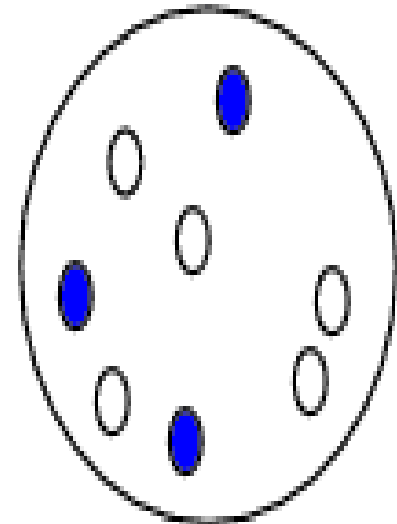
Agar plate
+ ampicillin
+bacteria
+no plasmid

B)



Agar plate
+ampicillin
+bacteria
+plasmid

C)



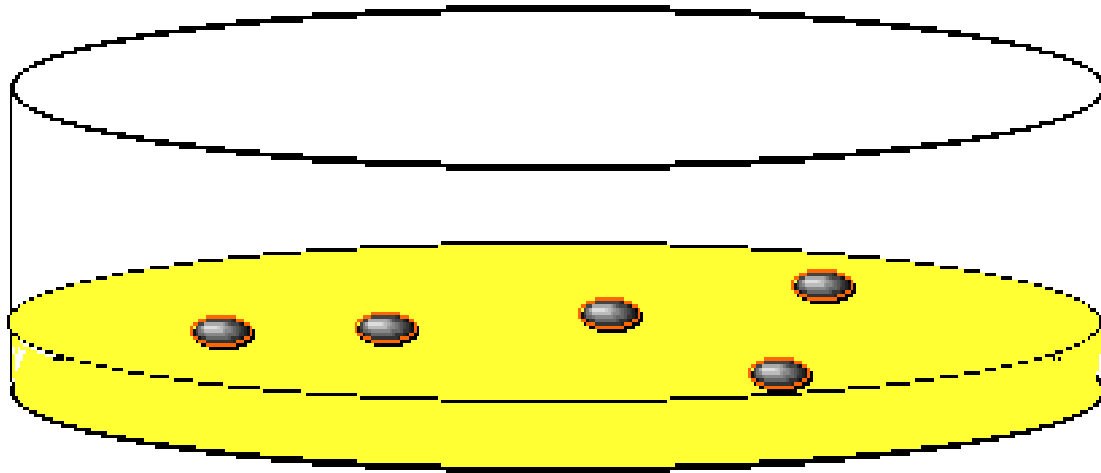
Agar plate +
ampicillin +
bacteria +
recombinant plasmid

BLUE – WHITE SCREENING



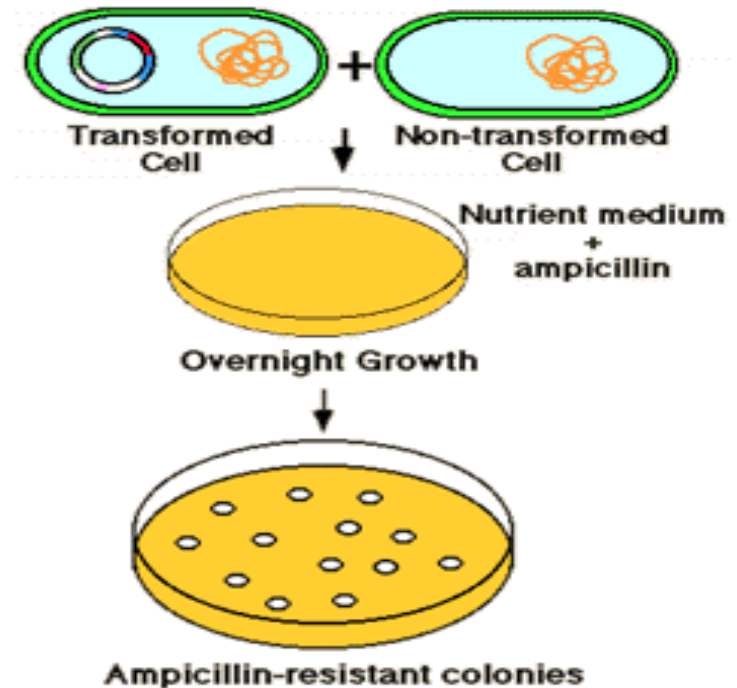
Screening with antibodies

→ (Antibiotic Resistance).



