



GEL ELECTROPHORESIS

BY

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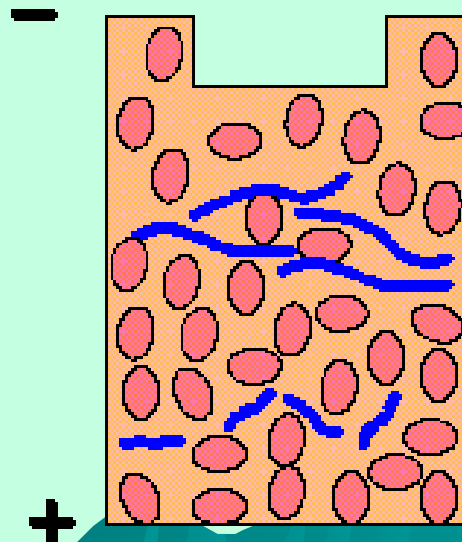
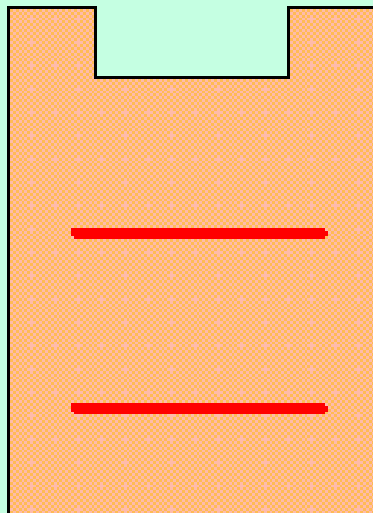
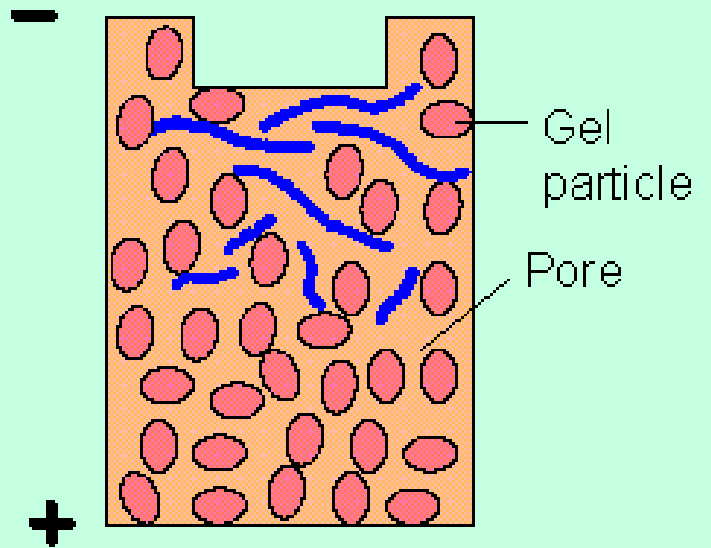
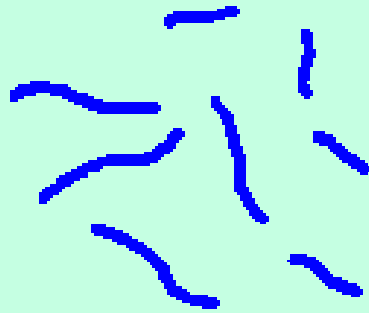
ELECTROPHORESIS:

Is the migration of charged molecules in solution in response to an electric field

The rate of migration depends on:

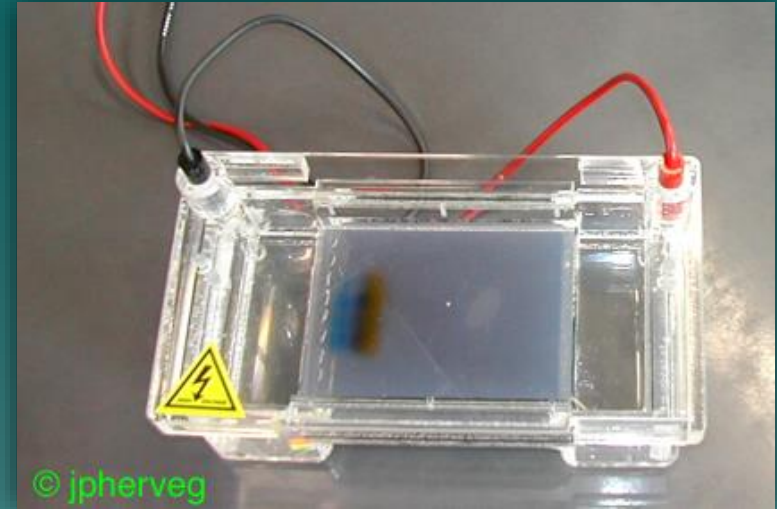
- ▣ Strength of the field
 - ▣ Net charge
 - ▣ Size & shape of the molecules
 - ▣ Ionic strength, viscosity & temperature of the medium
- (in which molecules are moving)

DNA fragments



AGAROSE

Is used to separate larger molecules such as nucleic acid, large proteins & protein complexes

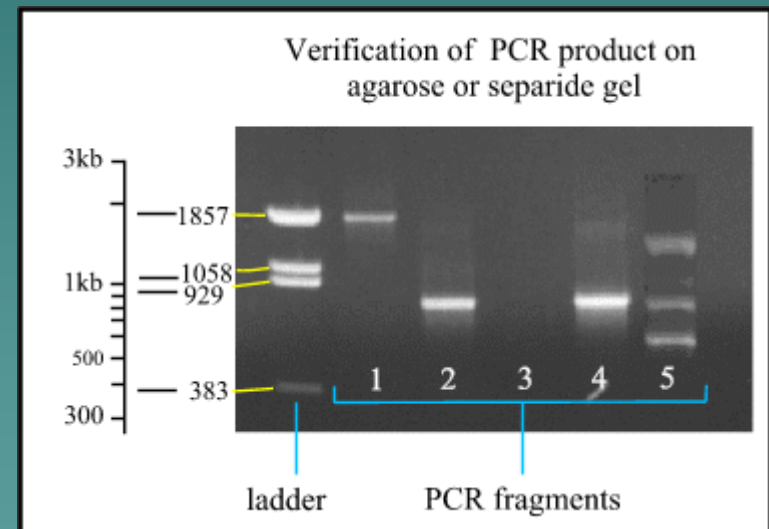
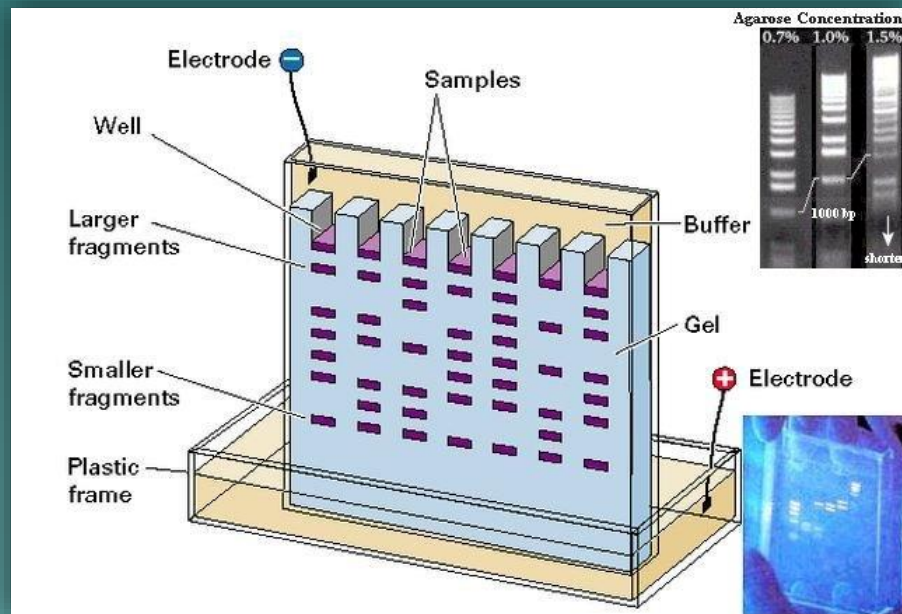


POLYACRYLAMIDE

Is used to separate most proteins & small oligonucleotides that require a small gel pore size for retardation



Dilute agarose gels are generally more rigid & easy to handle than polyacrylamide of the same concentration



There are 2 types of buffer systems in electrophoresis

Continuous System:

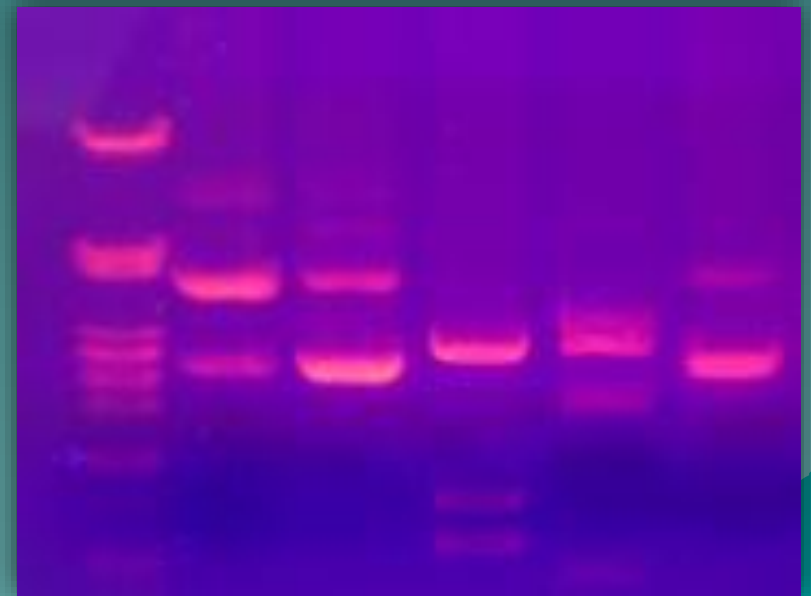
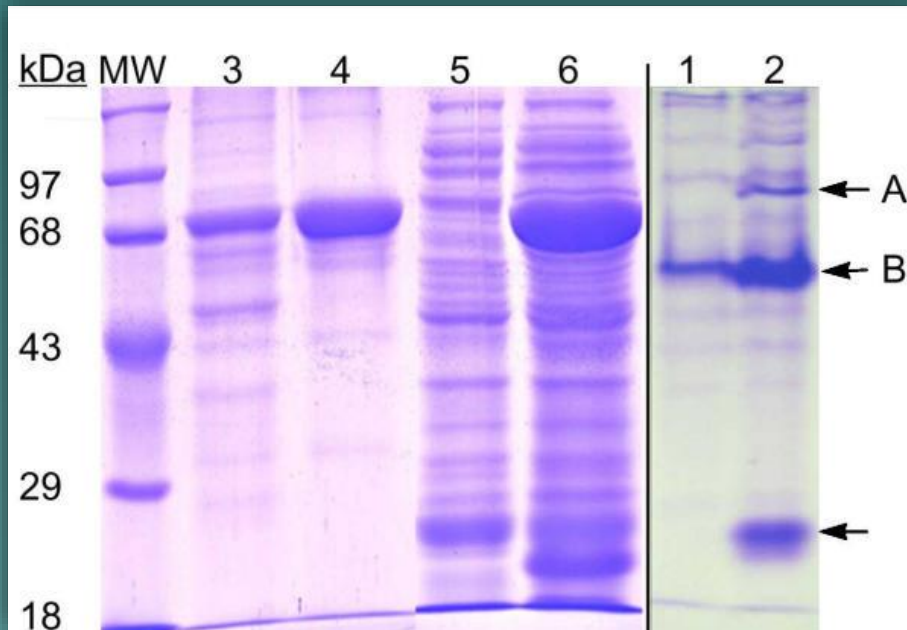
Has only a single separating gel & uses the same buffer in the tank & gel

