

# Introduction to Gene Cloning



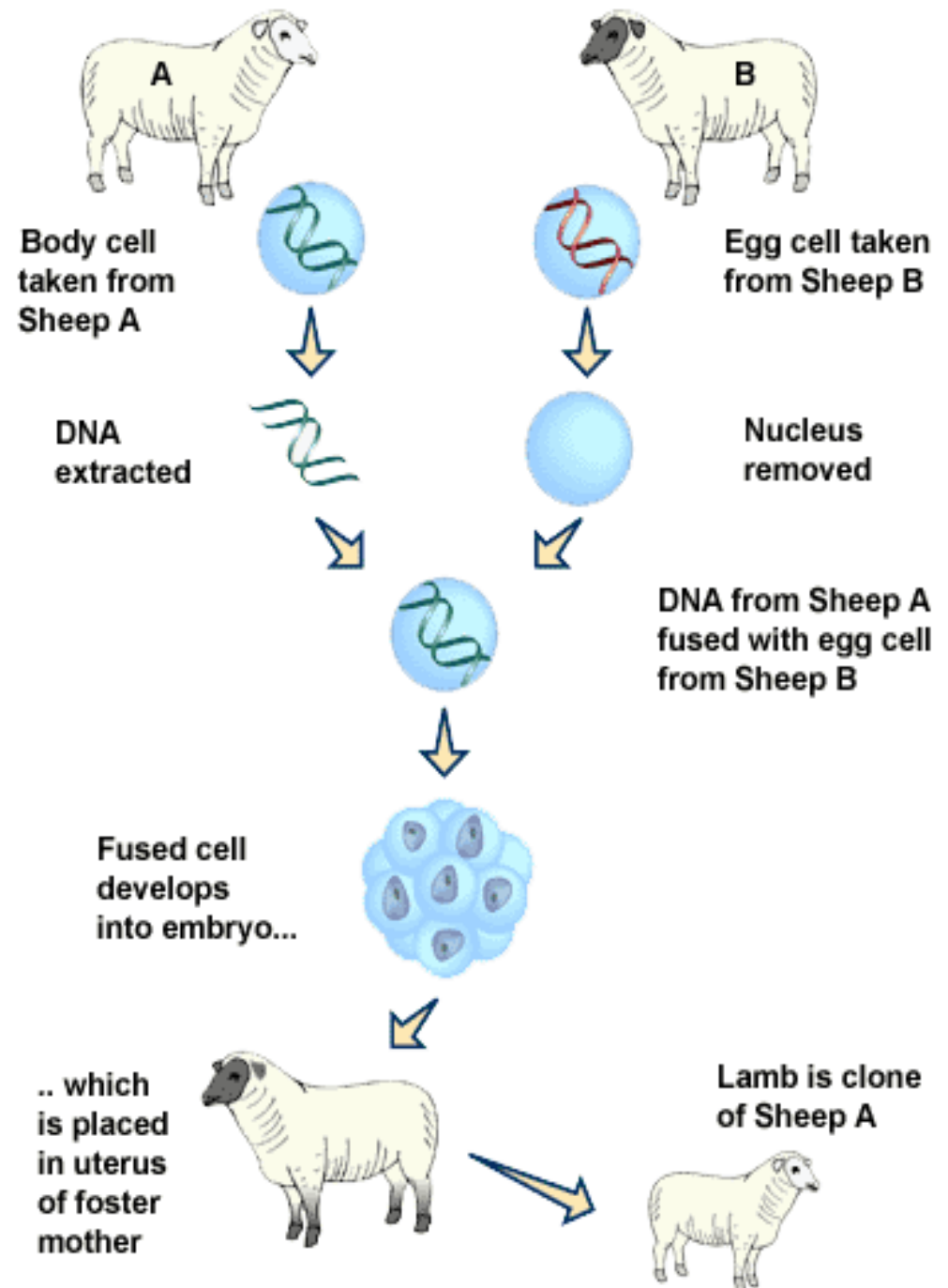
**Mohamed N. Seleem**

# Gene Cloning

- Making multiple copies of a target gene
- Generally use bacteria as the “factory”