agar medium with antibiotic

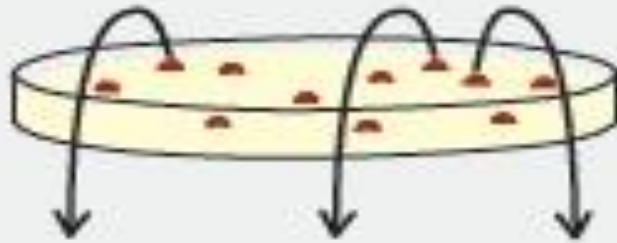
Plasmid vector contains an ampicillin resistance gene making the cell resistant to ampicillin containing medium. Growth of transformed cells (cells receiving the plasmid) can be identified on agar medium containing (e.g.) ampicillin. Thus, the cells with recombinant vector can be selected. This is an direct selection procedure.



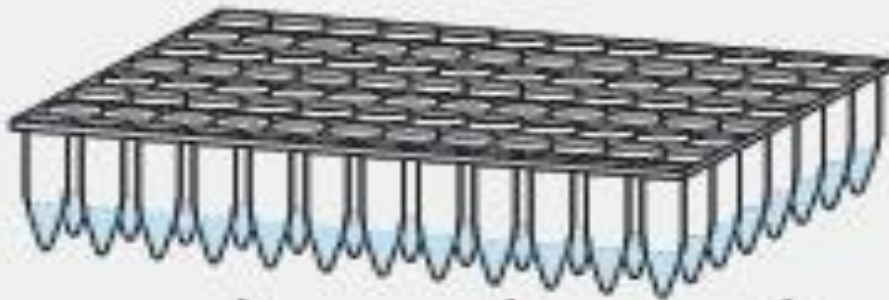
Confirmation

By using PCR:

Specific primer for the
Cloning Vector and the DNA
insert

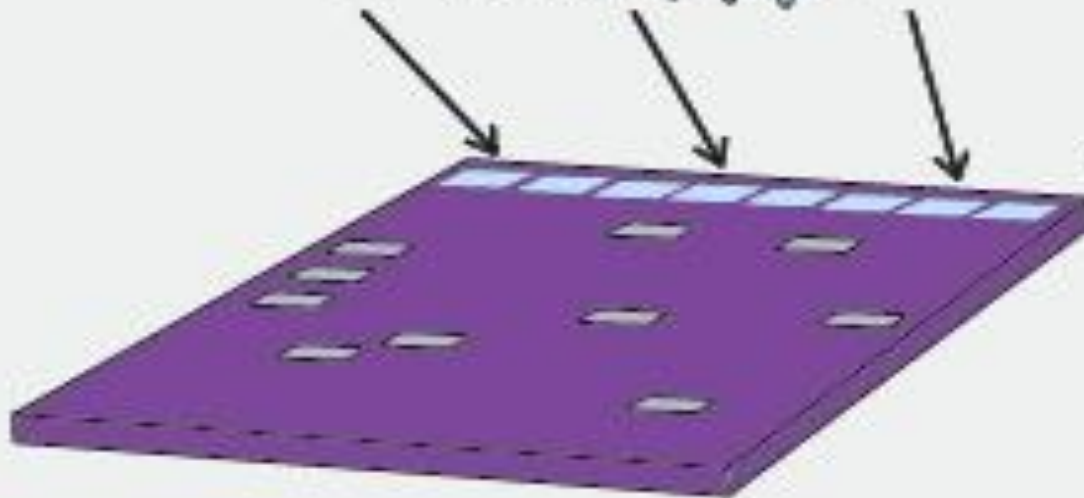


1. Pick colony into a microcentrifuge tube or microtiter well.



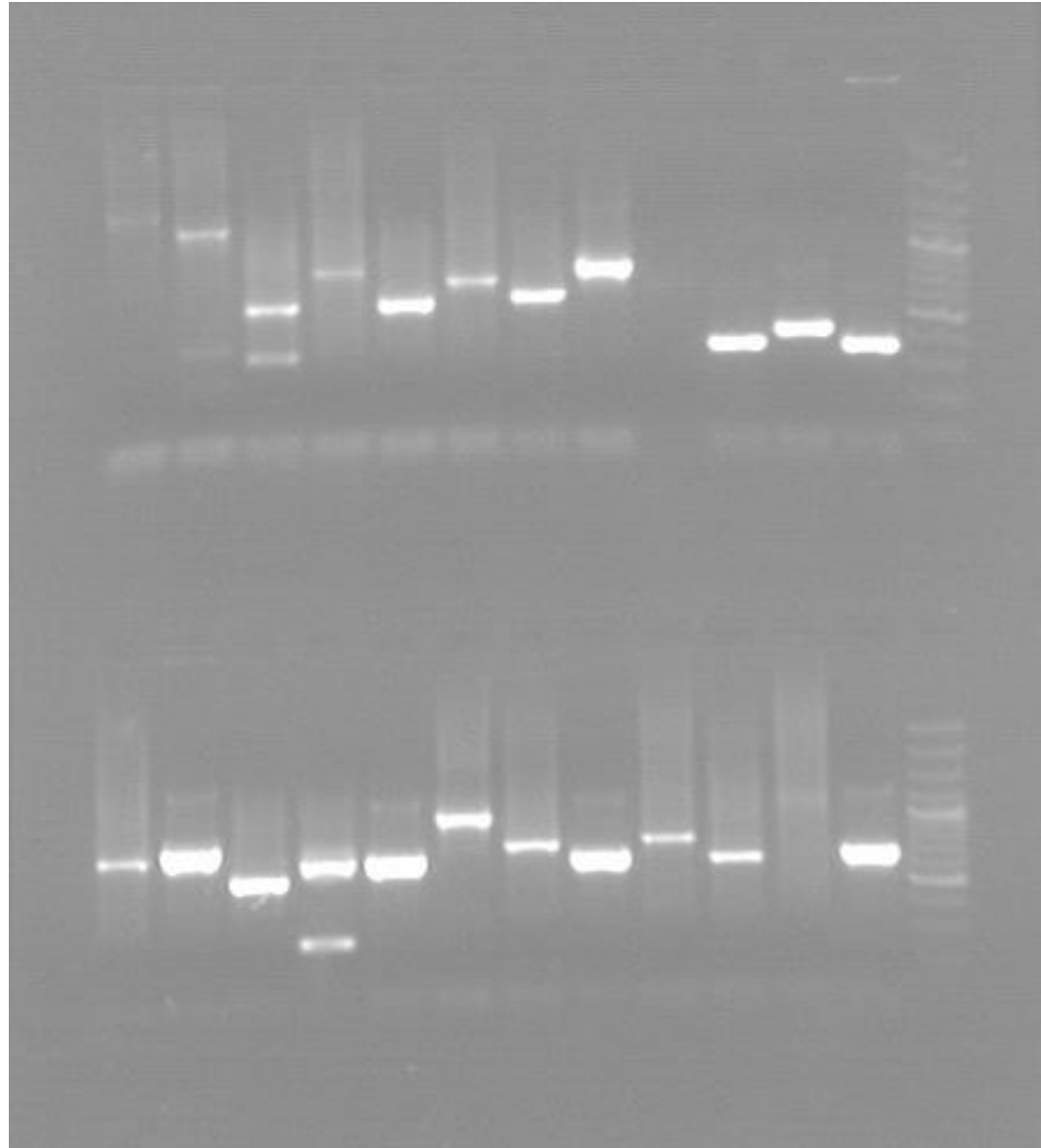
2. Add PCRLyse™ Solution, vortex, heat 5 min at 99°C.