Discontinuous System:

✓ A non-restrictive large pore gel (called stacking gel) is layered on top of a separating gel (called a resolving gel)

✓ Each gel is made with a different buffer, & the tank buffer are different from the gel buffers

THE RESOLUTION OBTAINED IN A DISCONTINUOUS SYSTEM IS **MUCH GREATER** THAN THAT OBTAINED WITH A CONTINUOUS SYSTEM



AGAROSE GEL ELECTROPHORESIS

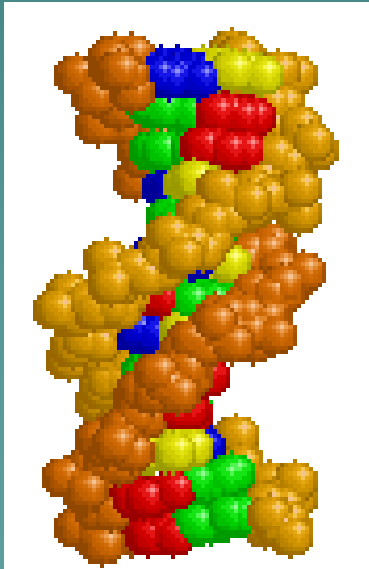
The standard method used to separate, identify & purify DNA fragments is **electrophoresis** through agarose gels

The technique is ➡ Simple

➡ Rapid to perform

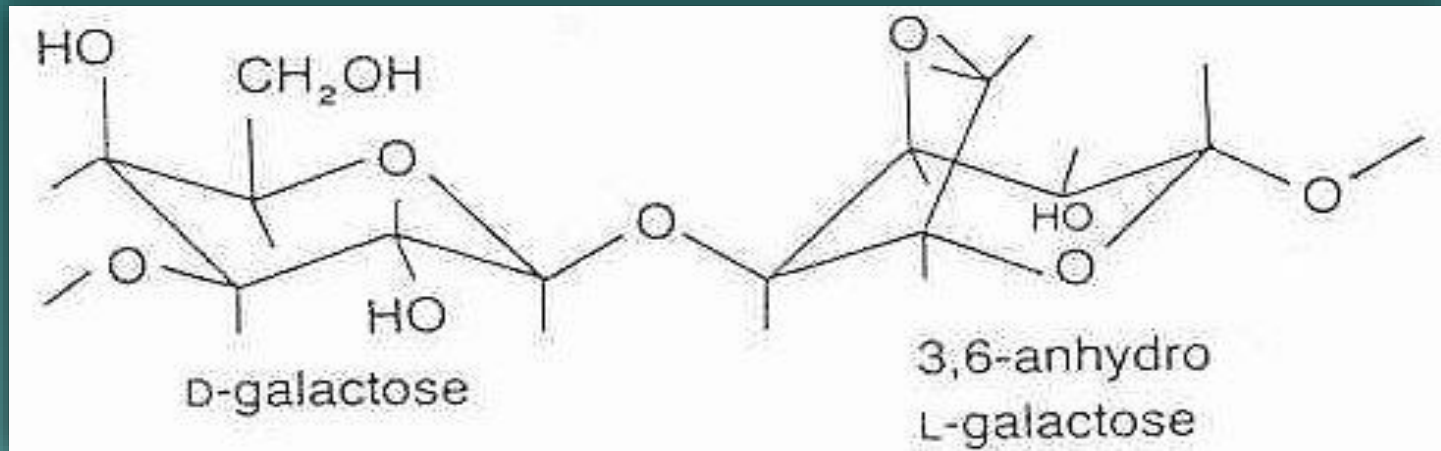
➡ Capable of resolving mixtures of DNA fragments

➡ The location of DNA within the gel can be determined directly



(AGAROSE)

Is extracted from seaweed & is a linear polymer whose basic structure is



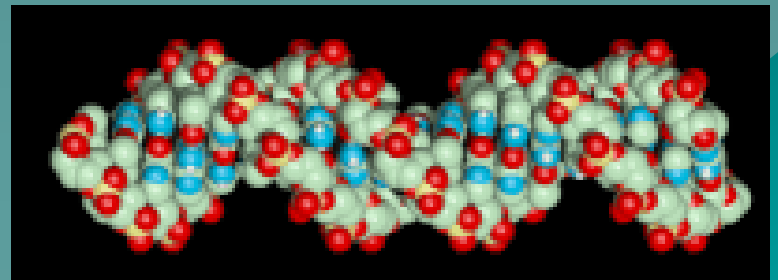
Commercially available agarose is not completely pure; it is contaminated with other polysaccharides, salts, & proteins

The amount of contamination varies from batch to batch & from manufacture to manufacture

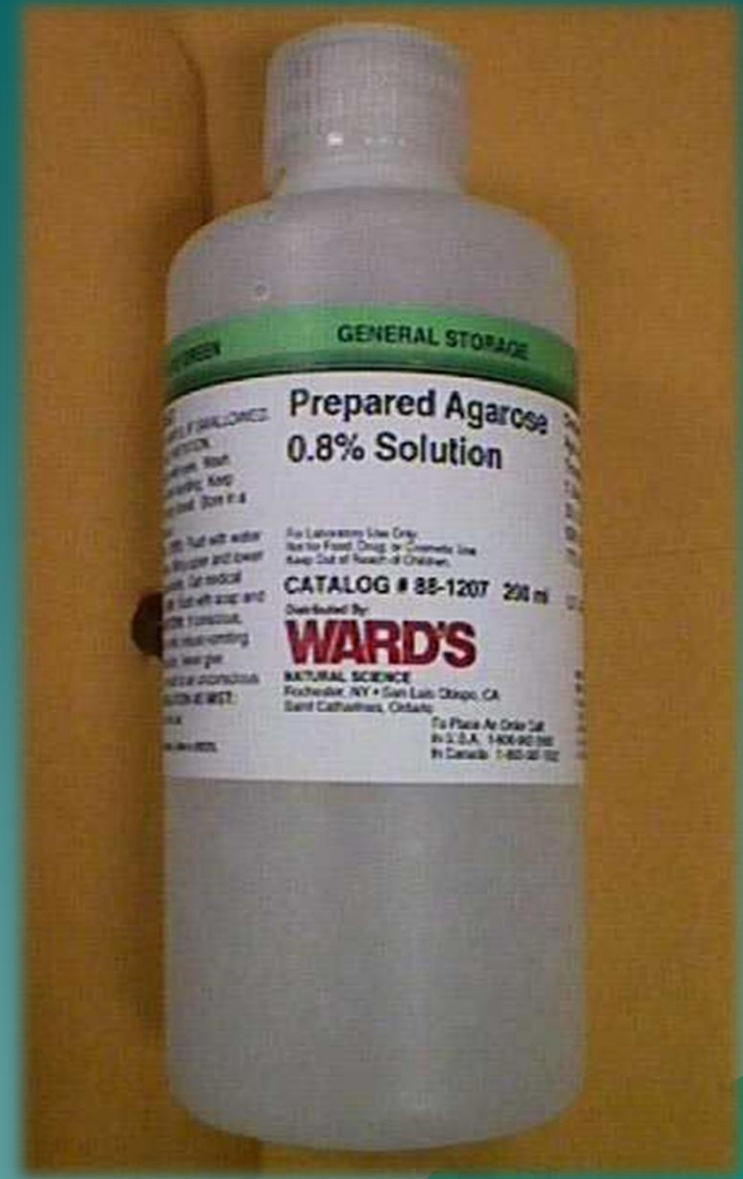
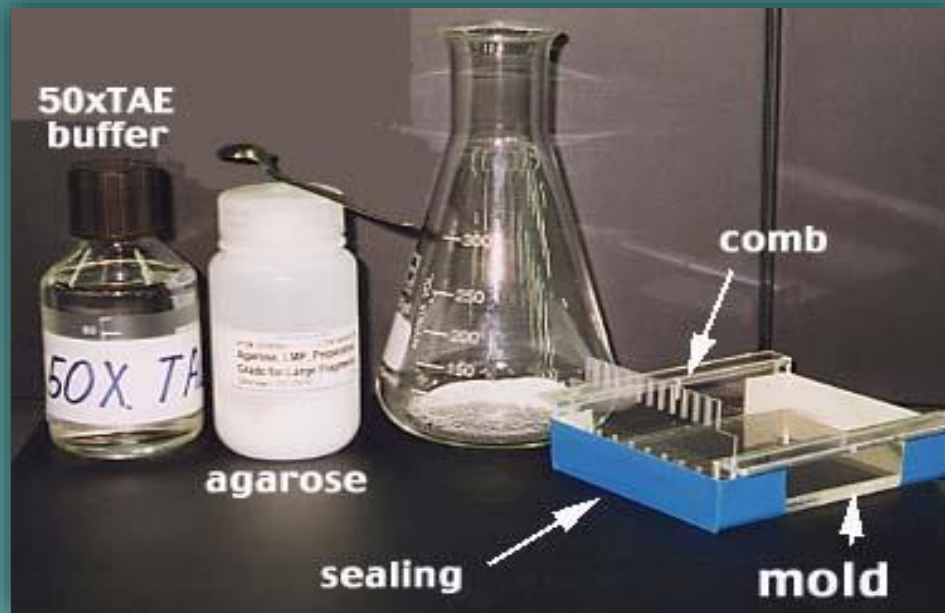
These difference can affect on