# Cloning





Ethical  
Issue  
of the Day #1

# Reproductive Cloning

Arguments for a ban  
on human reproductive\* cloning

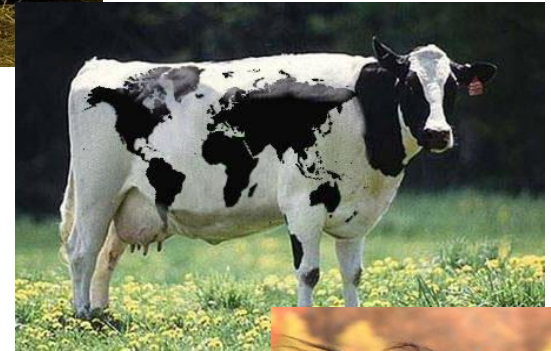
- hundreds of cloned embryos must be created and placed into a mother to produce a single live birth
- dying, stillborn and deformed babies frequent
- Even those who have lived a long time (such as Dolly) have a large number of serious health problems

\*cloning to make a baby



# Animal cloned so far

- Frog (1952)
- Sheep (1996)
- Mouse (1997)
- Cow (1998)
- Goat (2000)
- Cat (2001)
- Rabbit (2002)
- Horse, Rat (2003)



ANDi Monkey



April 2009 Injaz, or 'Achievement' Dubai



Snuppy the Afghan Hound

# Cloning a gene

- Insert a particular fragment of DNA (a particular gene) into a vector
- A vector is another DNA molecule that can be put into a host
- This creates a new DNA molecule  
--> Recombinant DNA
- Greatly aids further study of that gene, and its use in diagnosis or treatment