3. Perform PCR using an aliquot of the lysed cells.

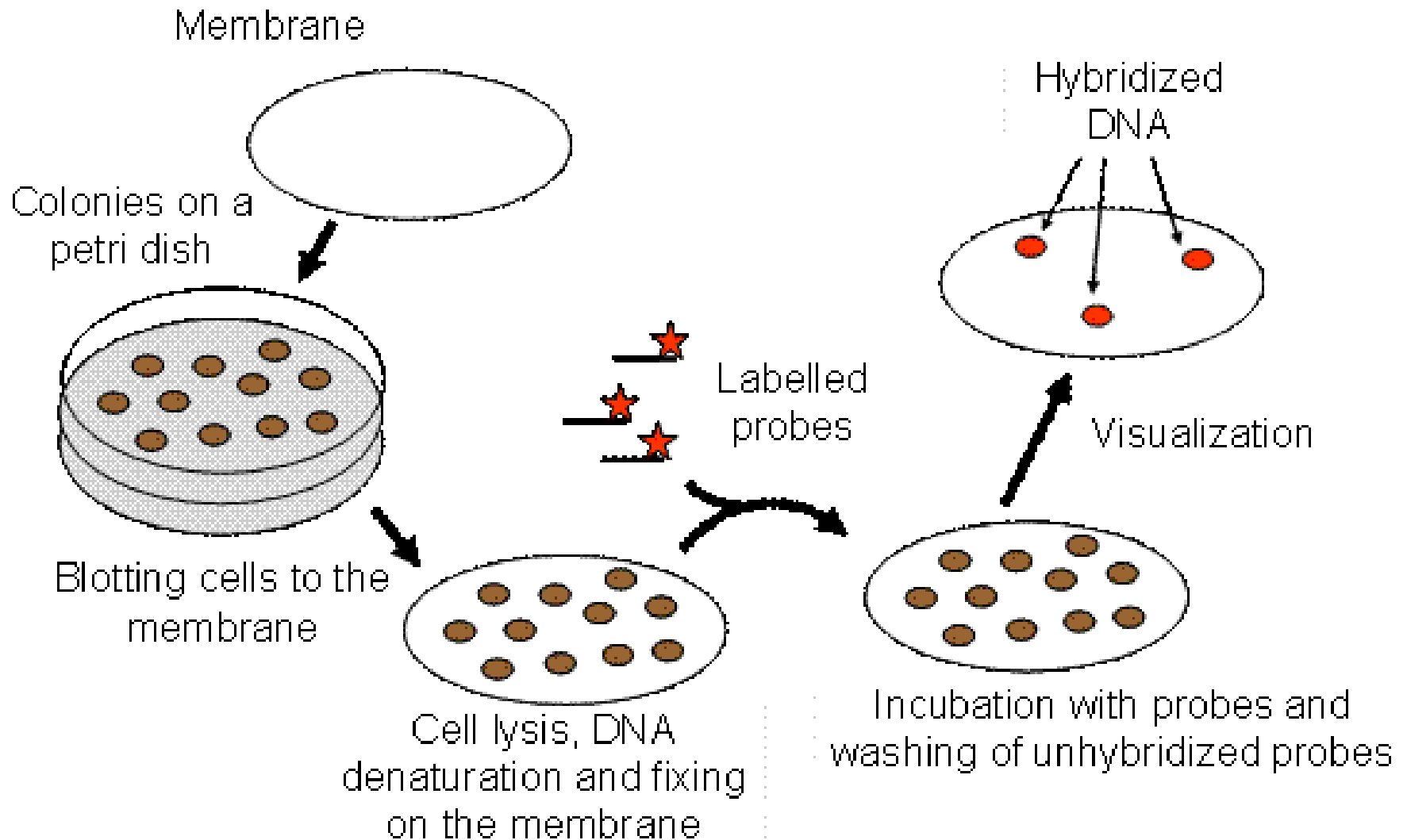


4. Analyze by gel electrophoresis.