➡ Migration of the DNA

➡ Ability of the DNA recovered from gel

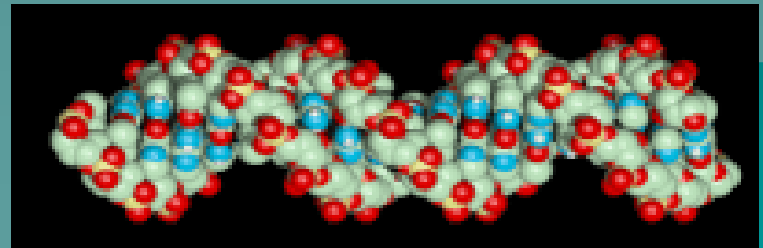


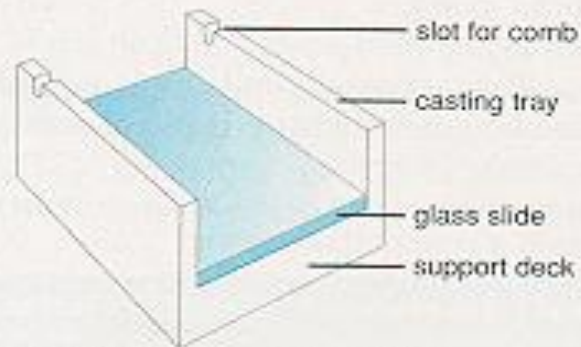
Now, most manufacturers prepare **special grades** of agarose that are screened for the presence of inhibitors & nucleases & for minimal background fluorescence after staining with ethidium bromide



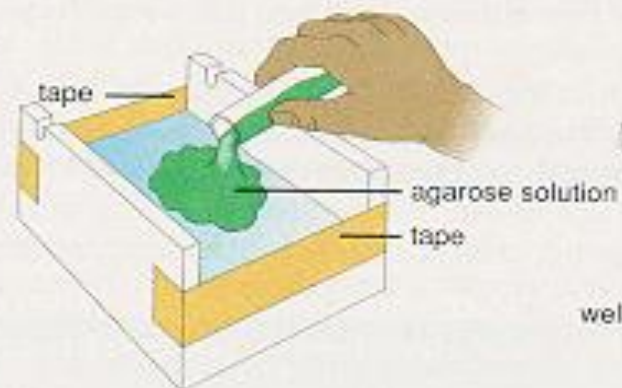
Agarose gels are cast by melting the agarose in the presence of the desired buffer until a clear, transparent solution is achieved

The melted solution is then poured into a mold & allow to harden. The agarose forms matrix & when an electric field is applied across the gel, DNA which is negatively charged at neutral PH, migrates toward the **anode**

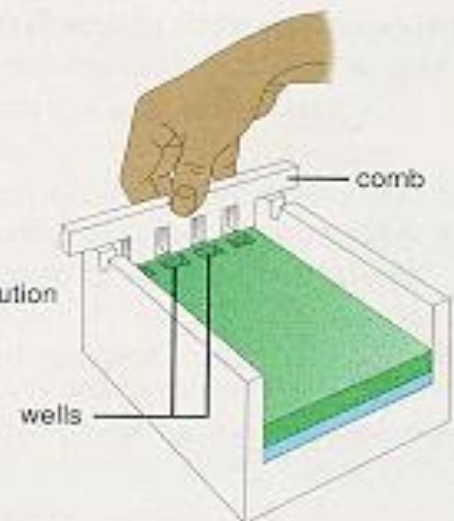




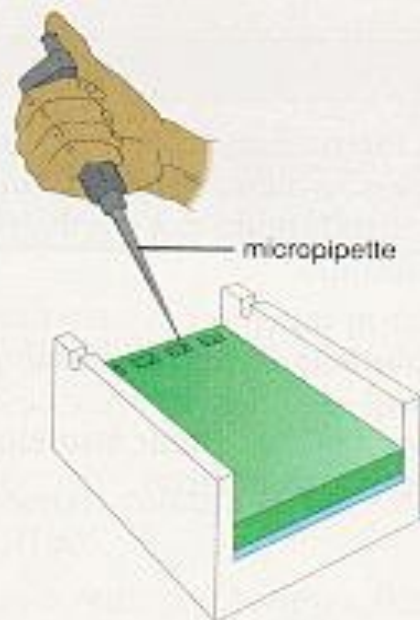
a. Casting tray for making gel slab



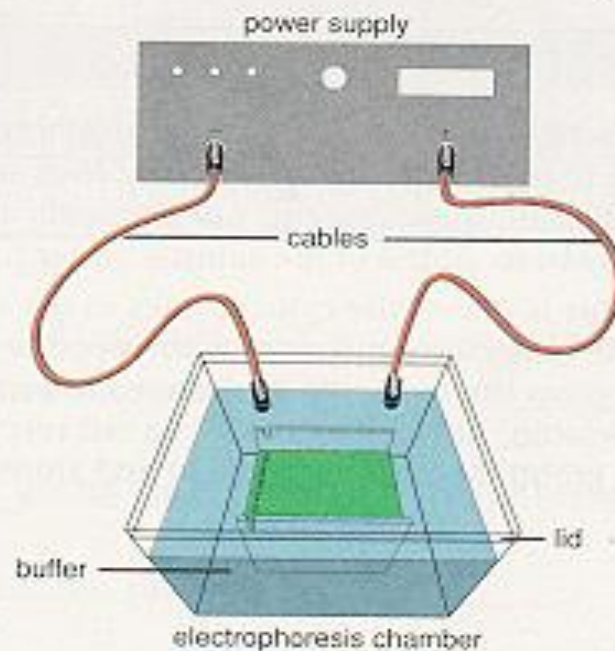
b. Agarose solution poured into casting tray



c. Comb that forms wells for samples



d. Wells that can be loaded with samples



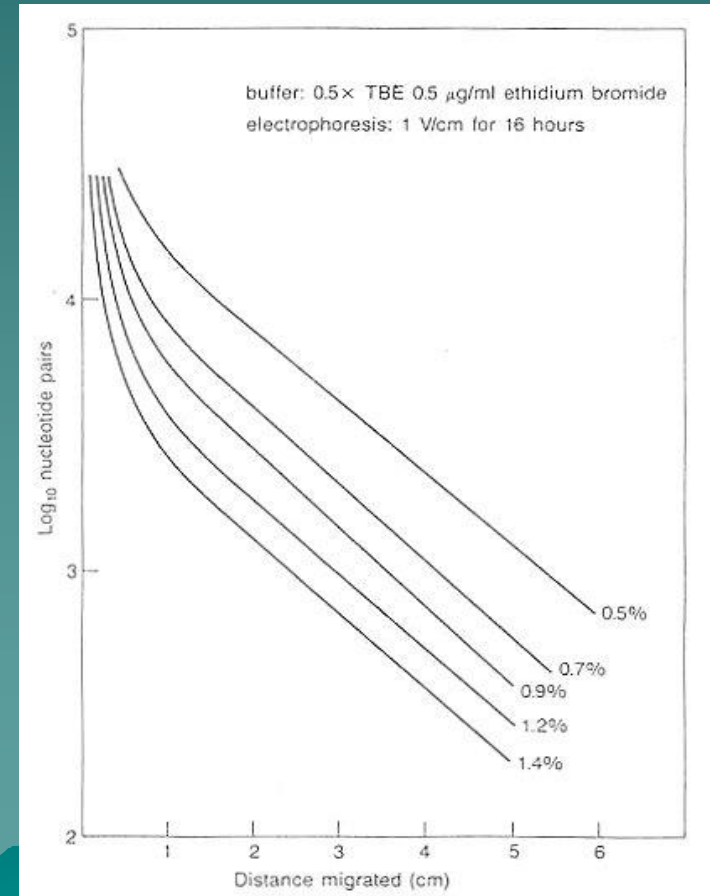
e. Electrophoresis chamber and power supply

Factors Affecting the Rate of DNA Migration in Agarose Gels

1) Molecular size of the DNA:

Larger molecules migrate more slowly than smaller molecules

They worm their way through the pores of the gel less efficiently than smaller molecules



2) Agarose Concentration:

By using gels of different concentrations, it is possible to resolve a wide size of DNA molecules

Range of separation in gels containing different amounts of agarose

Amount of agarose in gel (%[w/v])	Efficient range of separation of linear DNA molecules (kb)
0.3	5-60
0.6	1-20
0.7	0.8-10
0.9	0.5-7
1.2	0.4-6
1.5	0.2-3
2.0	0.2-3

3) DNA Conformation:

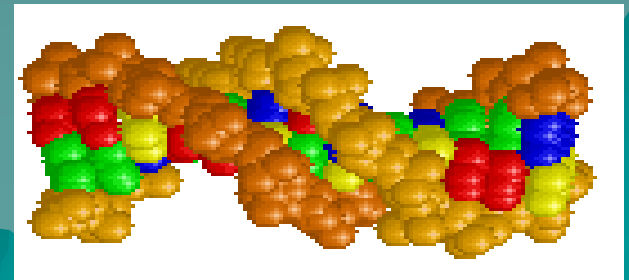
Superhelical circular DNAs

Niked circular DNAs

Linear DNAs

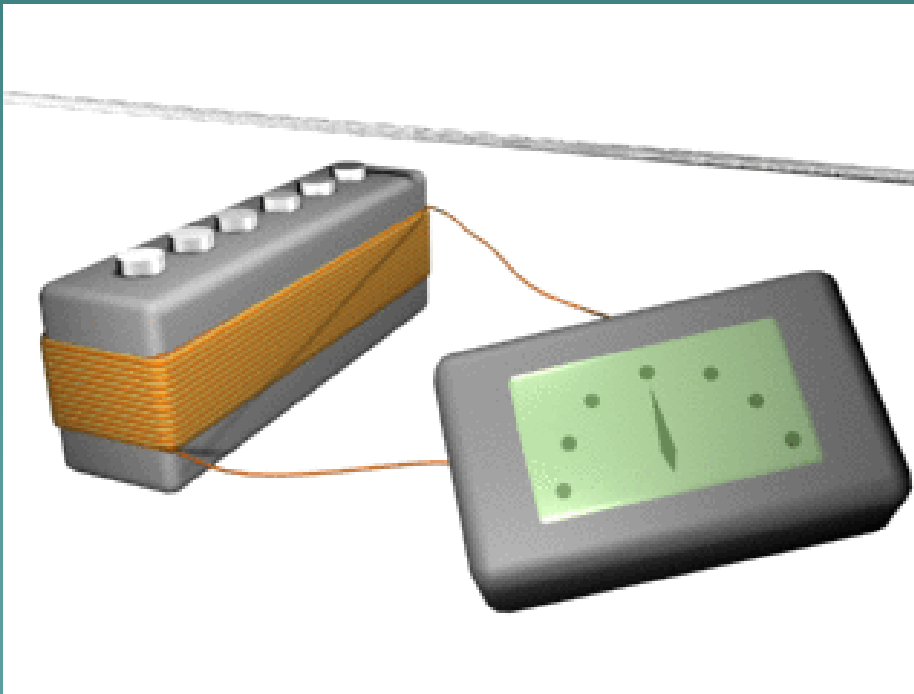
Of the same
molecular weight

Migrate through agarose gels at
different rates



4) Applied Voltage:

- The effective range of separation in agarose gels ↓ as the voltage ↑
- To obtain maximum resolution of DNA fragments < 2 kb size, agarose should be run at no more than 5 – 8 V/cm

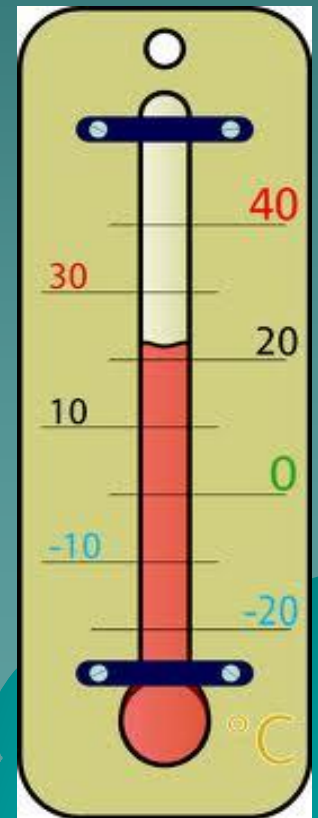


5) Direction of the electric field:

DNA molecules larger than 50-100 kb in length migrate through agarose at the same rate if the direction of the electric field remains constant

6) Base composition & Temperature:

In general, agarose gels are run at room temperature



7) Presence of Intercalating Dyes:

✍ The central dye in agarose gel electrophoresis is *ethidium bromide*

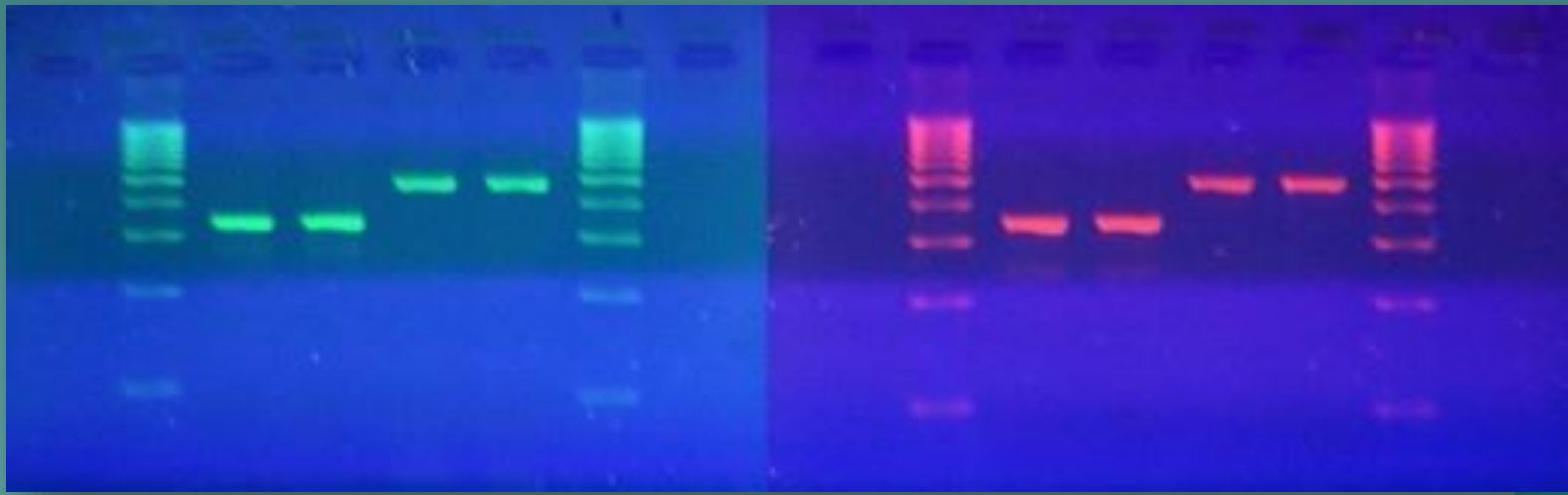
✍ It has the unique property of fluorescing under UV light when intercalated with DNA

✍ By running DNA through an EtBr-treated gel & exposing it to UV light, distinct bands of DNA become visible

Ethidium Bromide is a carcinogen & should be handled with care

Other dyes are sometimes used including **SYBER** green or **SYBER** safe. SYBER dyes are thought to be less carcinogenic than EtBr & to give cleaner, higher powered staining

Midori Green DNA Stain



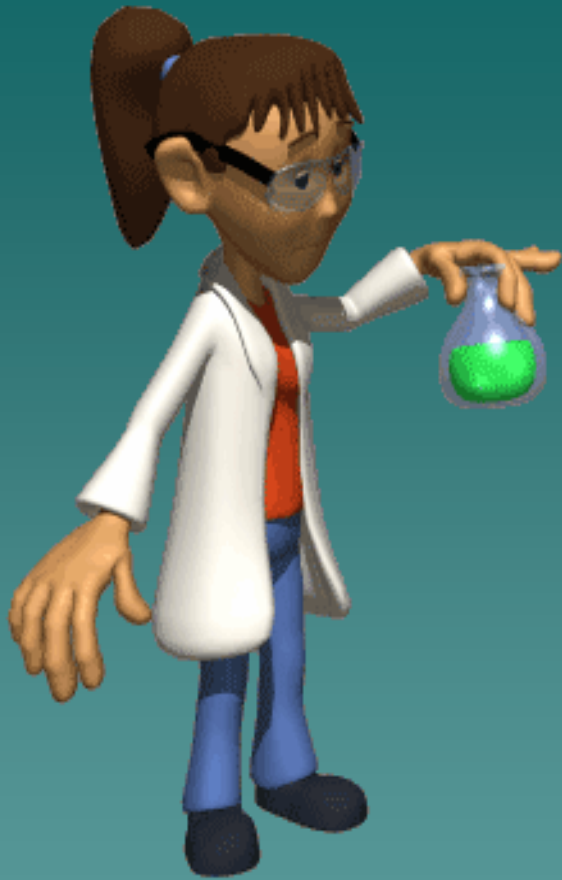
Left side Gel stained with Midori green stain & Right side Gel Stained with Ethidium Bromide

8) Electrophoresis Buffer:

The electrophoretic mobility of DNA is affected by the composition & ionic strength of the electrophoresis buffer

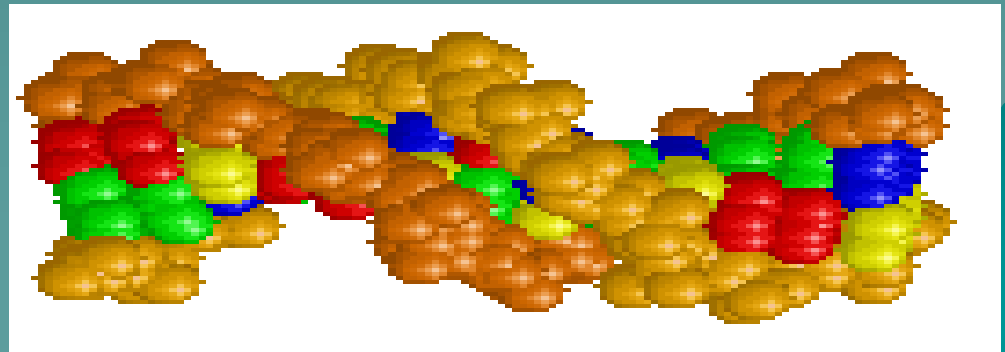
In the absence of ions → electrical conductance is minimal & DNA migrates very slowly

In buffers of high ionic strength → electrical conductance is very efficient & significant amount of heat are generated



Several different buffers are available for electrophoresis (**TAE, TPE & TBE**)

Electrophoresis buffers are usually made up as concentrated solutions & stored at room temp



Commonly used Electrophoresis Buffers

Buffer	Working solution	Conc. Stock solution (Per Liter)
Tris-acetate (TAE)	<u>1X</u> : 0.04 M Tris-acetate 0.001 M EDTA	<u>50X</u> : 243 g Tris base 57.1 ml glacial acetic acid 100 ml 0.5M EDTA (PH 8.0)
Tris-phosphate (TPE)	<u>1X</u> : 0.09 M Tris-phosphate 0.002 M EDTA	<u>10X</u> : 108 g Tris base 15.5 ml 85% phosphoric acid (1.679g/ml) 40 ml 0.5M EDTA (PH 8.0)
Tris-borate (TBE)	<u>0.5X</u> : 0.045 M Tris-borate 0.001 M EDTA	<u>5X</u> : 54 g Tris base 27.5 g boric acid 20 ml 0.5M EDTA (PH 8.0)
Alkaline	<u>1X</u> : 50 mM NaOH 1 mM EDTA	<u>1X</u> : 5 ml 10 N NaOH 2 ml 0.5M EDTA (PH 8.0)

9) Gel Loading Buffers:

- Gel- loading buffers are mixed with samples before loading into the slots of the gel