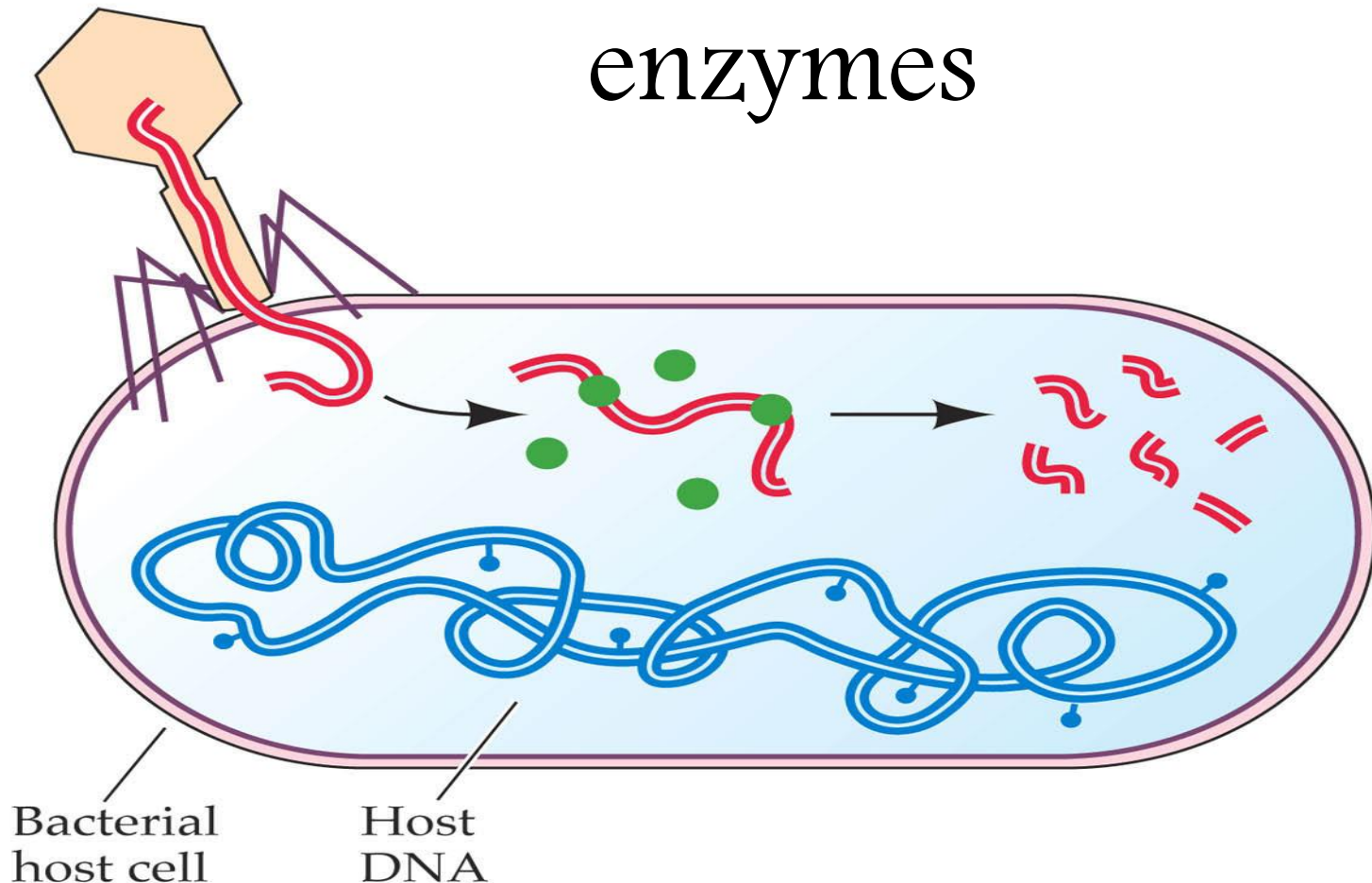
# Cloning Tools

- Restriction endonucleases
- Ligase
- Vectors
- Host
- Methods for introducing DNA into a host cell



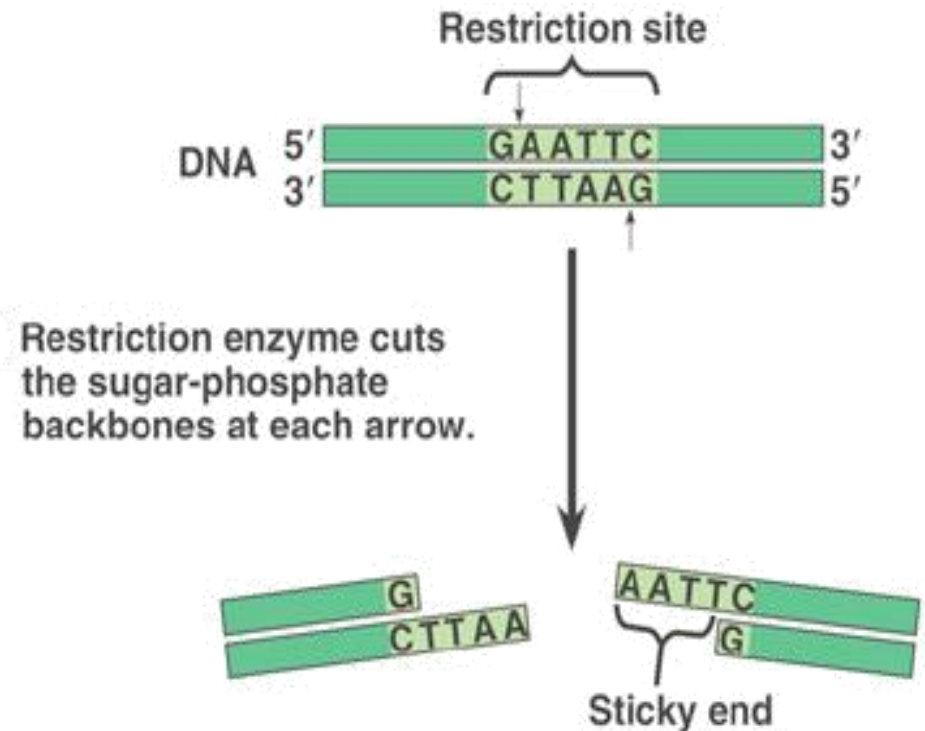


# Restriction endonucleases enzymes



# Cutting DNA

- Restriction endonucleases (restriction enzymes)
  - sticky ends
  - blunt ends
- Nomenclature
  - *EcoRI*
  - *E* = genus (*Escherichia*)
  - *co* = species (*coli*)
  - *R* = strain
  - *I* = # of enzyme