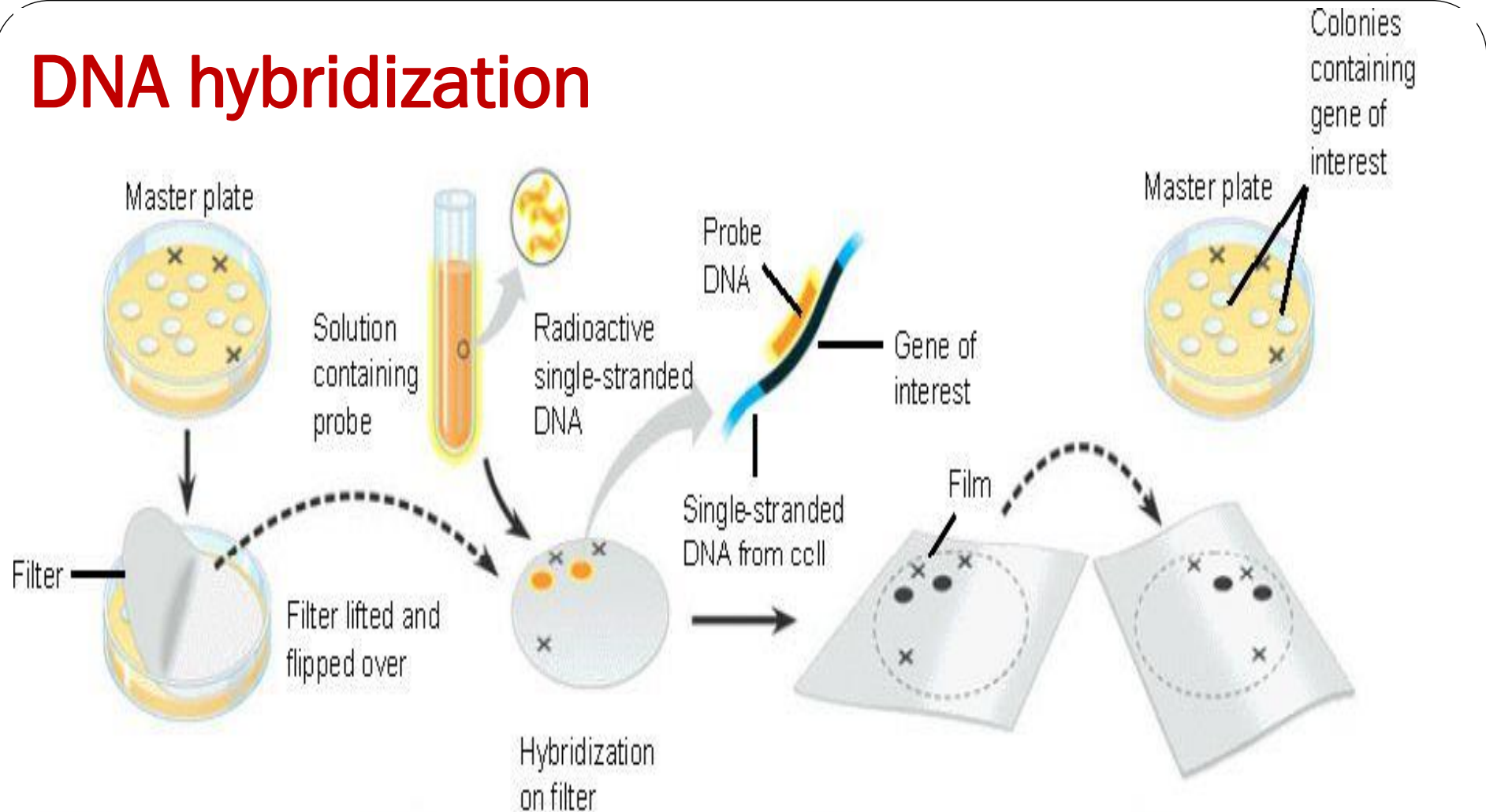
PCR for detection of the vector which carry the insert:



DNA hybridization

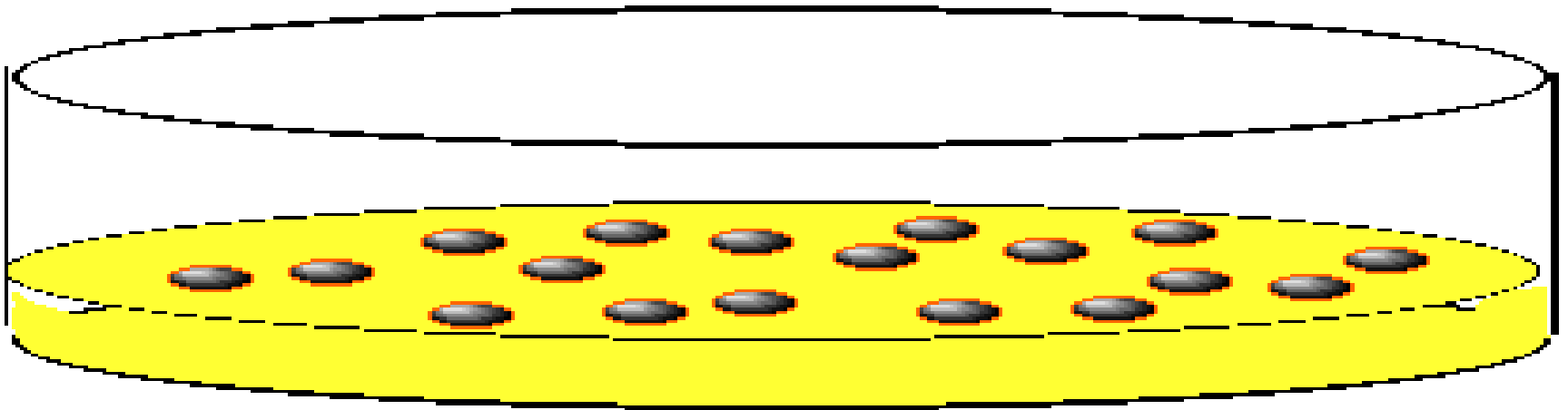


DNA hybridization

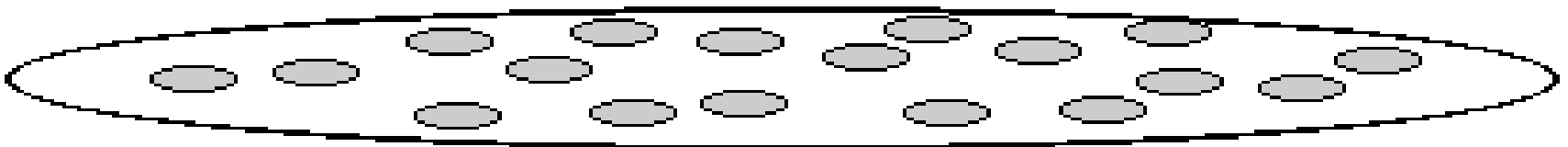


- 1 A special filter paper is pressed against the master plate, transferring cells to the bottom side of the filter.
- 2 The filter is treated to break open the cells and denature their DNA; the resulting single-stranded DNA molecules are treated so that they stick to the filter.
- 3 The filter is laid under photographic film, allowing any radioactive areas to expose the film (autoradiography).
- 4 After the developed film is flipped over, the reference marks on the film and master plate are aligned to locate colonies carrying the gene of interest.