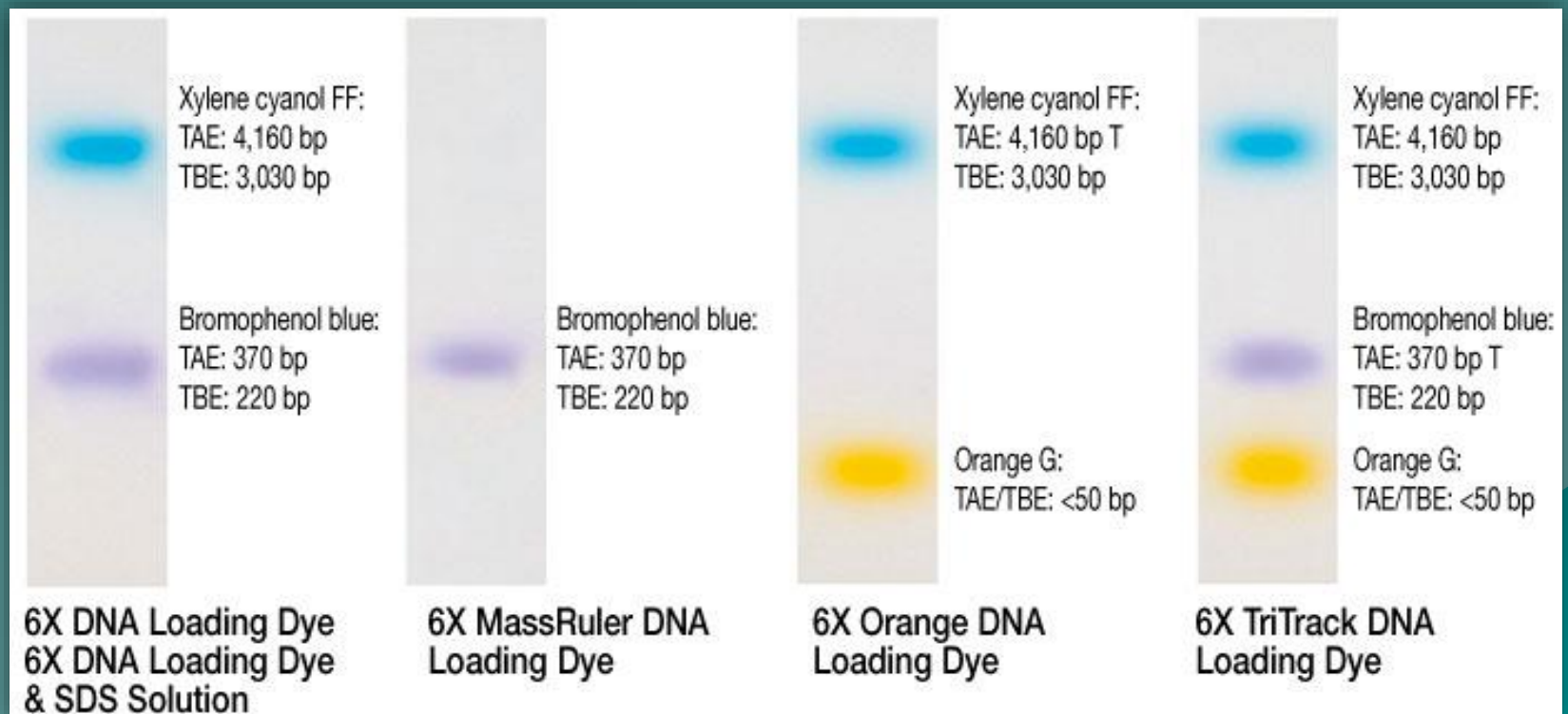
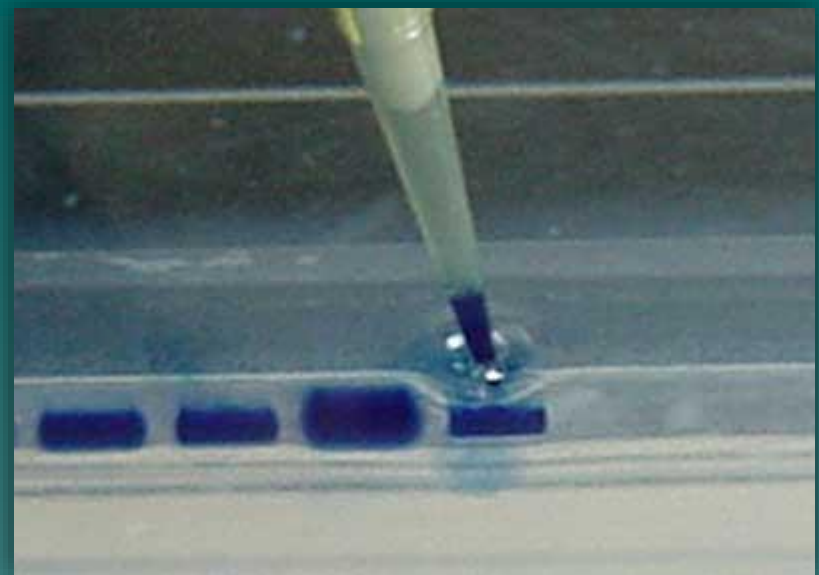
The purposes of that buffer are:

➡ ↑ The density of sample

➡ Ensuring that DNA sinks evenly into the well

➡ Add color to the sample

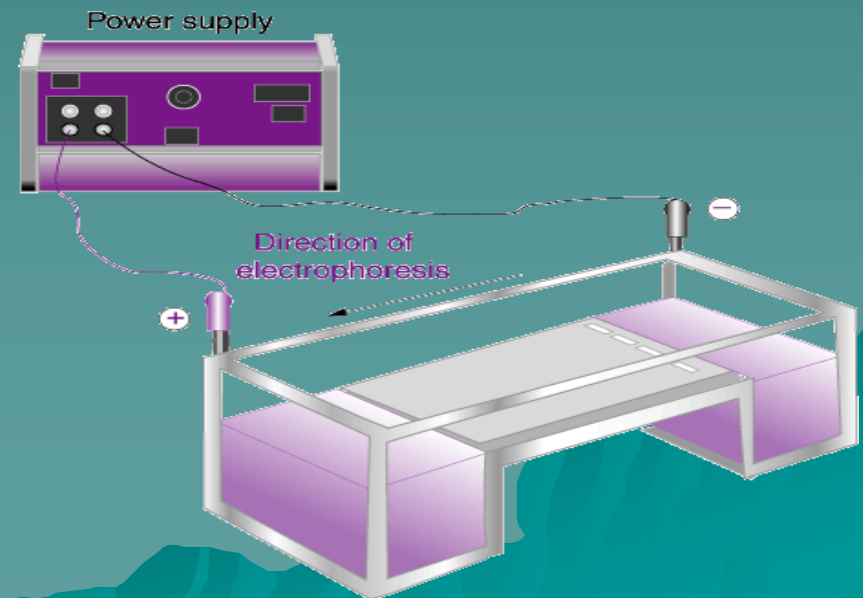
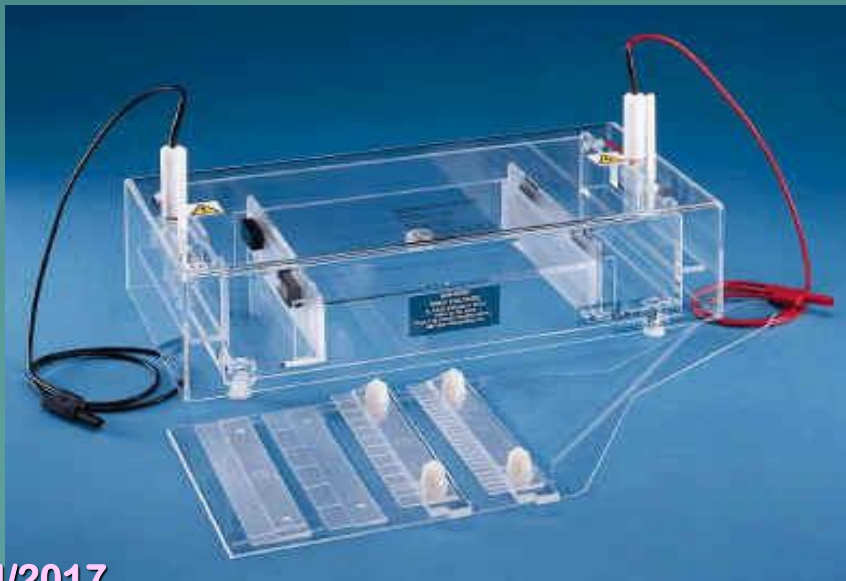
➡ The dye is negatively charged & move in the same direction as the DNA



Apparatuses Used For Agarose Gel Electrophoresis

The most commonly used configuration is the horizontal slab gel

Horizontal slab gels are usually poured on a glass plate or plastic tray that can be installed on a platform in electrophoresis tank

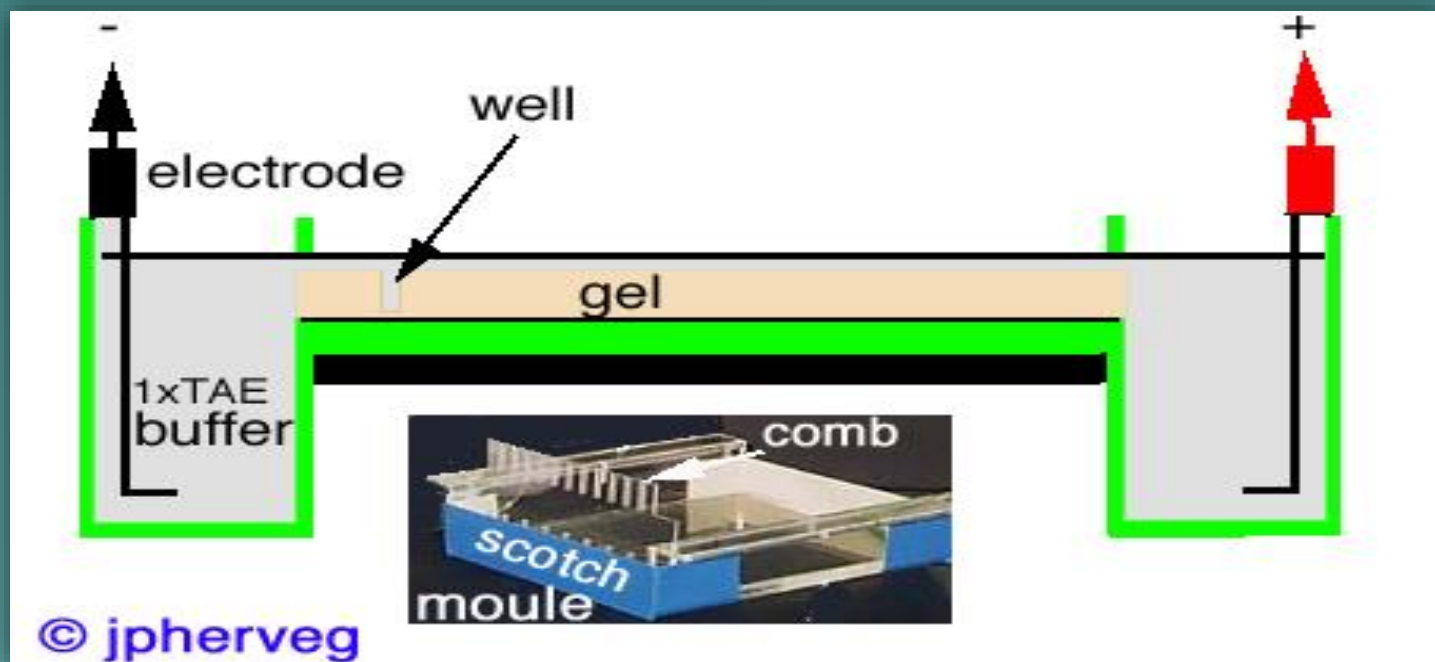


Preparation & Examination of Agarose gels

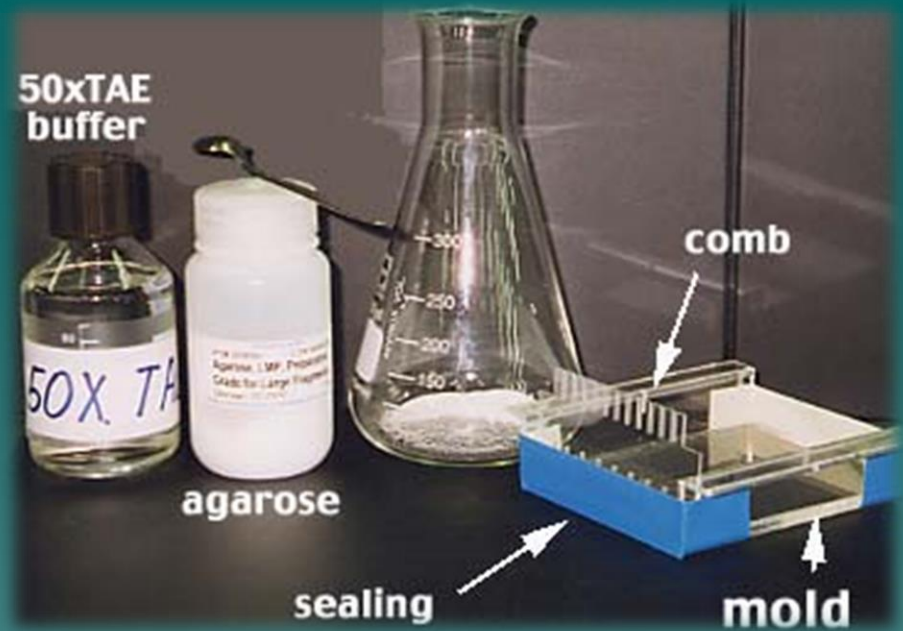
1. Seal the edges of a clean, dry, glass plate (plastic) with tape so as to form a mold. Set the mold on a horizontal section of bench