# Some restriction enzymes

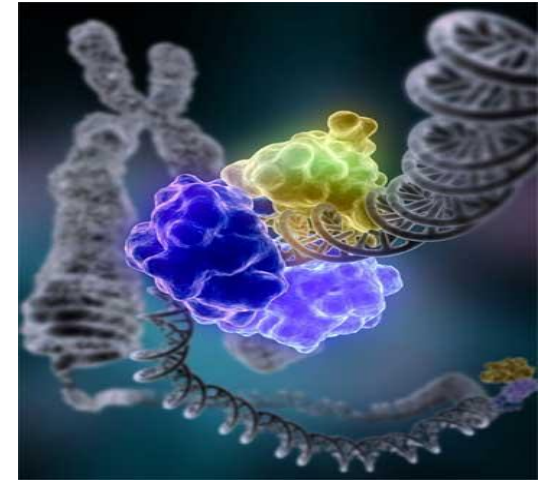
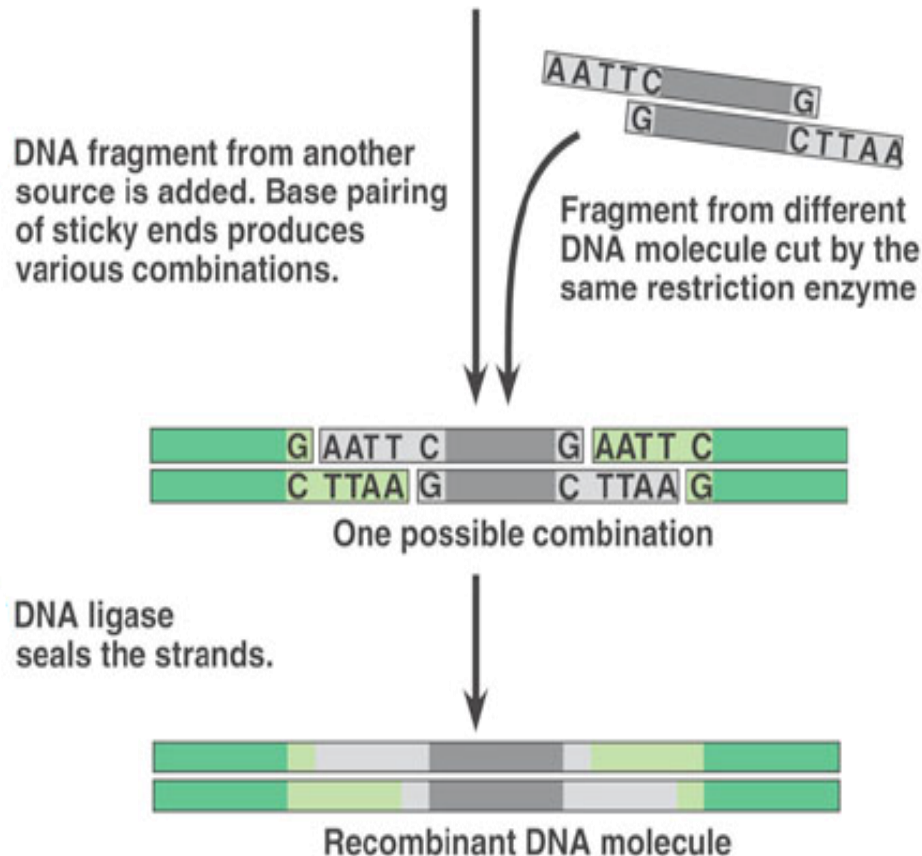
**Table 8.1** Properties of Some Restriction Enzymes

Enzyme	Bacterial Source	Restriction Site*
<i>Bam</i> HI	<i>Bacillus amyloliquefaciens</i> H	G <sup>↓</sup> GATCC CCTAG <sup>↑</sup> G
<i>Eco</i> RI	<i>Escherichia coli</i> RY13	G <sup>↓</sup> AATTC CTTAA <sup>↑</sup> G
<i>Eco</i> RII	<i>E. coli</i> R245	CC <sup>↓</sup> GG GG <sup>↑</sup> CC
<i>Hind</i> II	<i>Haemophilus influenzae</i> Rd	GTPy <sup>↓</sup> PuAC CAPu <sup>↑</sup> PyTG
<i>Hind</i> III	<i>H. influenzae</i> Rd	A <sup>↓</sup> AGCTT TTCGA <sup>↑</sup> A
<i>Hin</i> FI	<i>H. influenzae</i> Rf	G <sup>↓</sup> ANTC CTNA <sup>↑</sup> G
<i>Hpa</i> I	<i>H. parainfluenzae</i>	GTT <sup>↓</sup> AAC CAA <sup>↑</sup> TTG
<i>Msp</i> I	<i>Moraxella</i> sp.	CC <sup>↓</sup> GG GG <sup>↑</sup> CC
<i>Sma</i> I	<i>Serratia marcescens</i>	CCC <sup>↓</sup> GGG GGG <sup>↑</sup> CCC

\*Arrows indicate sites of cleavage; Py = pyrimidine (either T or C); Pu = purine (either A or G); N = any nucleotide (A, T, G, or C).