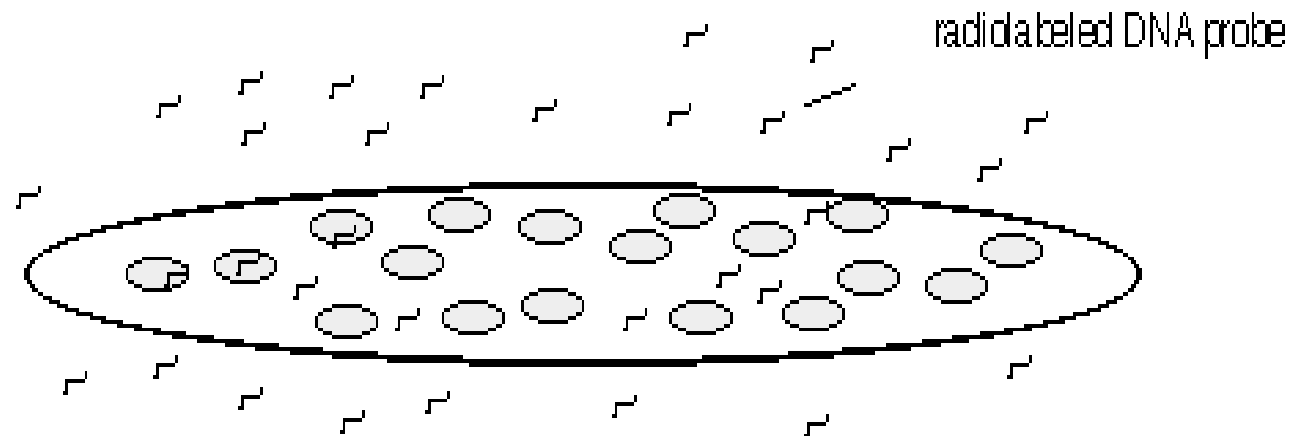
DNA hybridization



TRANSFER COLONIES TO
A NYLON FILTER

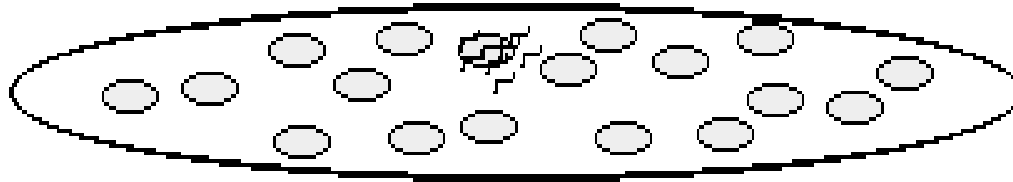


DNA hybridization

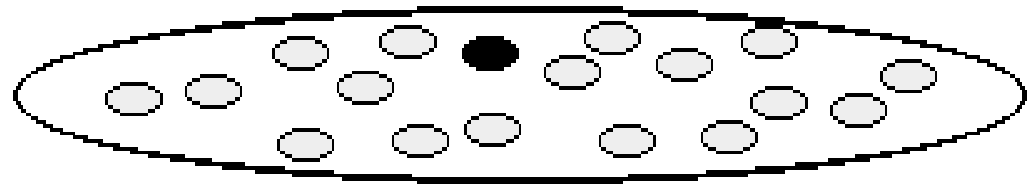


1. Synthesize a DNA fragment complementary to any strand of "our" gene.
2. Radiolabel the probe.
3. Lyse cells on the filter and denature DNA.
4. Hybridize the probe with the filter.
5. Wash out excess of the probe.

DNA hybridization



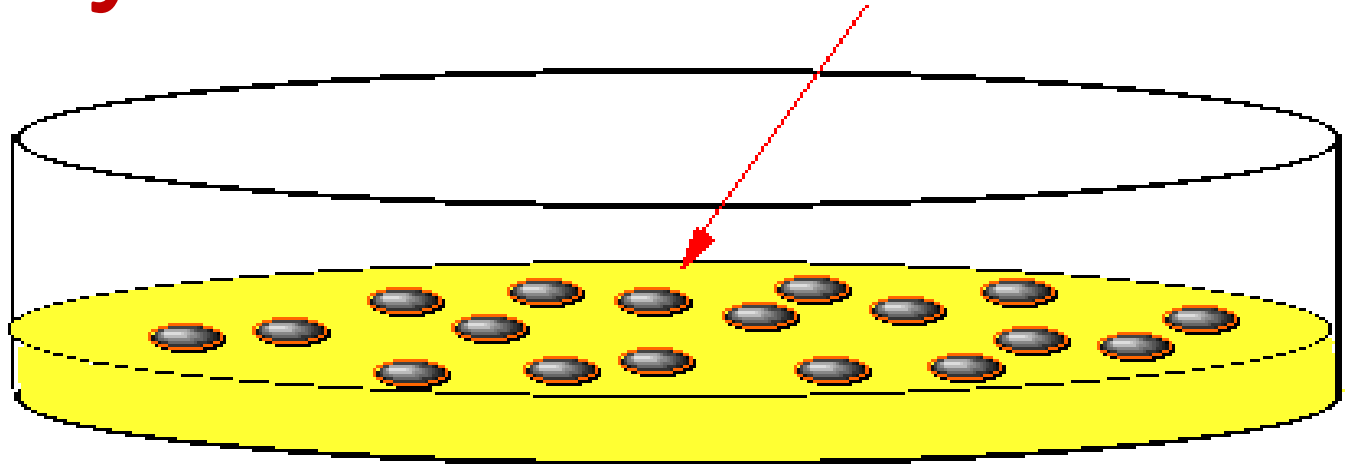
Only the colony that contains plasmids with “our gene” will hybridize with the probe and become radiolabeled.



Expose the filter to the X-ray film: a black spot will appear on the place of a colony with “our gene”.

DNA hybridization

Master plate



X-ray film



Find the colony on the master plate that contains the cloned gene.

Bacterial colonies containing cloned segments of foreign DNA

Radioactive DNA

1 Transfer cells to filter

Filter

2 Treat cells on filter to denature DNA

3 Add probe to filter

Solution containing probe

Probe DNA

Gene of interest

Single-stranded DNA from cell

Hybridization on filter

4 Autoradiography

Colonies containing gene of interest

Developed film

5 Compare autoradiograph with master plate

Master plate

DNA on membrane (a band "up close")

Low specific activity probe bound to DNA on membrane

weak signal on Southern

High specific activity probe bound to DNA on membrane

strong signal

DNA hybridization

If two single stranded DNA molecules have complementary nucleotide sequences they can **hybridize**: form a stable double stranded complex

3' CTGTCAGTCAGTCAGTCA 5'