2. Prepare the materials you will need





3. Prepare mixture



4. Gel the mixture

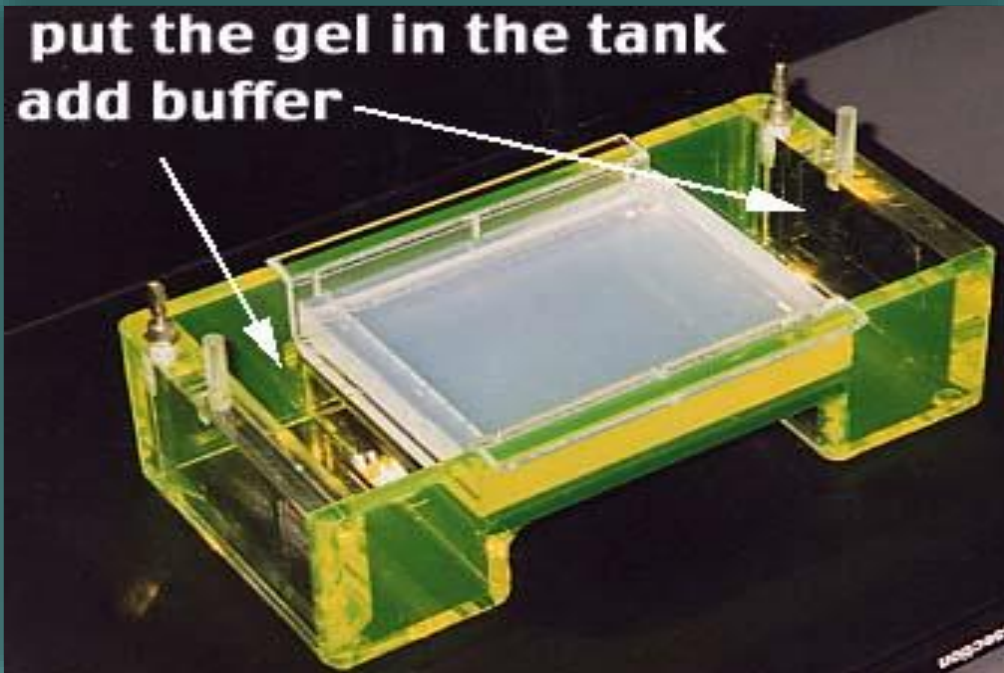
5. Cast gel (add ethidium bromide to the cooled gel)

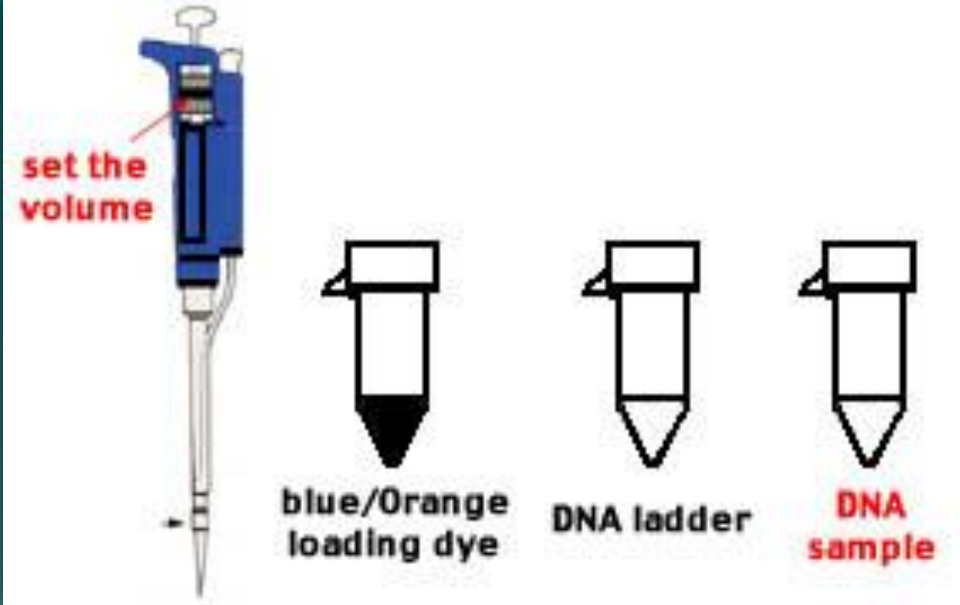


6. Place mold in the tank

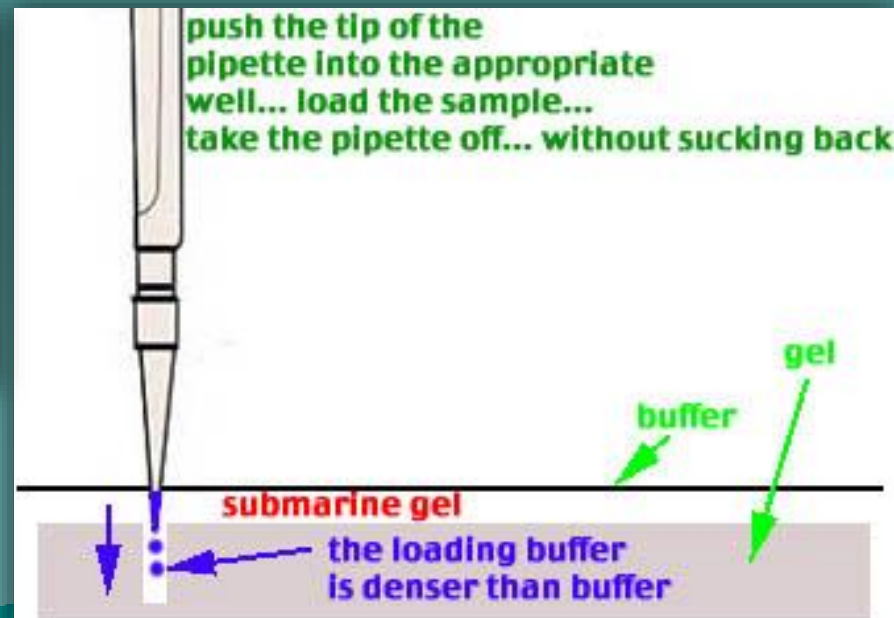
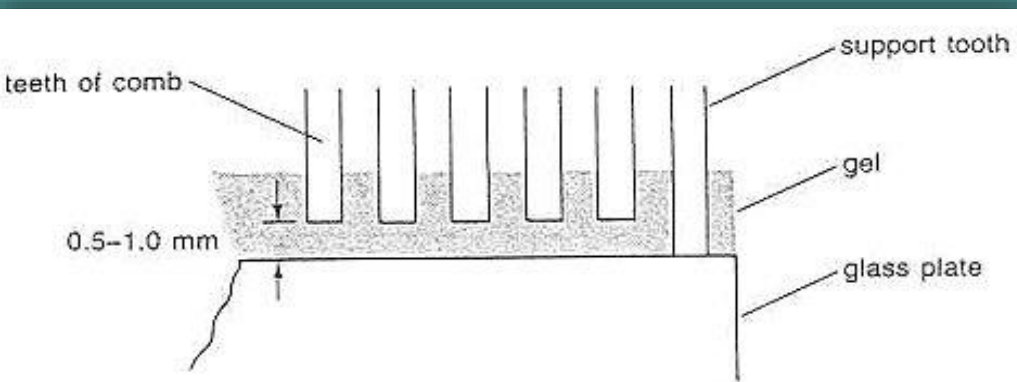