- 100's of restriction enzymes (RE's) are commercially available
- Artificial RE sites can be inserted at ends of any gene
- RE's and ligase allow precise cutting and pasting of any DNA sequences

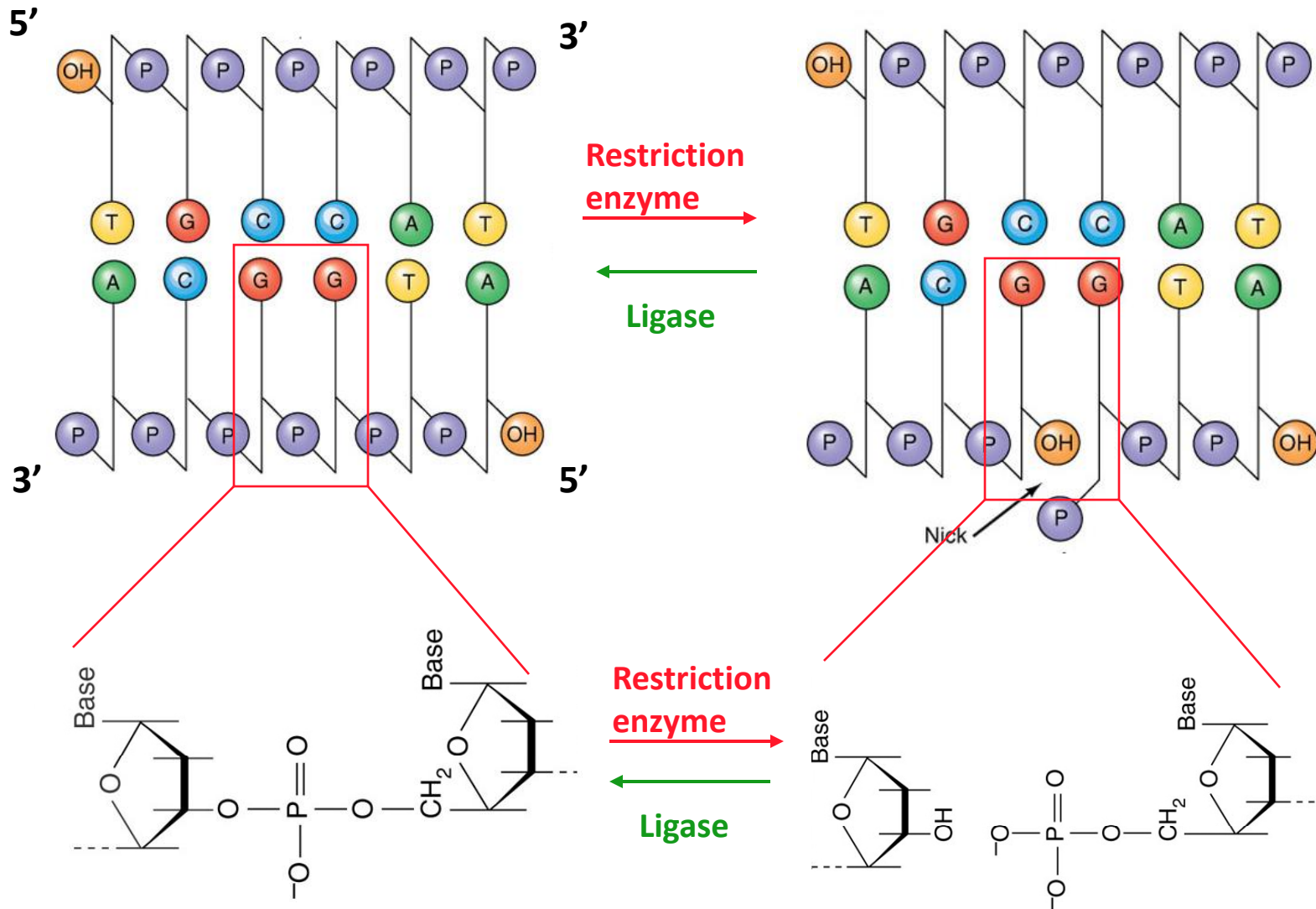