+

5' GATCGTCGATTACCAAATGCAGTCAGACAGTCAGTCAGTGAGTCCAGTCCCCATTGAG



Incubate DNA fragments together

3' CTGTCAGTCAGTCAGTCA 5'

| | | | |

5' GATCGTCGATTACCAAATGCAGTCAGACAGTCAGTCAGTGAGTCCAGTCCCCATTGAG

unstable complex

3' CTGTCAGTCAGTCAGTCA

| | | | |

5' GATCGTCGATTACCAAATGCAGTCAGACAGTCAGTCAGTGAGTCCAGTCCCCATTGAG

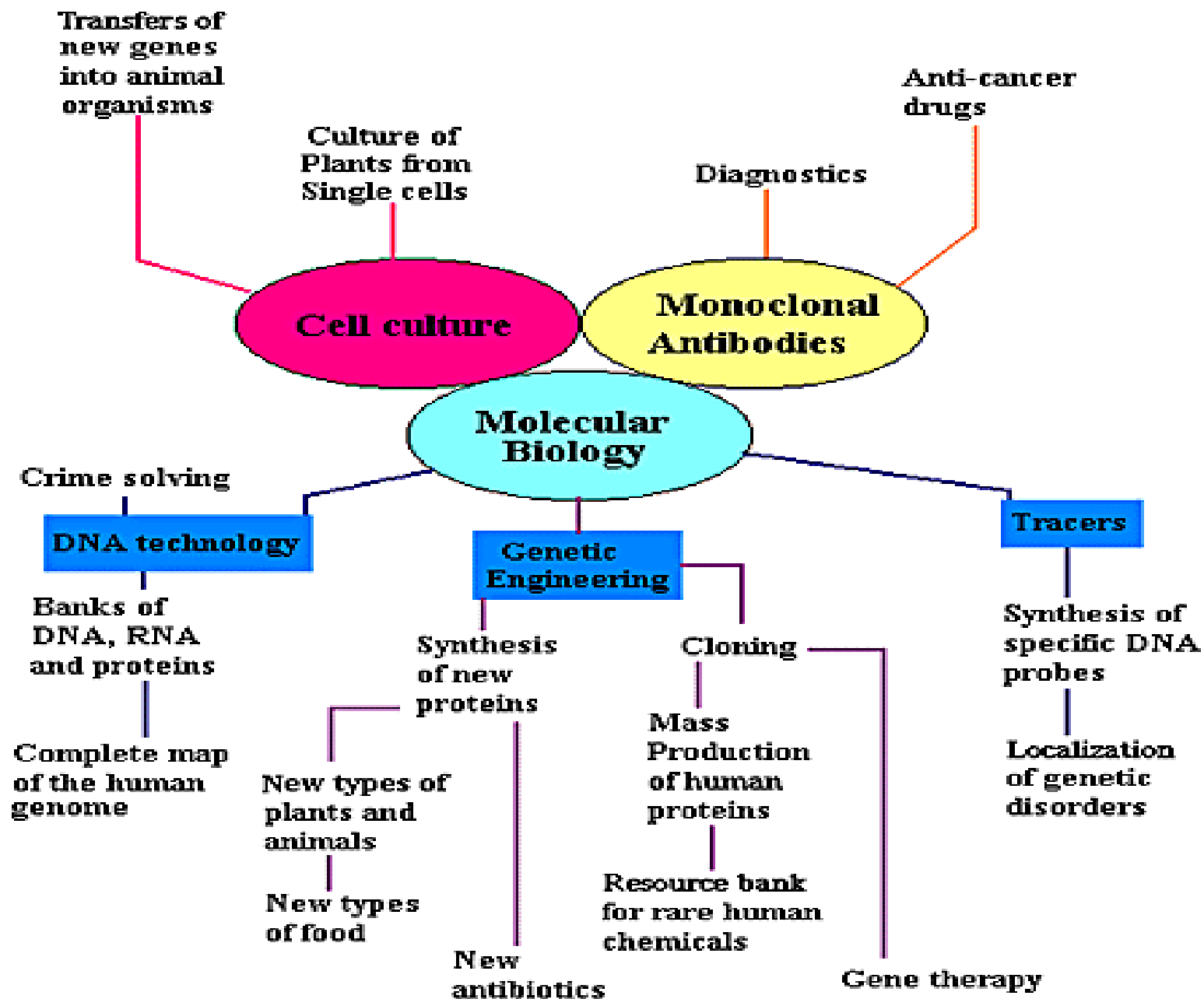
unstable complex

3' CTGTCAGTCAGTCAGTCA !


| | | | | | | | | | |

5' GATCGTCGATTACCAAATGCAGTCAGACAGTCAGTCAGTGAGTCCAGTCCCCATTGAG

stable complex



Good Friends
Never Say Goodbye
They Simply Say
"See You Soon"



Always laugh when you
can. It is cheaper than
medicine.

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Thanks a lot

with my Best Regards and My Best wishes

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