put the gel in the tank
add buffer

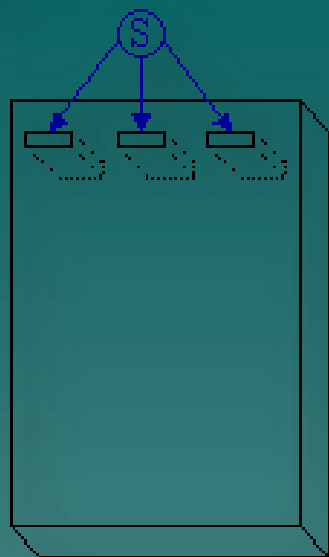




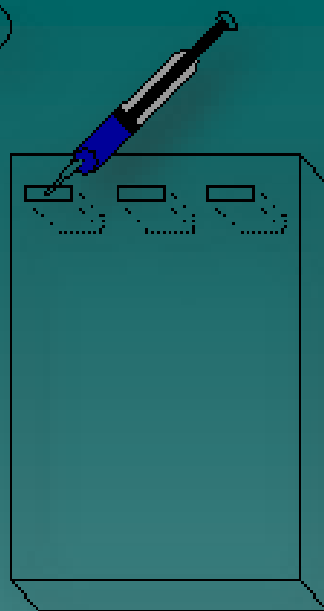
7. Charge the wells



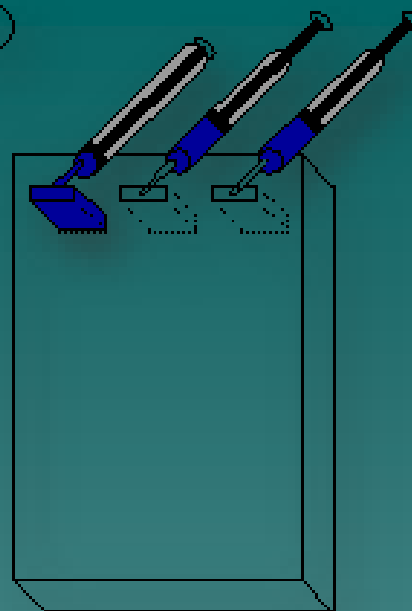
①



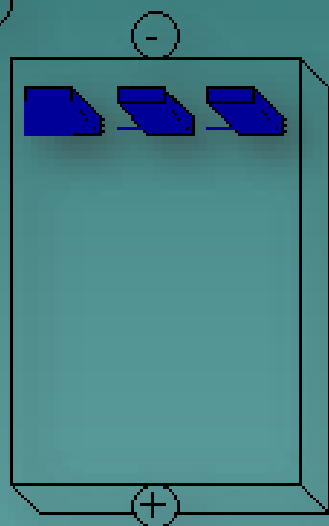
②



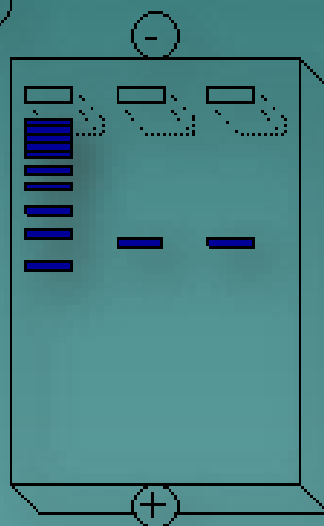
③



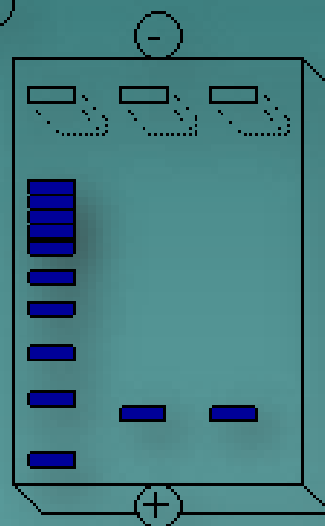
④

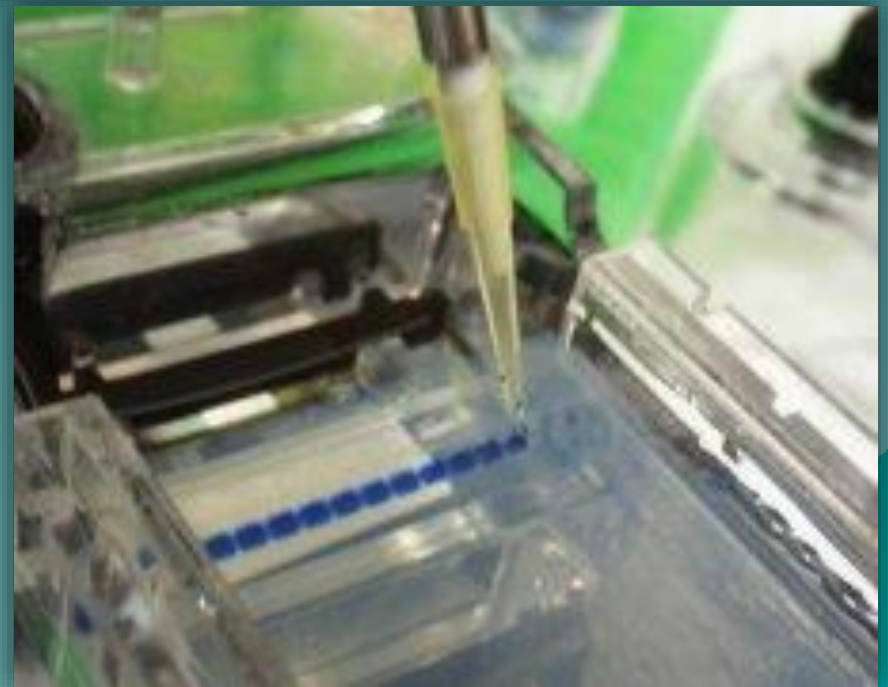


⑤

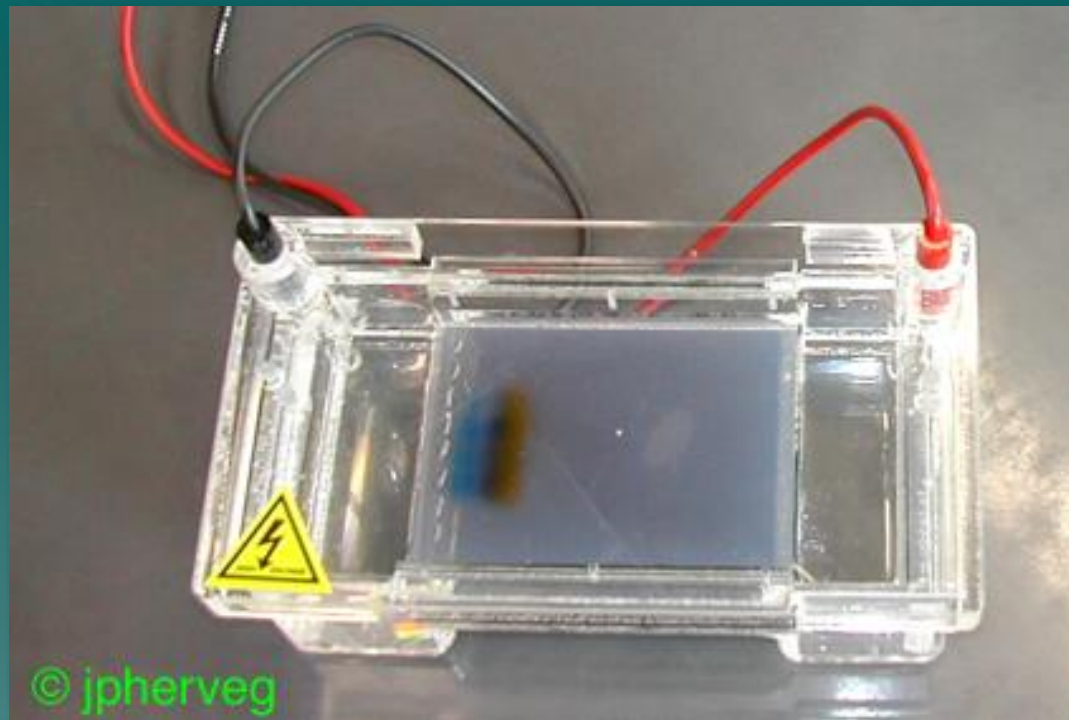


⑥

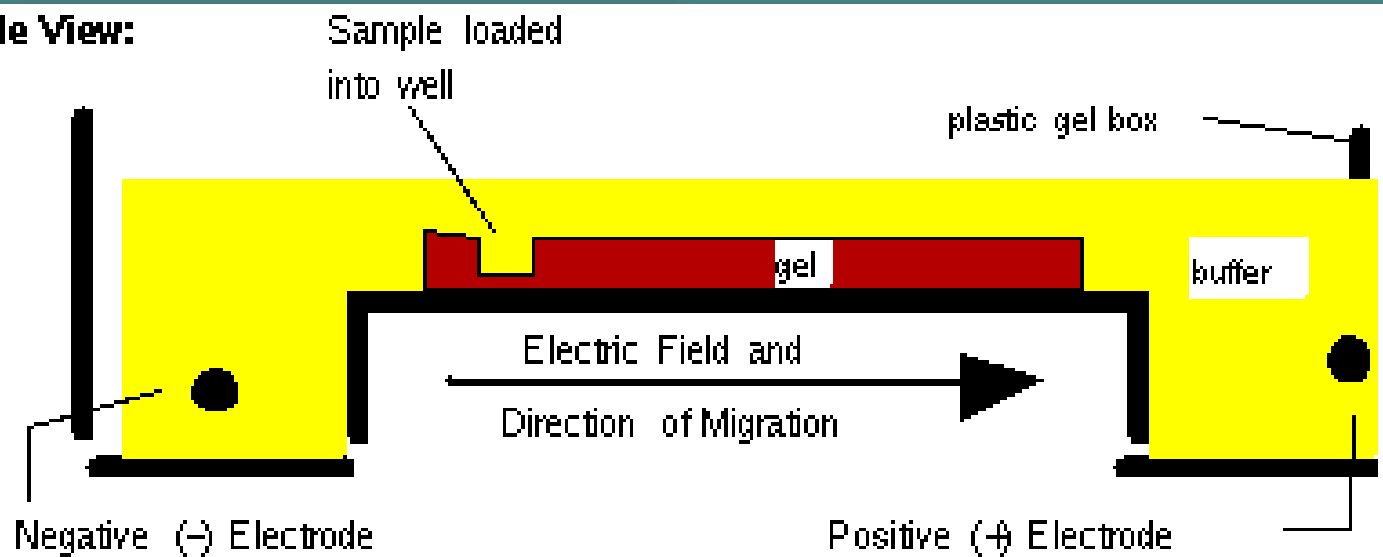




8. Plug in



Side View:





© jpherveg



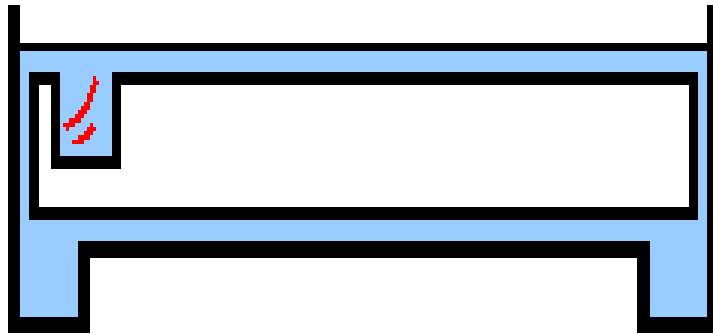
3/14/2017

Electrode

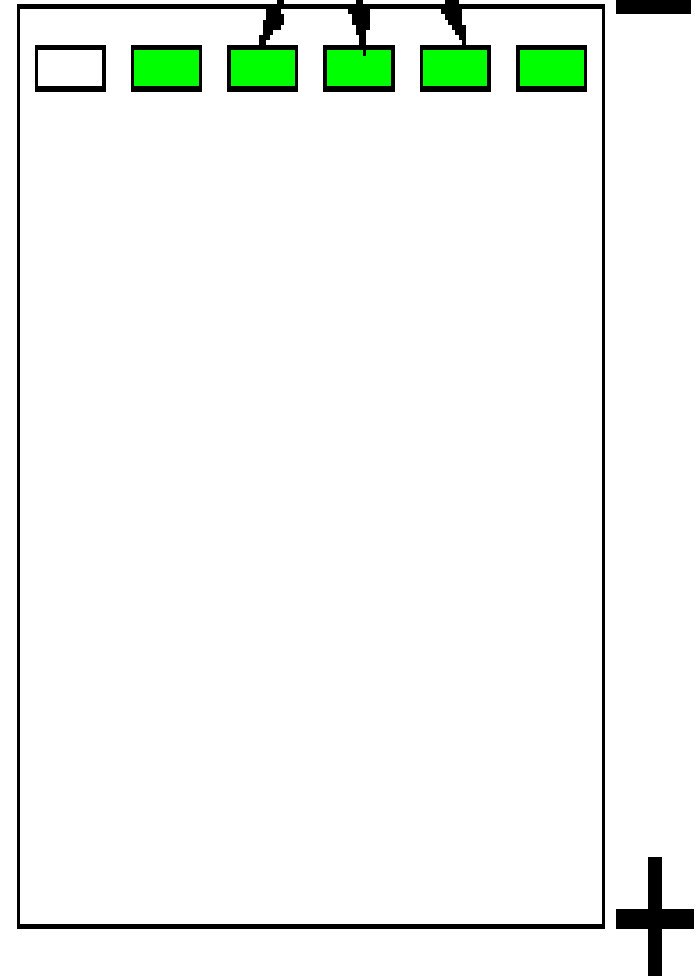
—

Electrode

+



START WELLS FILLED
WITH SAMPLE



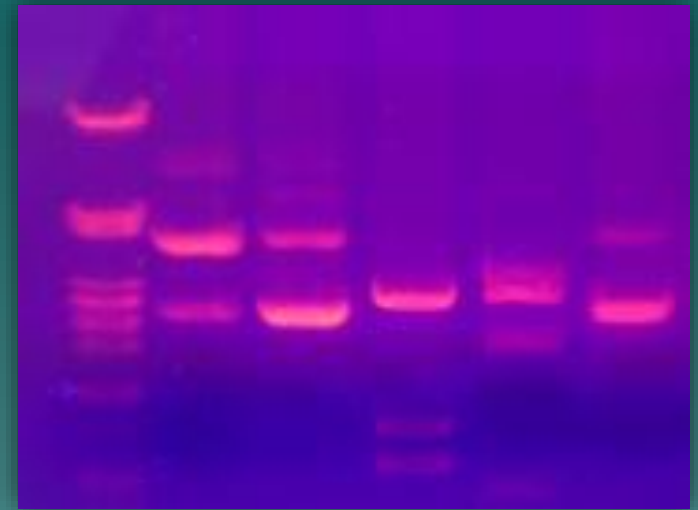
welder masks equipped
with glass windows

camera

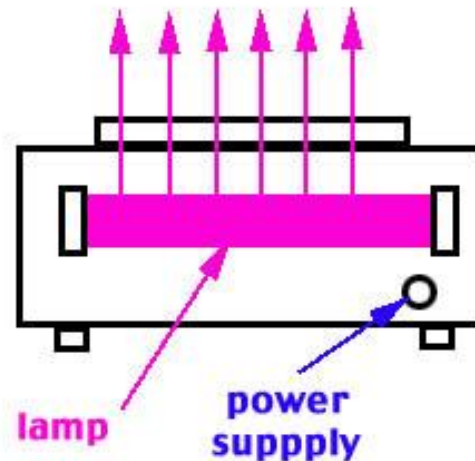
UV transilluminator

on/off switch

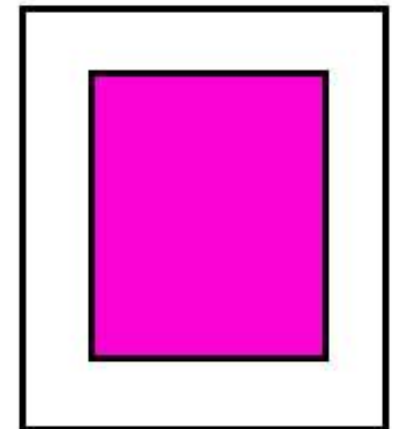
9. Examine gel



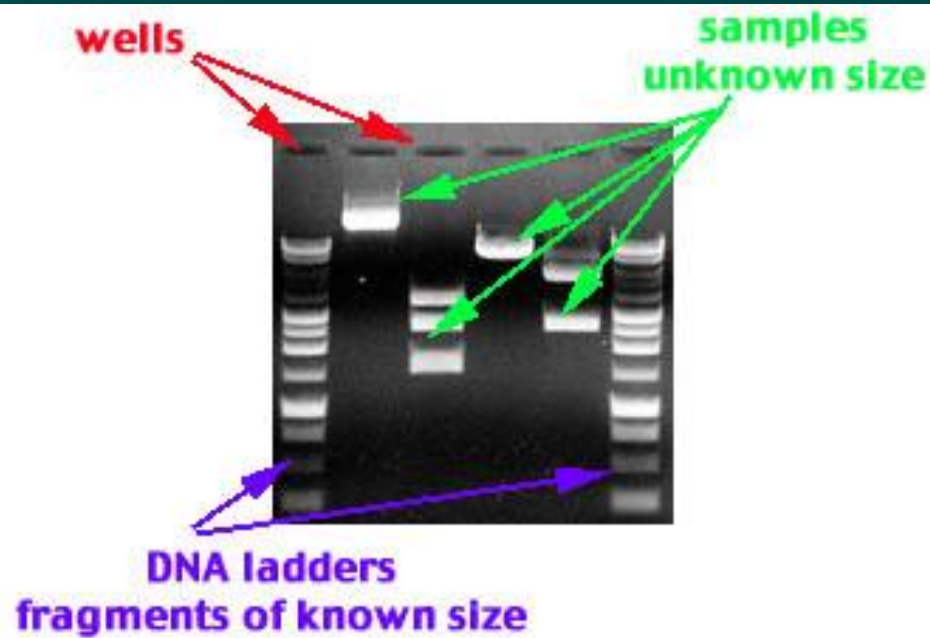
UV transilluminator



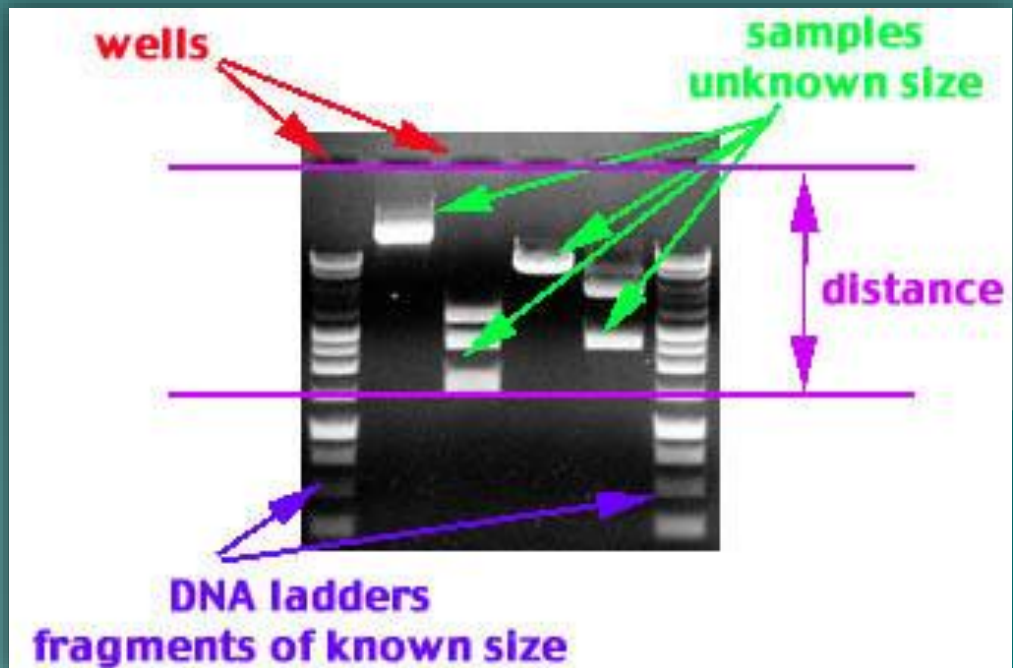
front view



top view



9. Results

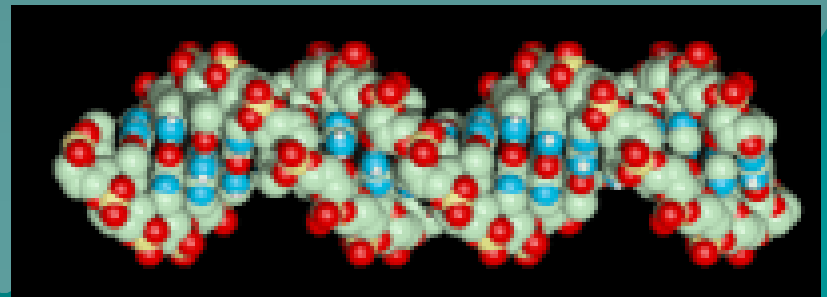


Decontamination of Ethidium Bromide Solutions

Ethidium bromide is a powerful mutagen & is moderately toxic

For this reason, gloves should be worn when working with solutions contain this dye

Also, after use, this solution should be decontaminated



A) Decontamination of concentrated solutions of ethidium bromide:

Method:

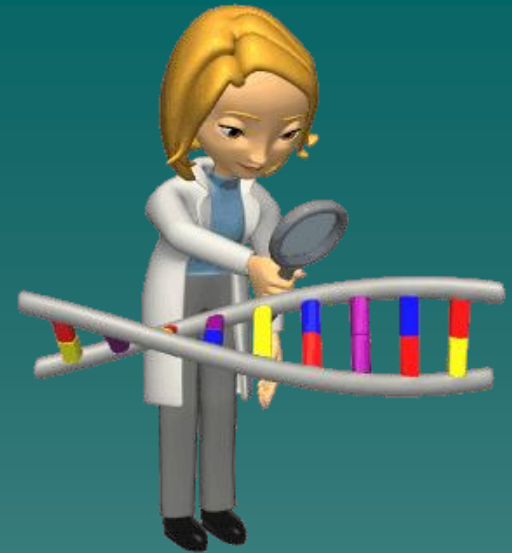
- a) Add **sufficient water** to reduce the concentration of ethidium bromide to < 0.5 mg/ml
- b) Then add 0.2 volume of fresh 5% hypophosphorous acid & 0.12 volume of fresh 0.5 M sodium nitrite. Mix carefully (The pH of solution is < 3.0)
- c) After incubation for 24 hours at room temperature, add a large excess of **1 M sodium bicarbonate**. The solution may now be discarded

B) Decontamination of dilute solution of ethidium bromide:

Method:

- a)** Add 100 mg of powdered activated charcoal for each 100 ml of solution
- b)** Store the solution for 1 hour at room temperature, shaking it intermittently
- c)** Filter the solution through a Whatman No.1 filter, & discard the filtrate
- d)** Seal the filter & activated charcoal in a plastic bag, & dispose of the bag in the hazardous waste

3/14/2017



***Thank you for your
attention!***



3/14/2017