# Pasting DNA



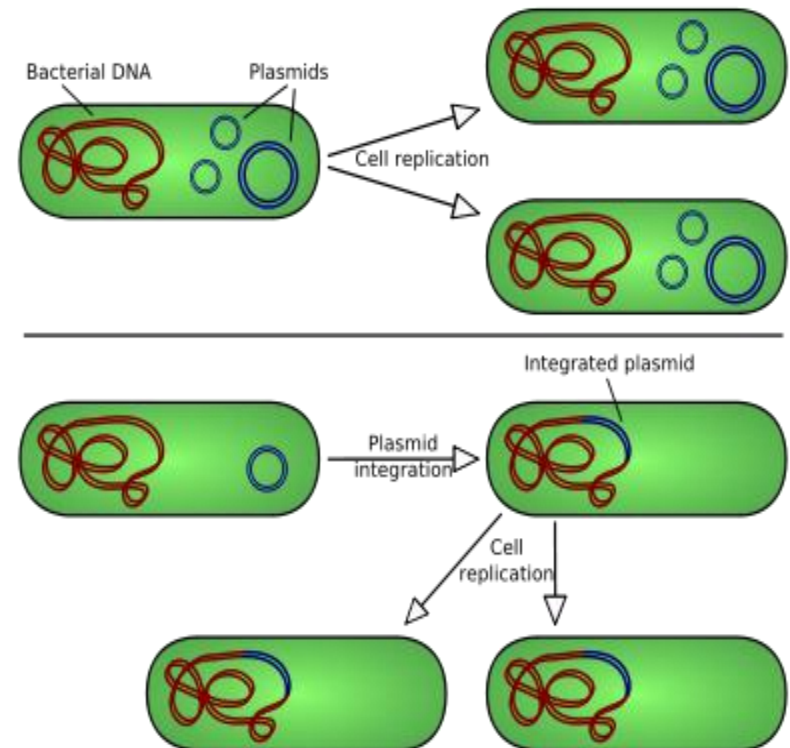
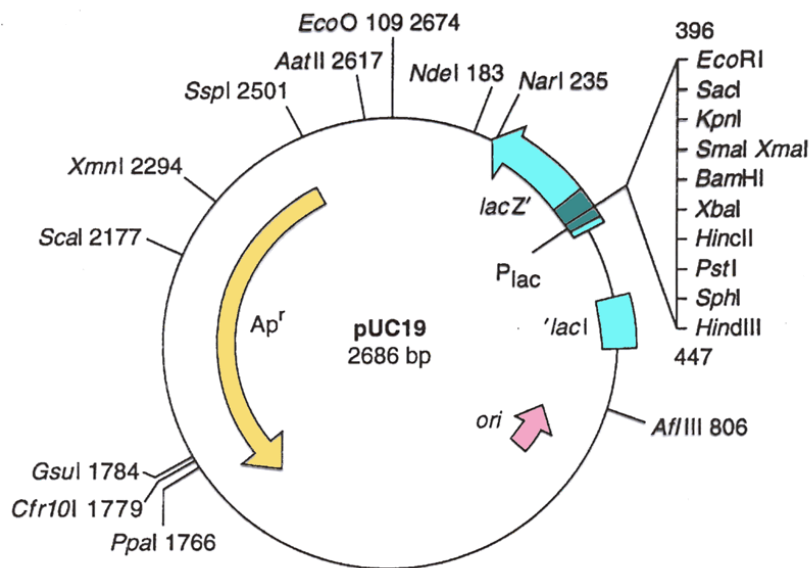
- Complementary ends (sticky ends) H-bond
- Ligase forms phosphodiester bond to seal strands together.



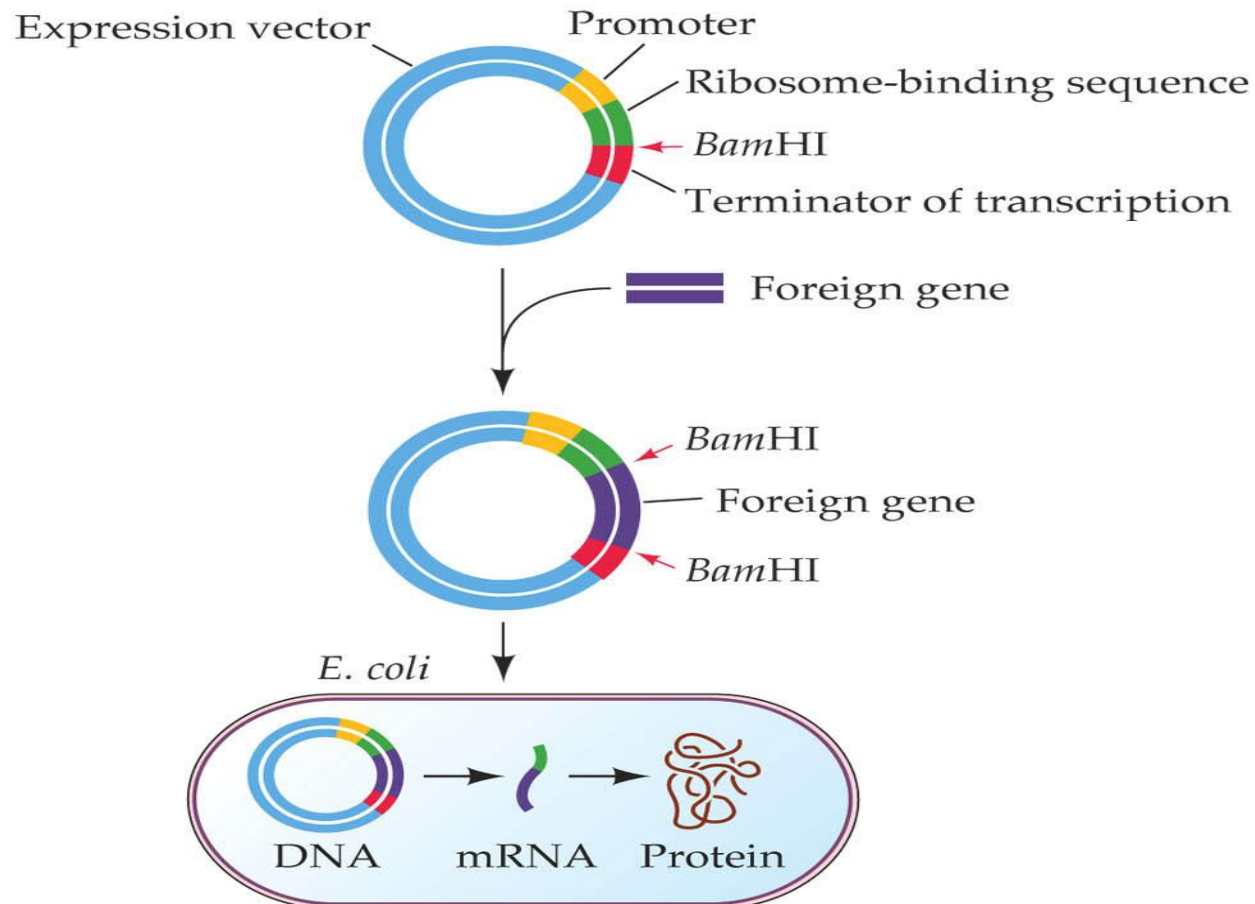
# DNA ligase covalently links two DNA strands



# Plasmid vectors



*An Expression Vector Allows a Foreign Gene to Be Expressed in a Host Cell*



# Cloning vectors

allowing the exogenous DNA to be inserted, stored, and manipulated mainly at DNA level.

1 Plasmid vectors

2 Bacteriophage vectors

3 Cosmids

4 BACs & YACs



# What determines the choice vector?

- insert size

- Ω vector size

- Ω restriction sites

- Ω copy number

- Ω cloning efficiency

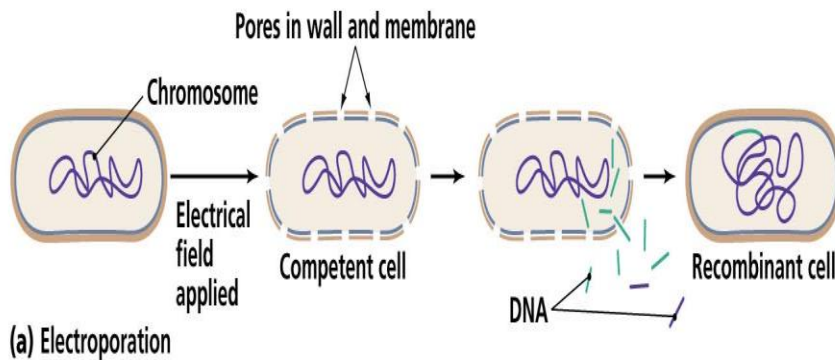
- Ω ability to screen for inserts

- Ω what down-stream experiments do you plan?

**Table 5.1** Maximum DNA insert possible with different cloning vectors. YACs are discussed on p. 159.

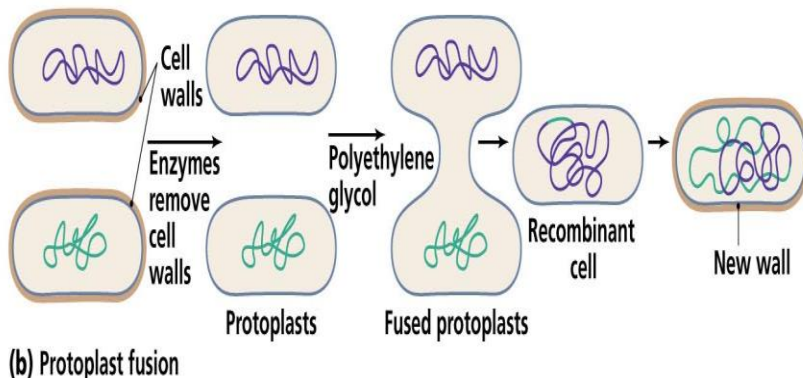
Vector	Host	Insert size
λ phage	<i>E. coli</i>	5–25 kb
λ cosmids	<i>E. coli</i>	35–45 kb
P1 phage	<i>E. coli</i>	70–100 kb
PACs	<i>E. coli</i>	100–300 kb
BACs	<i>E. coli</i>	≤ 300 kb
YACs	<i>Saccharomyces cerevisiae</i>	200–2000 kb

# Getting DNA into cells--Transformation of bacteria



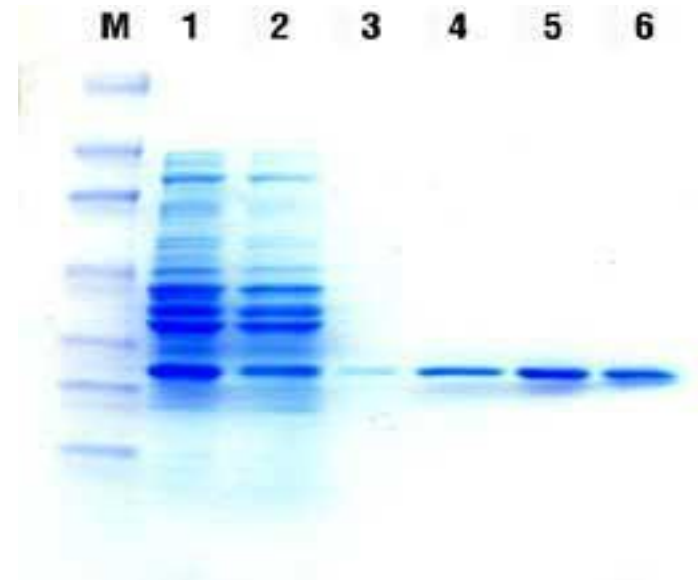
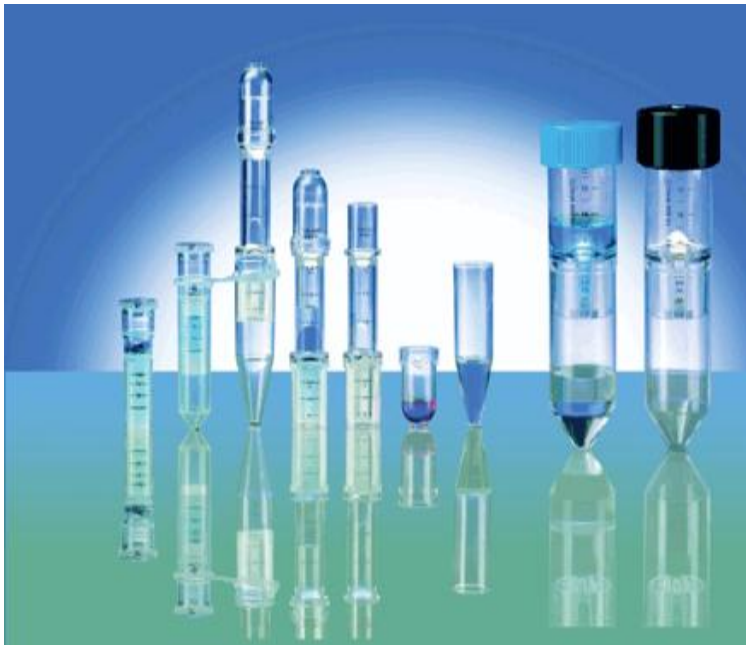
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- Cells are made competent via electroporation or via pre-treatment with  $\text{CaCl}_2$ /cold
- DNA (plasmid) is added and cells that have taken up the plasmid are identified by plating on selective media



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# Protein Purification



# Bacterial Expression System

## Shortcomings

- There are problems with expression of eukaryotic proteins in a bacterial system
  - Bacteria may recognize the proteins as foreign and destroy them
  - Posttranslational modifications are different in bacteria
  - Bacterial environment may not permit correct protein folding
- Very high levels of cloned eukaryotic proteins can be expressed in useless, insoluble form